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2025

..., 2025, 27.







**Н. .**

E-mail: *postnabu@mail.ru*

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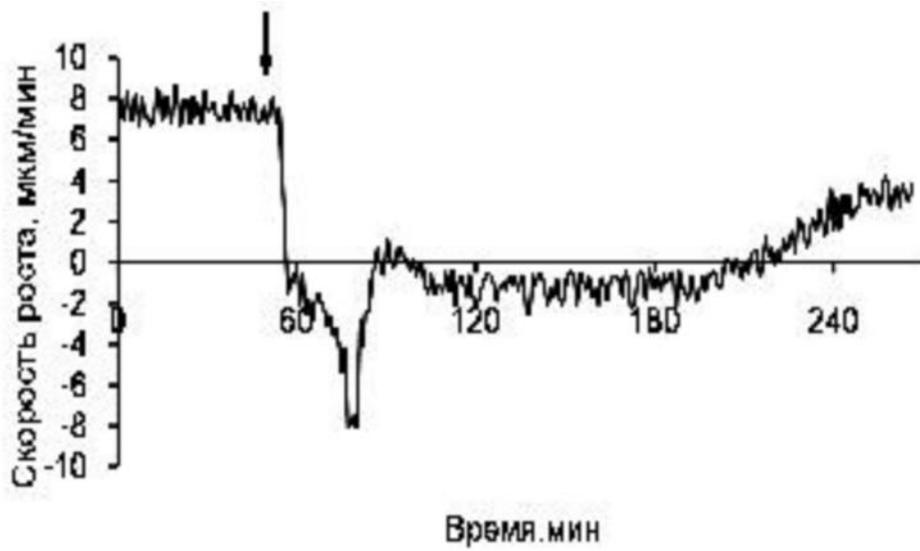
NaCl

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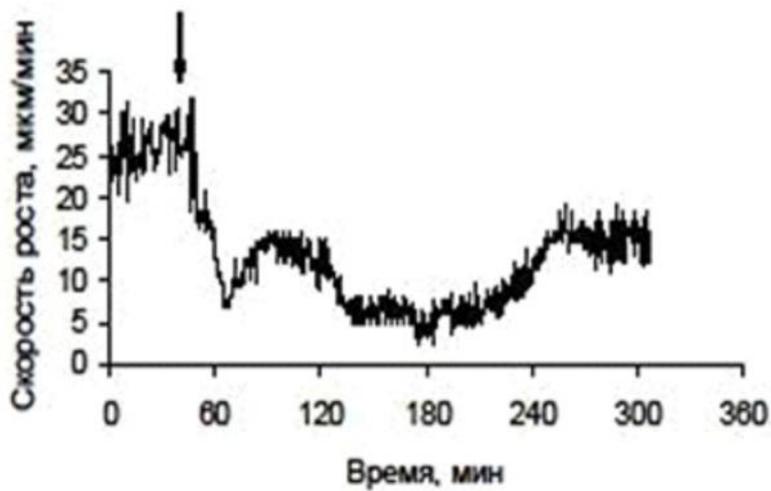
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[7].

[7].



.1. NaCl (80 ) 2- 21-  
 , ↓ NaCl ( ) .

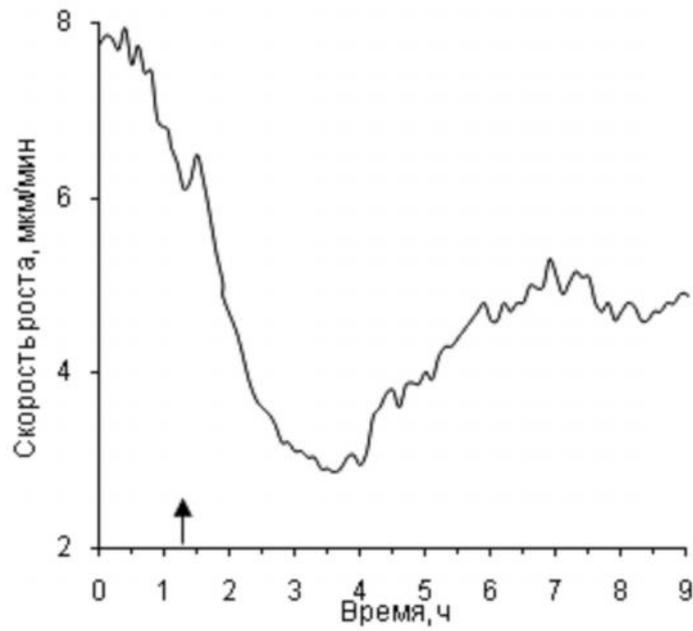


.2. NaCl (100 ) 2- 12-  
 , - ↓ NaCl ( ) .

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.3.

(1,5 )

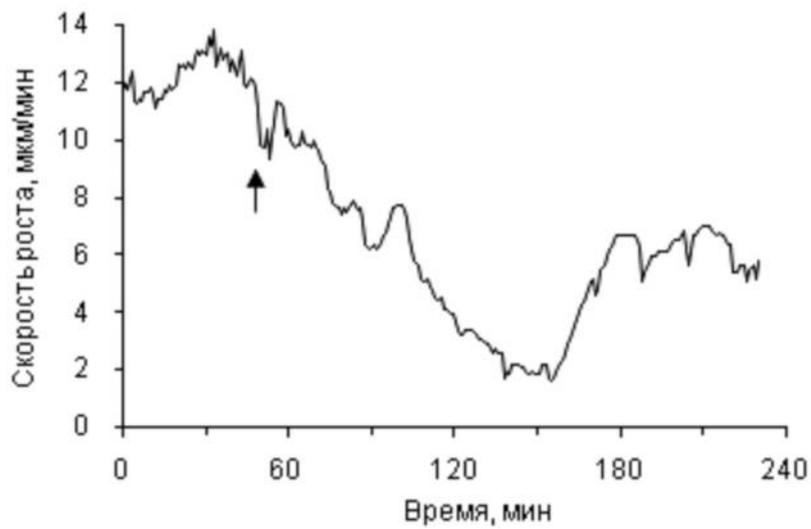
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( .4).



.4.

(6 )

12-

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( ) .

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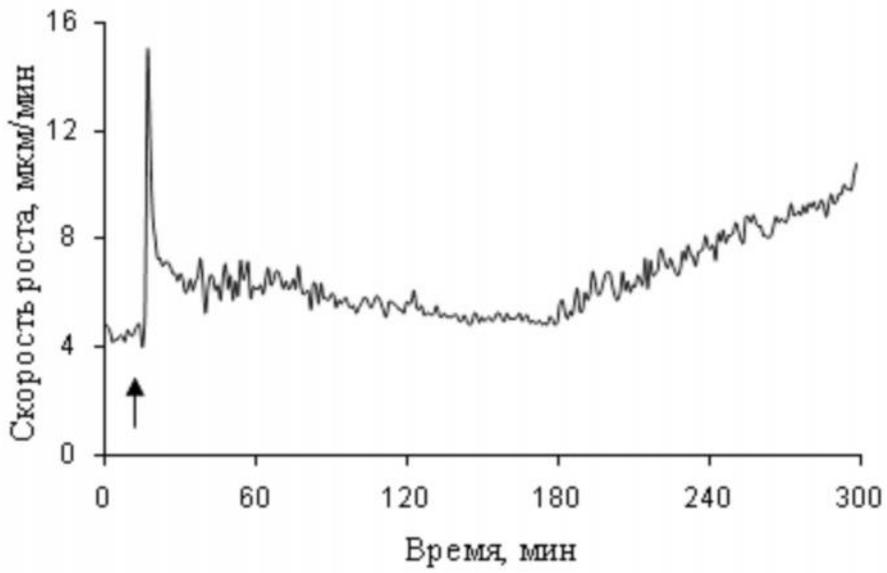
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[8].

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*de novo,*

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(0,01 )

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(NaCl,

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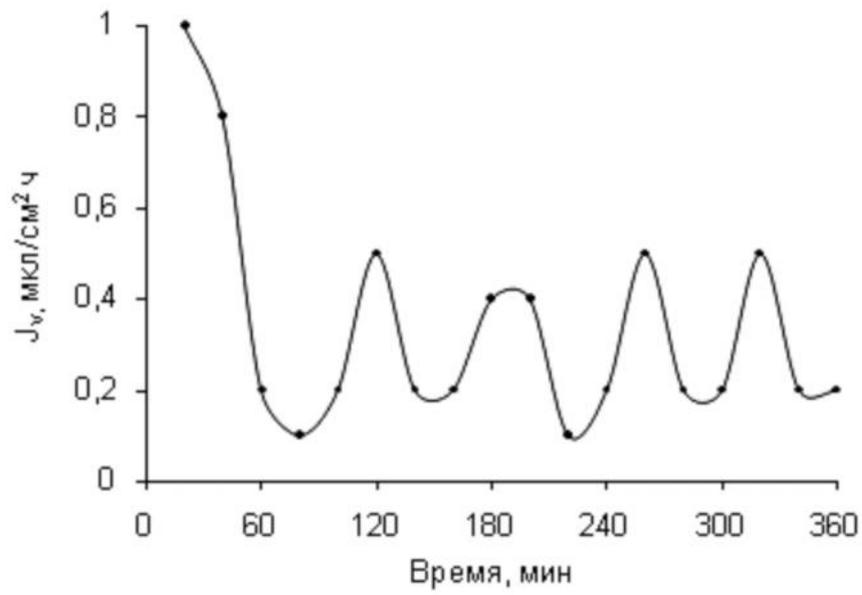
NaCl

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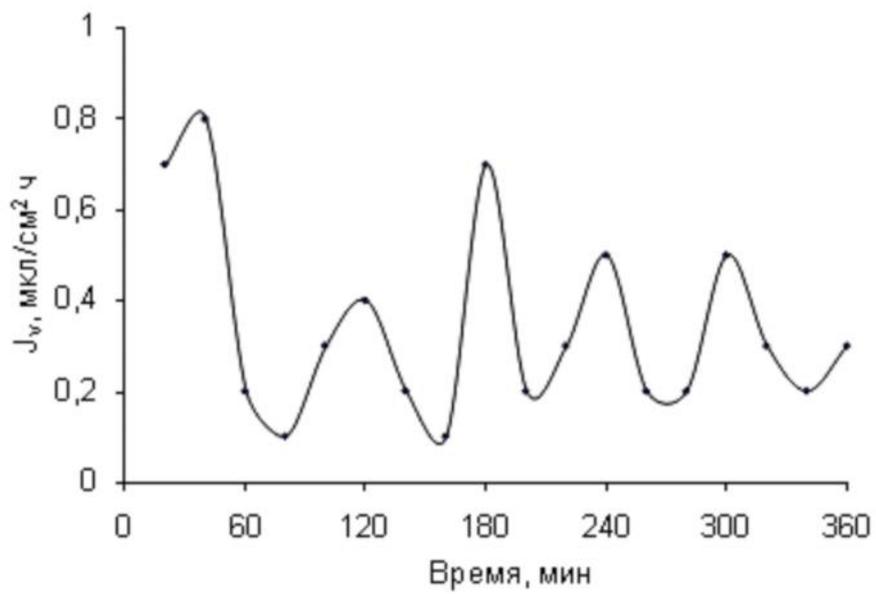
NaCl)

NaCl.



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1.

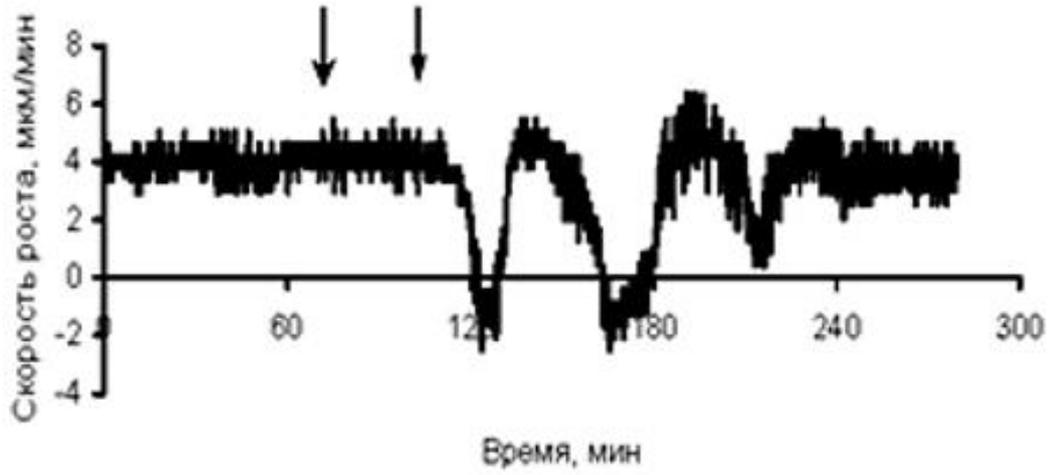


.7.

2.

2- ( .10).  
NaCl

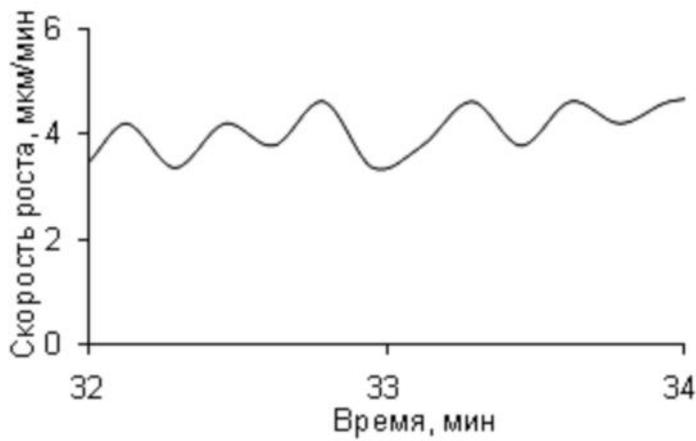
, , 2 , .8.



.8. 3- 16- NaCl (0,5 , 800  
↓ - ) ( 1 ) , ↓ -

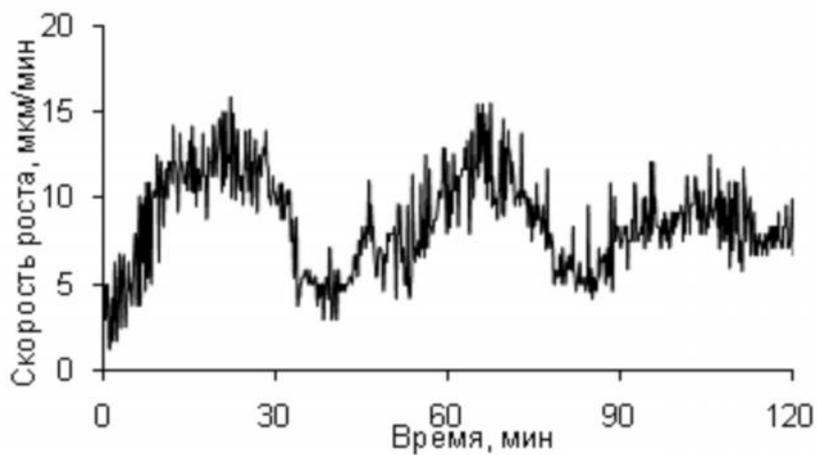
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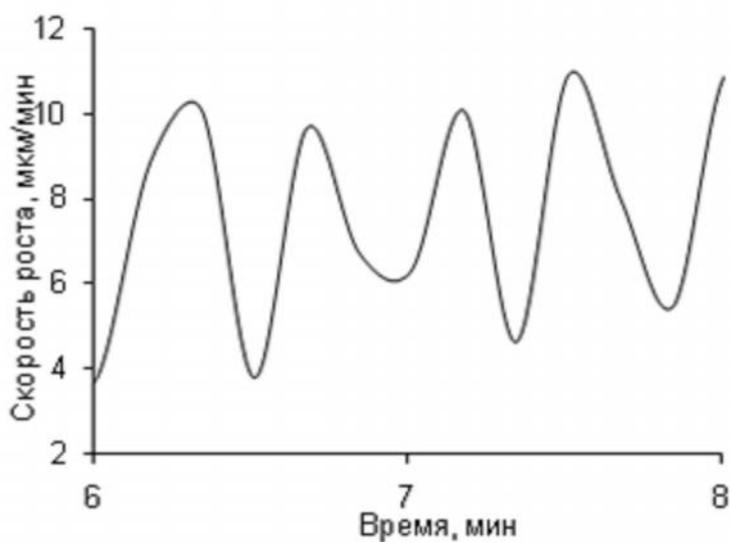


. 10. 2- 15-  
NaCl (1,0 , 800 ) ( )  
NaCl.

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11)

[11].

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(NaCl, , )  
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*de novo*

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1. Budagovskaya, N.V. "Golden section" in proportions of structural and functional parameters of photoautotrophic and heterotrophic organs and whole plant as an index of harmonic development //Plant Physiol. Biochem. – 1996 - sp. issue: SO4-2.
2. Budagovskaya, N.V. Golden section in structure-functional organization of plants //Biophotonics and Coherent Systems, L., Belousov, F-A., Pop, V., Voeikov, and R., van Wijk, (Eds.), Moscow University Press, Moscow, - 2000 - P. 67-74.
3. Budagovskaya, N.V., Interaction of plant shoots and roots: dynamics and stability //Biophotonics and Coherent Systems, Belousov L.V., Voeikov V.L., Martinuik V., (Eds.), Springer. - New-York, - 2007. - P. 213-223.





**FEOX-1**

• „ • „ • „ • „  
• •

-mail: *simacoffee@mail.ru*

*Sphaerotilus mobilis* Feox-1

*mofA*

IV)

Mn (III)

(III,

Mn

[2].

MofA

*Sphaerotilus*,

(II)

(II) [1].

*Sphaerotilus* – *S. mobilis* Feox-1.

( / ): – 0,25; –  
 0,25; MgSO<sub>4</sub>\*7H<sub>2</sub>O – 0,1; CaCl<sub>2</sub> – 0,05; MnSO<sub>4</sub>\*5H<sub>2</sub>O – 0,05;  
 – 1 ; 1 / (B12, B1)  
 1 / [3].  
 4  
 27 10

( )  
 – 0,05; 0,1; 0,15.

(II).

(IV)

BLAST. o o o o  
 - o RAST c c o o x o o o c c ([http://  
 rast.nmpdr.org/](http://rast.nmpdr.org/)).

ExtractRNA (« »),

)

1,5 %

Fisher Scientific, )

Qubit® RNA HS Assay Kit (Thermo  
 Qubit 2.0 (Thermo Fisher Scientific,

).

M-MulV (SibEnzyme, ) Eppendorf Mastercycler

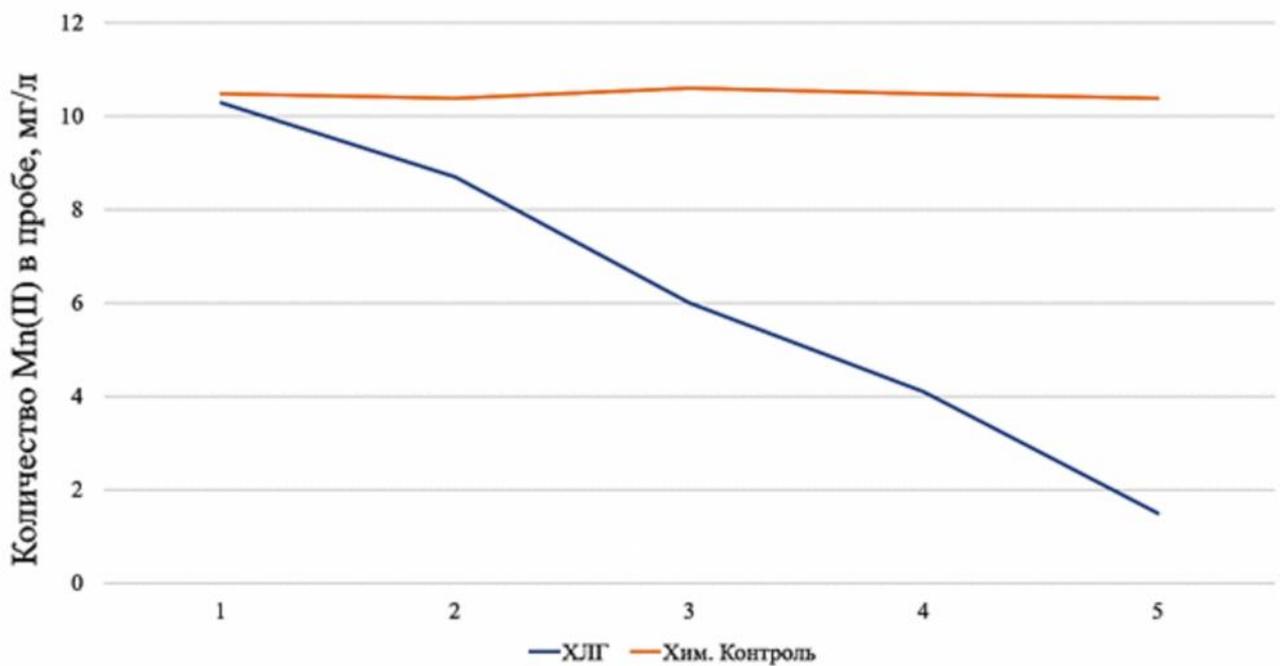
personal» [4].

MofA – (EC 1.16.3.3).

*S. mobilis* Feox-1

*S. discophora*.

*Sphaerotilus*



1. Mn (II) *S. mobilis* Feox-1.

*L. mobilis* Feox-1

*mofA* –

*mofA*

72

4

(рис. 2).

Mn (II)

Mn<sup>2+</sup>.

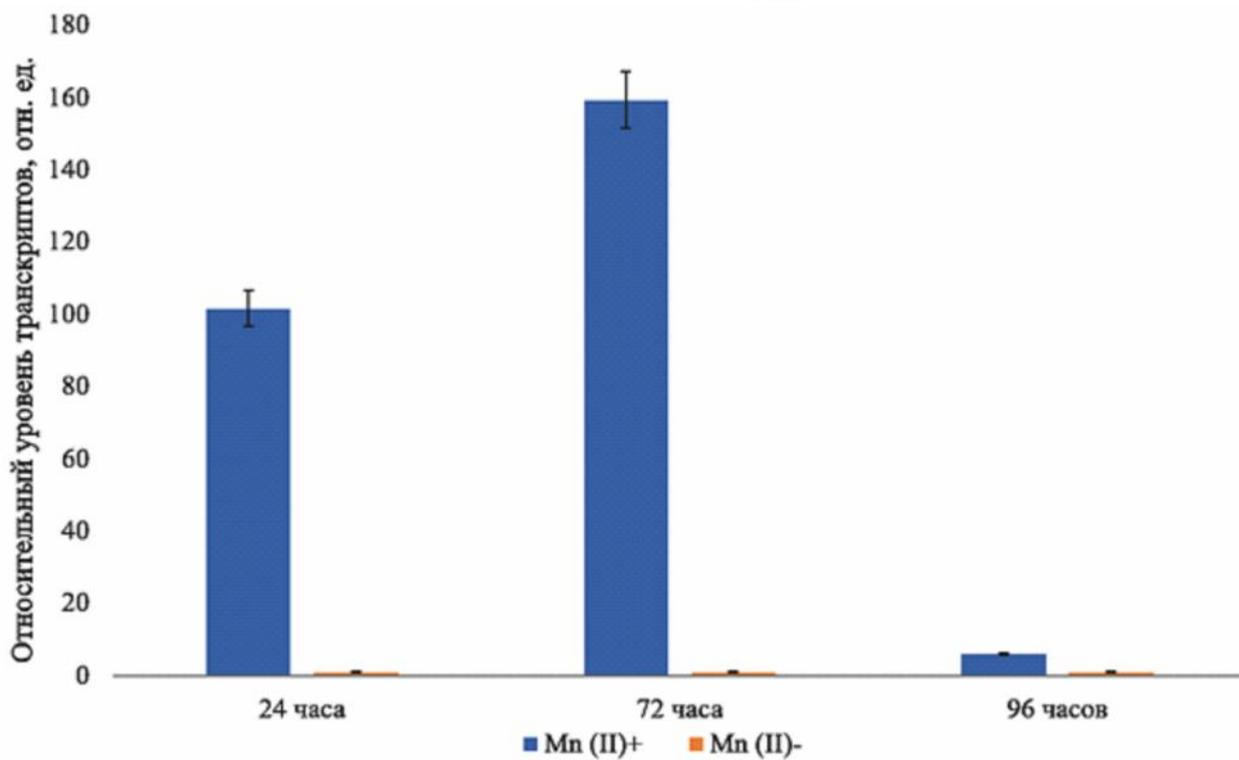


рис. 2.

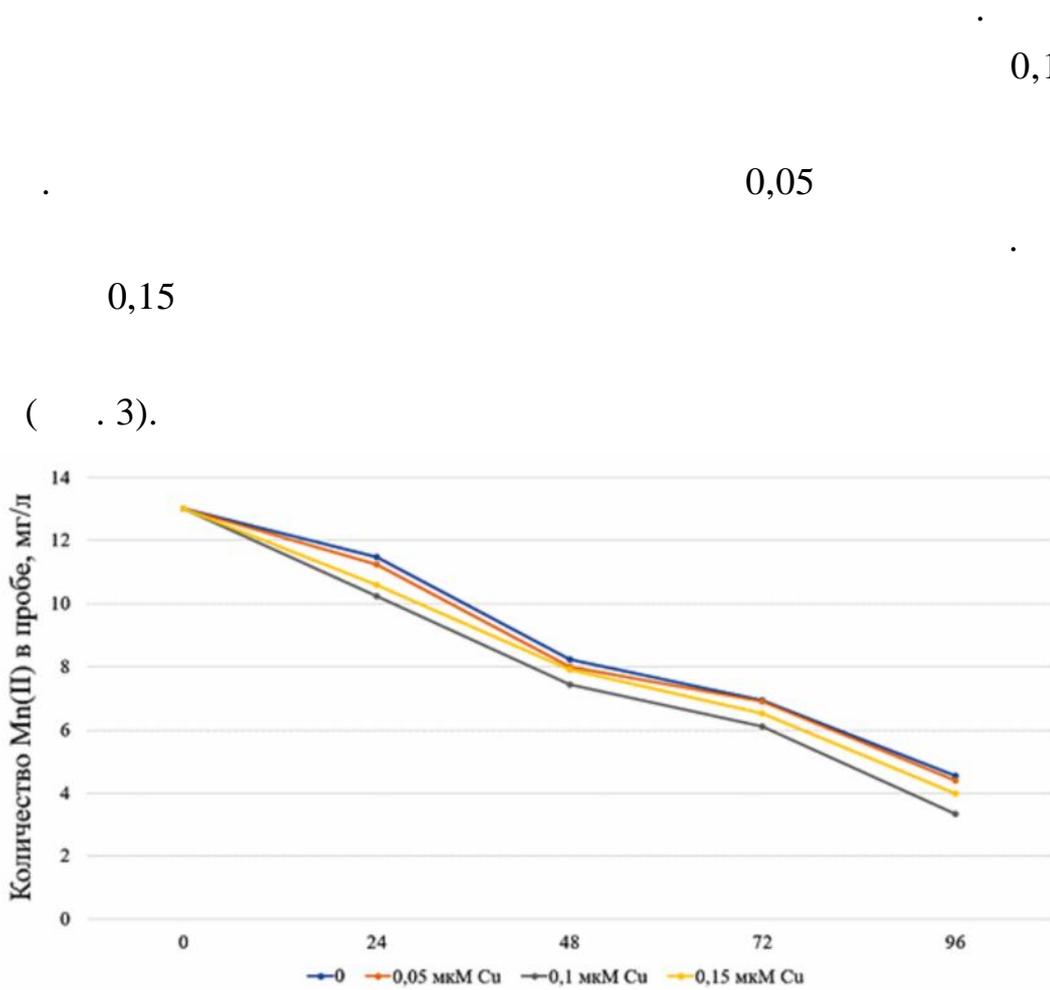
*L. mobilis* Feo -1.

*mofA*

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Cu (II)),

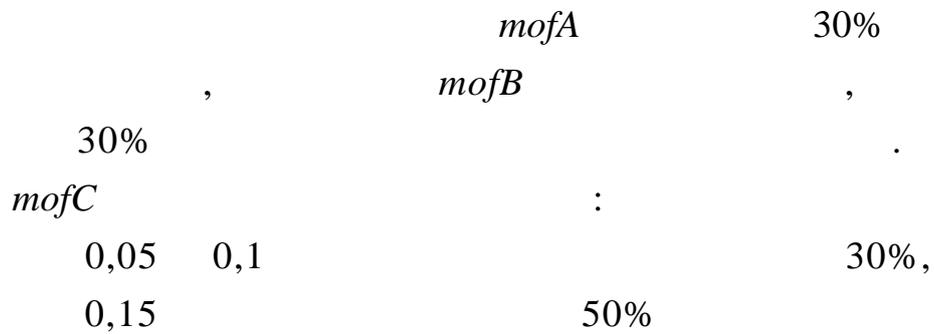
*L. mobilis* Feox-1



3. Mn (II)  
*S. mobilis* Feox-1.

*mofA*

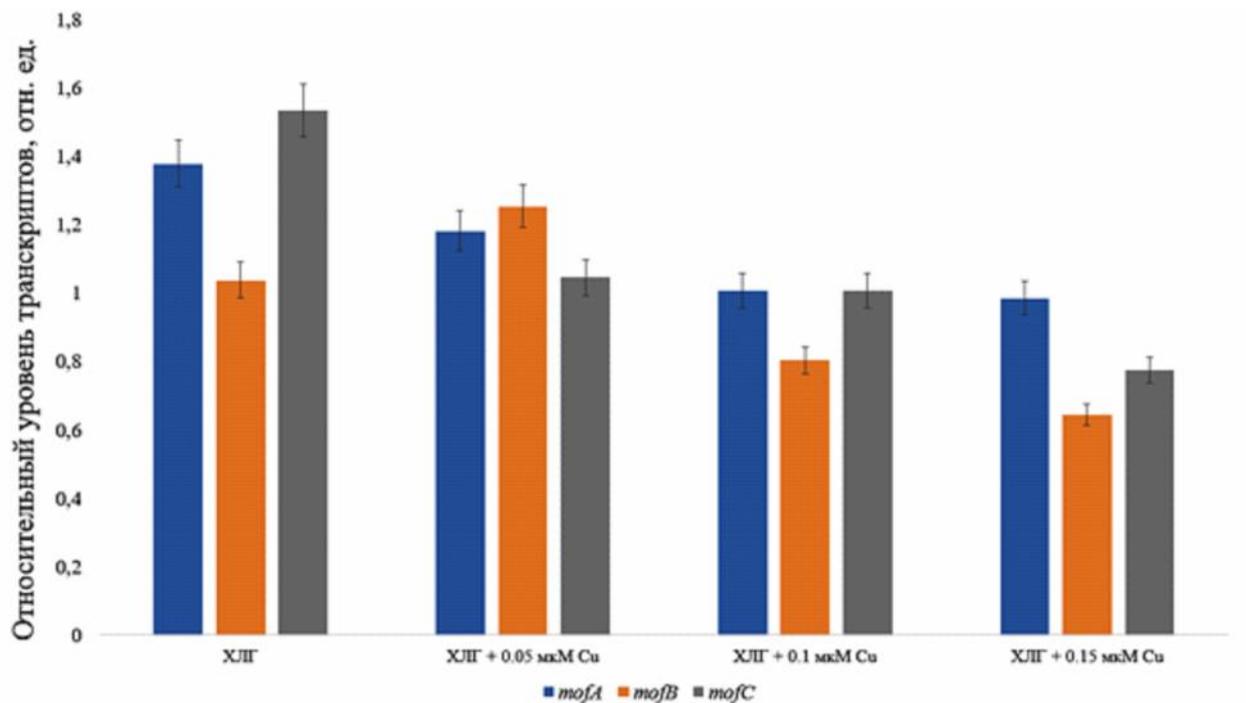
4).



*Sphaerotilus – S. discophora*

SS-1,

Cu (II)



4. *mofA* *L. mobilis* Feo -1 4

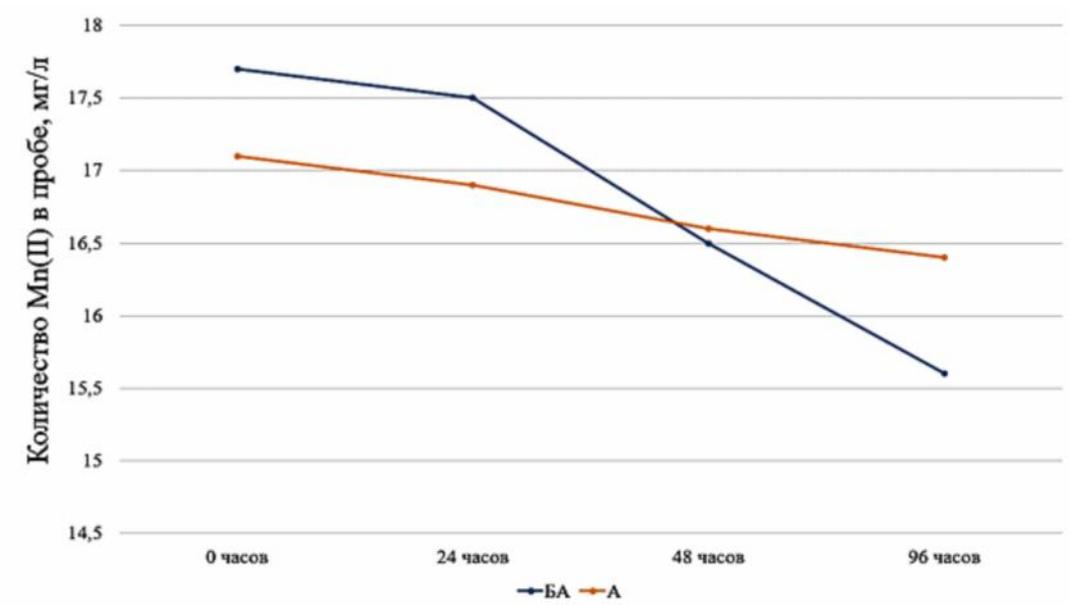
(NaN<sub>3</sub>) –

IV

8 / 19 /

96

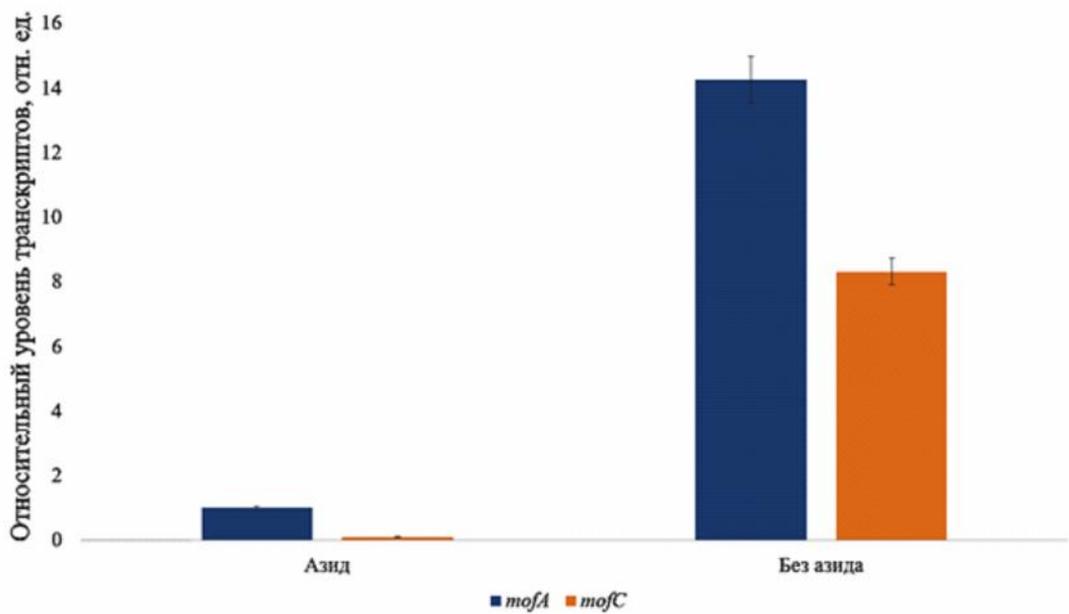
96 ( .5).



5. Mn (II)  
*L. mobilis* Feox-1.

(  
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14,2

*mofA*  
*mofA – mofB mofC*  
*mofC* 8,1 ( . 6).



6. *mofA, mofC* *S. mobilis* Feo -1.

NaN<sub>3</sub>

MofA

( ofC),

ofA

*S. mobilis*

1. . . . // . – 2017. – . 9. – . 4. – . 370-379.
2. Hansel C. M. Manganese in marine microbiology //Advances in microbial physiology. – 2017. – . 70. – . 37-83.
3. Okazaki M. et al. Partial purification and characterization of manganese-oxidizing factors of *Pseudomonas fluorescens* GB-1 //Applied and Environmental Microbiology. – 1997. – . 63. – . 12. – . 4793-4799.
4. Rudenko T. S. et al. *Azospirillum thiophilum* BV-S // . – 2018. – . 18. – . 3. – . 438-442.



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PEPCK-C

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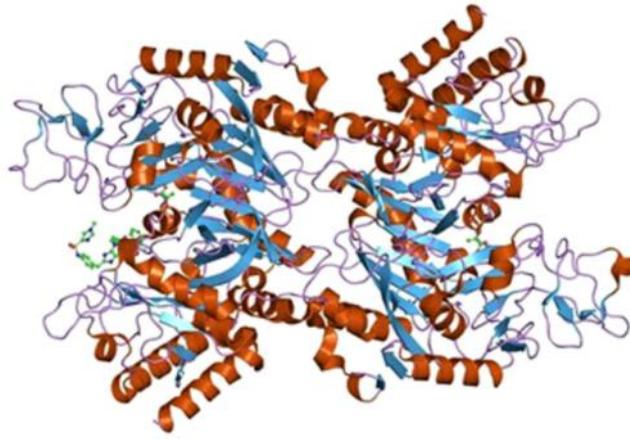
9

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PEPCK-

C

[5].



. 2.

[4].

PEPCK-C

60%

1. Advances in transgenic technologies and their impact on animal husbandry / L. Zhou, H. Chen // – *Animal Biotechnology*. – 2021. – Vol. 32, No. 1. – P. 1–14. – DOI 10.1080/10495398.2020.1758466.

2. A review of transgenic animal techniques and their applications / W. M. E. Shakweer, A. Y. Krivoruchko, Sh. M. Dessouki, A. A. Khattab // *Journal of Genetic Engineering and Biotechnology*. – 2023. – Vol. 21, No. 1. – P. 55. – DOI 10.1186/s43141-023-00502-z. – EDN CVIAQX.

3. Genetic modification of mice to study metabolic disorders / H. Wang, M. Zhang / – *Diabetes*. – 2018. – Vol. 67, No. 7. – P. 1273–1283. – DOI 10.2337/db18-0040.

- ....
4. Mitochondrial phosphoenolpyruvate carboxykinase (PEPCK-M) is a pro-survival, endoplasmic reticulum (ER) stress response gene involved in tumor cell adaptation to nutrient availability / A. M ndez-Lucas, P. Hyro ov , L. Novellademunt [et al.] // *Journal of Biological Chemistry*. – 2014. – Vol. 289, No. 32. – P. 22090-22102. – DOI 10.1074/jbc.M114.566927. – EDN YCOTIG.
  5. PEPCK-C and its role in muscle metabolism and exercise capacity / M. Parvin Hakimi, A. Koshy, M. Zubair // – *American Journal of Physiology-Endocrinology and Metabolism*. – 2019. – Vol. 317, No. 6. – P. E1054-E1064. – DOI 10.1152/ajpendo.00233.2019.
  6. The role of PEPCK-C in energy metabolism and physical performance in transgenic mice / Y. Zhou, K. Chen // – *American Journal of Physiology-Endocrinology and Metabolism*. – 2020. – Vol. 129, No. 4. – P. 1243–1253. – DOI 10.1152/ajpendo.00436.2020.
  7. The role of transgenic mice in understanding the mechanisms of learning and memory / T. J. Huang, Y. Liu // – *Frontiers of Behavioral Neuroscience*. – 2018. – Vol. 12. – P. 51. – DOI 10.3389/fnbeh.2018.00051.
  8. Transgenic mice: a powerful tool for studying human diseases / T. Miyazaki, T. Saito // – *Nature Reviews Genetics*. – 2019. – Vol. 20. – P. 579–598. – DOI 10.1038/s41576-019-0102-0.
  9. Transgenic models for studying the mechanisms of memory and learning / B. Li, Y. Wang // – *Journal of Neurology Methods*. – 2022. – Vol. 358. – Article 109157. – DOI 10.1016/j.jneumeth.2022.109157.
  10. Transgenic mouse models for cancer research / Yu. Wang, Yu. Zhang // – *Cancer Research*. – 2016. – Vol. 76, No. 16. – P. 4672–4680. – DOI 10.1158/0008-5472.CAN-16-0726.
  11. Transgenic technology in mice: application and future prospects / K. Tamura, M. Tada // – *Molecular Genetics and Genomics*. – 2020. – Vol. 295, No. 1. – P. 1–11. – DOI 10.1007/s00438-019-01634-7.



C57Bl/6,

“ ” ( , ).

32

(n=12)

ad libitum

, (n=10)

L-

(Sigma-aldrich, St. Louis, MO, USA) (200 / )

(n=10)

(Grindex, ,

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Lueptow (2017).

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d1

; d2

(d3).

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d2

(p<0,05).

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(p<0,05),

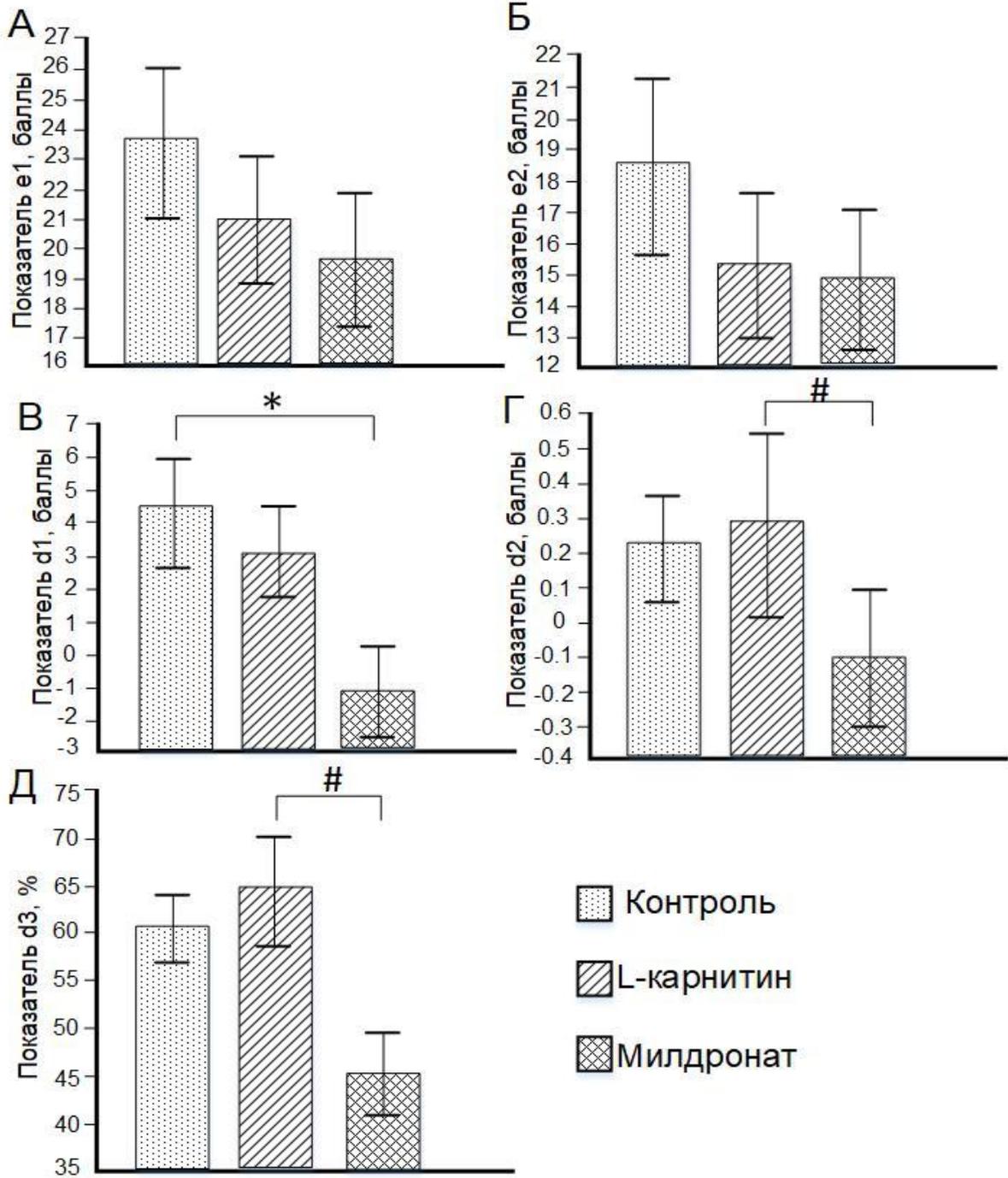
1,36

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( . 1).

L-

[3].



e1 ( ), p<0,05 .1. 2 ( ), L- d1 ( ), (#). d2 ( ), : d3 ( ), (\*),

[5].

1. Pharmacological effects of meldonium: Biochemical mechanisms and biomarkers of cardiometabolic activity/ M. Dambrova [et al.] // *Pharmacol Res.* – 2016. – Vol. 113. – P. 771-780.
2. Bellman V. Unlocking the Potential of Meldonium: From Performance Enhancement to Therapeutic Insights *Psychoactives.* – 2024. – Vol. 3. – P. 235-247.
3. Ferreira G.C. L-Carnitine and Acetyl-L-carnitine Roles and Neuroprotection in Developing Brain / G.C. Ferreira, M.C. McKenna // *Neurochem Res.* – 2017. – Vol. 42. – P. 1661-1675.
4. Neuroprotective Effects of Carnitine and Its Potential Application to Ameliorate Neurotoxicity / L.E. Latham [et al.] // *Chem Res Toxicol.* – 2021. – Vol. 34. – P. 1208-1222.
5. Long-term mildronate treatment increased Proteobacteria level in gut microbiome, and caused behavioral deviations and transcriptome change in liver, heart and brain of healthy mice / A.P. Gureev [et al.] // *Toxicol Appl Pharmacol.* – 2020. – Vol. 398. – P. 115031.



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-mail: *bc366@bio.vsu.ru*

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[1].

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[3].

( , , 4.2.1.2) –

[2,4].

*(Rattus norvegicus L.)*

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NaCl.

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240 5

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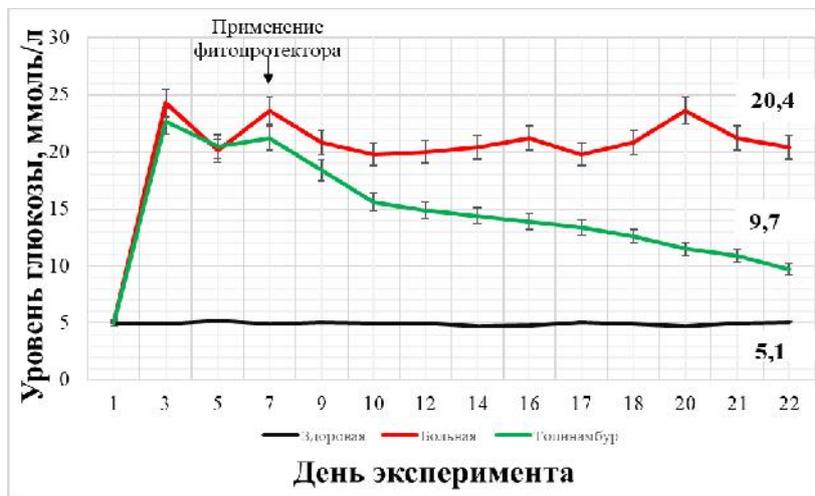
[2,6].

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[7]. 10

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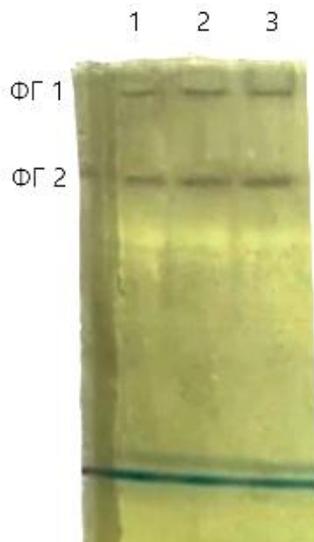
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0,28 / . . .

0,59 / . . . , 2,1



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3).

Rf 1 2 0,12 0,20,

1. 1994. — 383 .
2. — 2005. — 224 .
3. « 2- / . — 2008. — 4. — . 17-18.
4. : , 2012. — 528 .
5. Lenzen S. Alloxan : history and mechanism of action / S. Lenzen, U. Panten // Diabetologia. 1988a. — Vol. 31. — P. 337—342.
6. // . — 2017. — .7, 1. — .27—37.
7. / . — 2- ., 2006. — 134 .



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*IN VIVO*

• „ • „ • •

E-mail: *dasadragunova96129@gmail.com*

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150 M 200

[1].

( , 1.2.2.16).

[2].

( , 1.1.1.61),

[3].

(*Zea mays* L.).

(*Zea mays* L.)

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“LabTech” ( )

25 / 2

25 .

24 .

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: 50 Tris-HCl ( 9.0), 0,3 , 3 , 0,1

CaCl<sub>2</sub> 0,05% Tween 80, 3000 /

5 .

Evolution 260 Bio (Thermo Fisher

Scientific, )

340

Aldrich, ), 1 , : 16  
, 100 Tris-HCl ( 9.0).

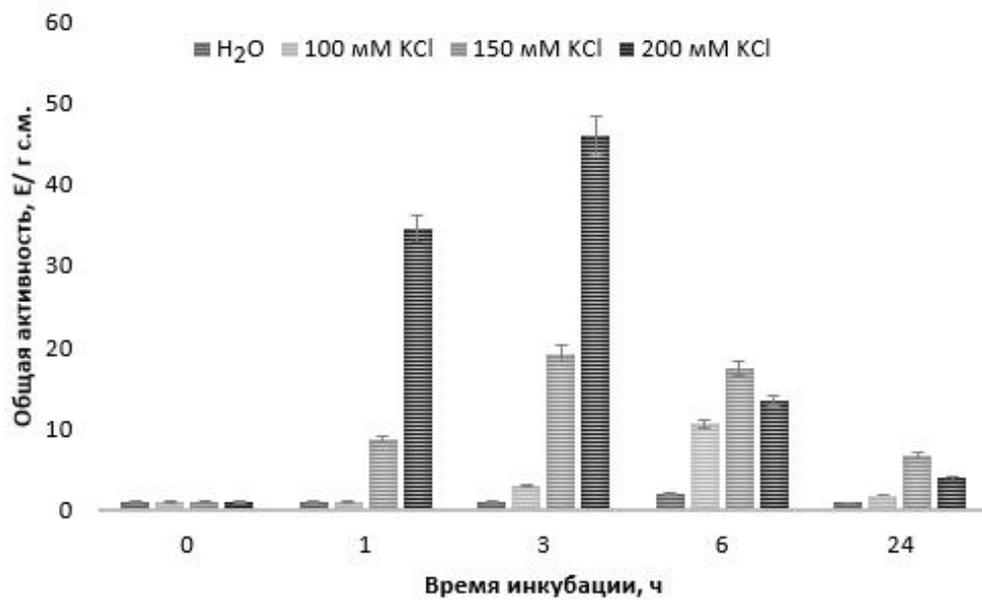
3-

STATISTICA

12.0.

± (SEM).  
( $p < 0.05$ ).

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KCl. 100 , 150 200 -  
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KCl

1. . . . //  
( 18-04-20028 18-04-20023)  
. – 2018. – . 343.
2. Ji J. et al. Roles of -aminobutyric acid on salinity-responsive genes at transcriptomic level in poplar: Involving in abscisic acid and ethylene-signalling pathways //Planta. – 2018. – . 248. – . 675-690.
3. Taxon E. S., Halbers L. P., Parsons S. M. Kinetics aspects of Gamma-hydroxybutyrate dehydrogenase //Biochimica et Biophysica Acta (BBA)-Proteins and Proteomics. – 2020. – . 1868. – . 5. – . 140376.



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E-mai: *sveta.v05@yandex.ru*

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METI,

Dnmt1

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*de novo.*

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Dnmt3  
*de novo.*

CpNpG,

SNF2/ SWI2

DDM1,

[3].

CG

CNG,

CNG

CNN.

*met1*

CNG CNN.

*met1-drm-1drm2*

CNG

CpG

[3].

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[3].

(*Zea mays* L.)

76,

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NaCl, 10

MgCl<sub>2</sub>, 1

DTT,

7,9, 80

SAM, 40

-HCl, 50  
[15].

256

S-

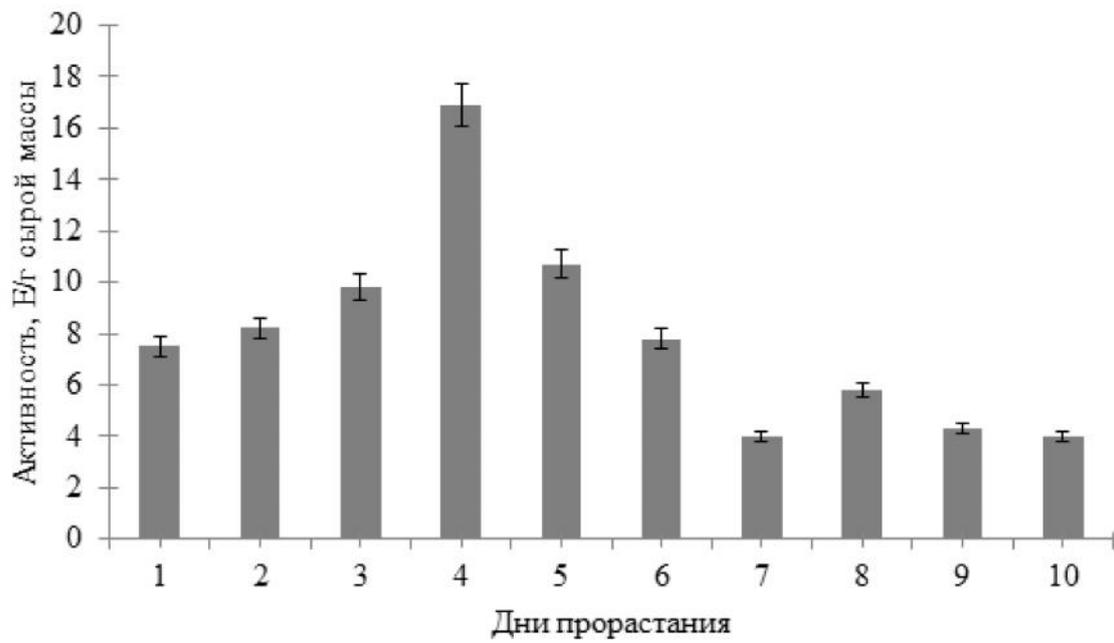
[12].

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16,9 / ( .1).

4,0 / 10



.1.

1. . . . . // .  
1991. . 56. . 361–368.
2. . . . . , 2001.
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. 1–14.
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. 2010. . 6. . 21–29.
6. . . . . / . . . . - : . . . . , 1990. - 351 .
7. . . . . , 2012.
8. . . . . // : . -  
2010 – . 496.
9. . . . . ? // . . . . ,  
. 2014.
10. Alexander Meissner, Jörn Walter. Epigenetic Mechanisms in Cellular Reprogramming. 2015 244 p.
11. Antequera F., Bird A. CpG islands as genomic footprints of promoters that are associated with replication origins // *Curr. Biol.* – 1999. – V. 9. R661-R667.
12. A simplified characterization of S-adenosyl-l-methionine-consuming enzymes with 1-Step EZ-MTase: a universal and straightforward coupled-assay for in vitro and in vivo setting / E.S. Burgos [et al.] // *Chem Sci.* – 2017. – V. 8. – P. 6601-6612.
13. Bartee L., Malagnac F., Bender J. Arabidopsis cmt3 chromomethylase mutations block nonCG methylation and silencing of an endogenous gene // *Genes Dev.* 2001. V. 15. P. 1753–1758.
14. Callebaut I., Courvalin J.C., Mornon J.P. The BAH (bromoadjacent homology) domain: A link between DNA methylation, replication and transcriptional regulation // *FEBS Lett.* 1999. V. 446. P. 189–193.
15. Cokus S.J. Shotgun bisulphite sequencing of the Arabidopsis genome reveals DNA methylation patterning / S.J. Cokus [et al.] // *Nature.* – 2008. - V. 452. - P. 215–219.
16. Finnegan E.J., Plant DNA methyltransferases / E.J. Finnegan, K.A. Kovac // *Plant Mol. Biol.* - 2000. - V. 43. - P. 189–210.



577.12

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-mail: *anna.khomutova2002@gmail.com*

- (RdDM)

RdDM,

RdDM

*thaliana.*

*Arabidopsis*  
RdDM

[1].

(RdDM) [2].

( ),

[3].

RdDM

RdDM,

[4].

( CG, CHG CHH- )

(piRNA). (siRNA), (miRNA), piwi-RdDM siRNA,

miRNA. MiRNA-

« » miRNA (20-22 ), miRNA [6].

miRNA,

RdDM Pol IV ( - 4),

RDR2 ( - - 2), DCL3 (Dicer-like 3) AGO4 (Argonaute 4) [7]. RdDM 24-

IV

(siRNA) Pol IV. Pol (ssRNA),

(dsRNA) RDR2. dsRNA

DCL3, siRNA (siRNA 24 ). siRNA

AGO4, AGO4 RISC ( - ) [8].

(pri-miRNA)

DGCR8 III DROSHA. 3 (METTL3)

pri-miRNA, m<sup>6</sup>A DGCR8.

METTL3 DGCR8

pri-miRNA miRNA

pri-miRNA. ,

m<sup>6</sup>A ,

miRNA [9].

miRNA (*Oryza sativa*)

miRNA, 24

miRNA ( 24 miRNA, lmiRNAs). [10]. *Arabidopsis thaliana* DCL3-

« » miRNA

[11].

lmiRNA

[10].  
 (RdDM),  
 miR2936 miR398,  
 AGO1 AGO4-  
 (RdDM). AGO4- RdDM  
 RPW8 4 (HR4)  
 HR4 ago4-1,  
 APETALA2 12 (RAP2.12),  
 [12].

(*Brassica oleracea*) 508  
 (DMR); 59,92% DMR  
 , 39,43% miRNA  
 31 (DEG)  
 miRNA, (ROS) (ABA),  
 , miRNA  
 ROS ABA [13].  
 miRNA  
 miRNA,  
 miRNA  
 . [14].  
 ..., 2025, 27.

-

....

RdDM,

*Marchantia polymorpha*

*Phsycomitrella patens. M. polymorpha*

[15].

*M. polymorpha P. patens* 21

24

*Arabidopsis* [16],

*Arabidopsis Oryza sativa*

24

*Arabidopsis*

[11].

24

miRNAs,

MpAGO4a, MpAGO9,

-

IV-V

MpDCL3

*M. polymorpha.*

, *M.*

*polymorpha*

(miR160, miR166, miR171, miR319/miR159,

miR390, miR408 miR529/miR156)

, miR160 miR166

MpC3HDZ1 MpARF3,

[15].

RdDM

RdDM –

1. . – 2006. – . 42. – . 1-14. //
2. Regulation and function of DNA methylation in plants and animals / He X.J. [et al.] // *Cell Res.* – 2011. – V.21. – P.442-465.
3. Erdmann R.M. RNA-directed DNA Methylation / R.M. Erdmann, C.L. Picard // *PloS Genetics.* – 2020. – V. 16:e1009034.
4. Chinnusamy V. RNA-directed DNA methylation and demethylation in plants / Chinnusamy V., Zhu JK // *Sci China C Life Sci.* – 2009. – V.52. – P.331-343.
5. Holoch D. RNA-mediated epigenetic regulation of gene expression / Holoch D, Moazed D // *Nat Rev Genet.* – 2015. – V.16. – P.71-84.
6. DNA Methylation in Plants by microRNAs / Teotia [et.al.] // *Plant Epigenetics. RNA Technologies.* – 2017. – V.13. – P.247-262.
7. Generation of a luciferase-based reporter for CHH and CG DNA methylation in *Arabidopsis thaliana* / Dinh T. [et al.] // *Silence.* – 2013. – V.4. – P.1–11.
8. Systematic genome-wide and expression analysis of RNA-directed DNA methylation pathway genes in grapes predicts their involvement in multiple biological processes / Xiang R [et.al.] // *Front. Plant Sci.* – 2022. – V13:e1089392.
9. N6-methyladenosine marks primary microRNAs for processing / Alarcon C. [et. al.] // *Nature.* – 2015. – V.519. – P.482-485.
10. DNA Methylation Mediated by a MicroRNA / Wu L [et al.] // *Molecular Cell.* – 2010. – V. 38. – P.465-475.
11. Evolution of *Arabidopsis* MIR genes generates novel microRNA classes / Vazquez F. [et al.] // *Nucleic Acids Res.* – 2008. – V.36. – P.6429-6438.
12. ARGONAUTE1 and ARGONAUTE4 Regulate Gene Expression and Hypoxia Tolerance / Loreti E [et al.] // *Plant Physiol.* – 2020. – V.82. –P. 287-300.
13. Global DNA Methylation and mRNA-miRNA Variations Activated by Heat Shock Boost Early Microspore Embryogenesis in Cabbage (*Brassica oleracea*) / Kong C. [et. al.] // *Int. J. Mol. Sci.* – 2022. – V.23:5147.
14. Evolution and Functional Diversification of MIRNA Genes / Josh T. [et.al.] // *The Plant Cell.* – 2011. – V. 23. – P.431-442.
15. Identification of miRNAs and their targets in the liverwort *Marchantia polymorpha* by integrating RNA-Seq and degradome analyses / Lin PC [et. al.] // *Plant and Cell Physiology.* –2016. – V.57. – P.339–358.
16. Lin SS, Bowman JL. MicroRNAs in *Marchantia polymorpha*. /Lin SS, Bowman JL. // *New Phytol.* – 2018. – V.2. – P.409-416.



582.623.2:573.6

**IN VITRO**

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« - ».

E mail: [natalya.vnuckova@yandex.ru](mailto:natalya.vnuckova@yandex.ru)

L-

*in vitro.*

200 /

*in vitro*

(1-10° ),

80-90%

12  
[1].

*in vitro*

2 4 .

( ).

[2, 3].

..., 2025, 27.

*in vitro* [4, 5].

L-

[6].

: 1 -

*Salix dasyclados* Wimm., 3 - *S. caspica* Pall., 5 - *S. purpurea* L., 6 - *S. palustris* Host. 7 - *S. viminalis* L. [7]. WPM,

( 25±2° , 2,0 ).

16 /8

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(30-80%),

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50%,

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(20-30%).

	WPM		%	1	1	,% <sup>1</sup>
	/	/				
1	-	-	100	4,6±0,42	4,3±0,50	100
	100	-	90	4,2±0,20	4,0±0,33	100
	200	-	100	3,4±0,33	3,6±0,29	100
	-	5	80	2,3±0,30 <sup>2</sup>	3,0±0,38	50
	-	10	60	1,4±0,39 <sup>3</sup>	1,5±0,51 <sup>3</sup>	30
3	-	-	100	4,4±0,32	7,1±0,62	100
	100	-	100	4,1±0,58	5,8±1,02	90
	200	-	100	3,7±0,19	6,9±0,57	100
	-	5	60	2,8±0,91	4,2±1,43 <sup>2</sup>	60
	-	10	60	2,7±0,74	4,0±0,89 <sup>2</sup>	30
5	-	-	100	3,3±0,32	3,6±0,22	100
	100	-	100	3,8±0,30	4,2±0,33	100
	200	-	100	3,5±0,34	3,8±0,41	100
	-	5	80	1,0±0,21 <sup>3</sup>	1,4±0,32 <sup>3</sup>	70
	-	10	50	0,9±0,25 <sup>3</sup>	1,0±0,09 <sup>3</sup>	30
6	-	-	100	4,7±0,30	7,7±0,44	100
	100	-	100	4,5±0,50	6,2±0,62	100
	200	-	100	3,3±0,43 <sup>2</sup>	6,0±0,63	100
	-	5	70	3,0±0,42 <sup>3</sup>	3,5±0,31 <sup>3</sup>	40
	-	10	30	2,9±0,49 <sup>3</sup>	3,3±0,51 <sup>3</sup>	20
7	-	-	100	4,2±0,43	5,7±0,78	100
	100	-	100	4,1±0,51	6,0±0,71	100
	200	-	100	4,4±0,57	6,1±0,47	100
	-	5	50	2,9±0,38	4,3±0,59	30
	-	10	50	2,7±0,70	4,0±0,64	20

3

2\_ <0,05

3\_ <0,001

(90-100%),

3

100%.

200 /

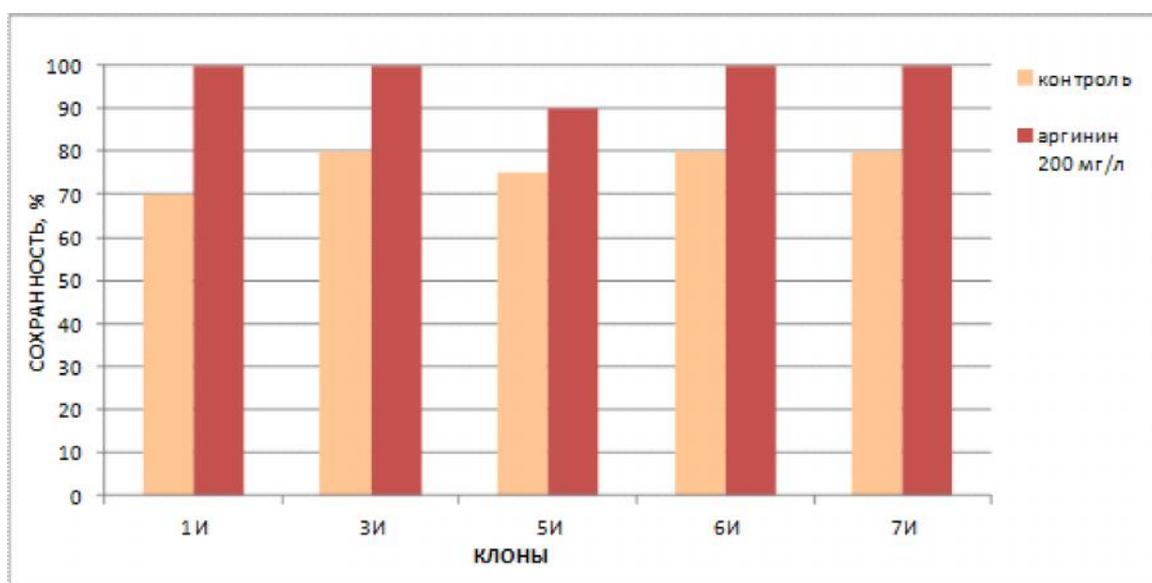
4 90-100% 70-80%

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WPM 2



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(5-10 / )

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(200 / ): 20%

8-





577.12

### MIR159

• „ • •

E-mail: *ulia.volk54@gmail.com*

miR159B

miR159b

[8].

(miR159 miR319),

(miR172),  
(miR165/166),

(miR164),

(miR160) [2].

miR159B

[2].

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, 11

miR159.

MIR159,

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[6].

mays L.) 76, (Zea

25 10 .

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NaCl;

3, 6, 12 24 .

- -  
LiCl.

M-MuLV ( , )

miR159b.

16°C - 30 , 42°C - 30 , 85°C - 5 [1, 4].

AmpliSence ( , ).

*ef-1* [13].

mir: 5'-ACCTGGCG GGA GAAG-3',

5'-GTGCAGGGTCCGAGGT-3'.

- 95° 5 , - 95° - 30 ., 58° - 30  
., 72° - 30 . ( ), - 72° - 10 .

Opticon Monitor™ Software (Bio-

Rad, )

miR159b 2-<sup>Ct</sup>- [5].

« - » miR159b

1.

« - », [7].

miR\_159b

[4]:

1)

5'-UUUGGAUUGAAGGGAGCUCUU-3'

44

« - », 5'GTCGTATCCAGTGCAGGGTCCGAGGTATTCGCACTGGA  
TACGAC-3' 3'-

« - » miR159b:

5'GTCGTATCCAGTGCAGGGTCCGAGGTATTCGCACTGGATACGACA  
AGAGC-3'

2)

5'-ACCTGGCG GGA GAAG-3'

3)

5'-CAG CAT AGG TCA CGT CCC A-3'

44

3'-

21-

65

5'-

5'-

miR159b

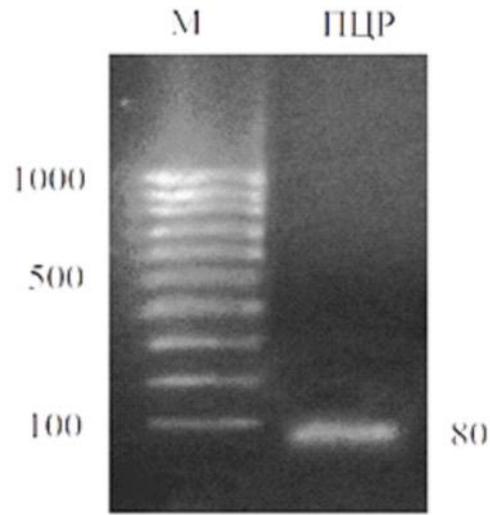
71-

miR-H1,

miR159b

miR159b

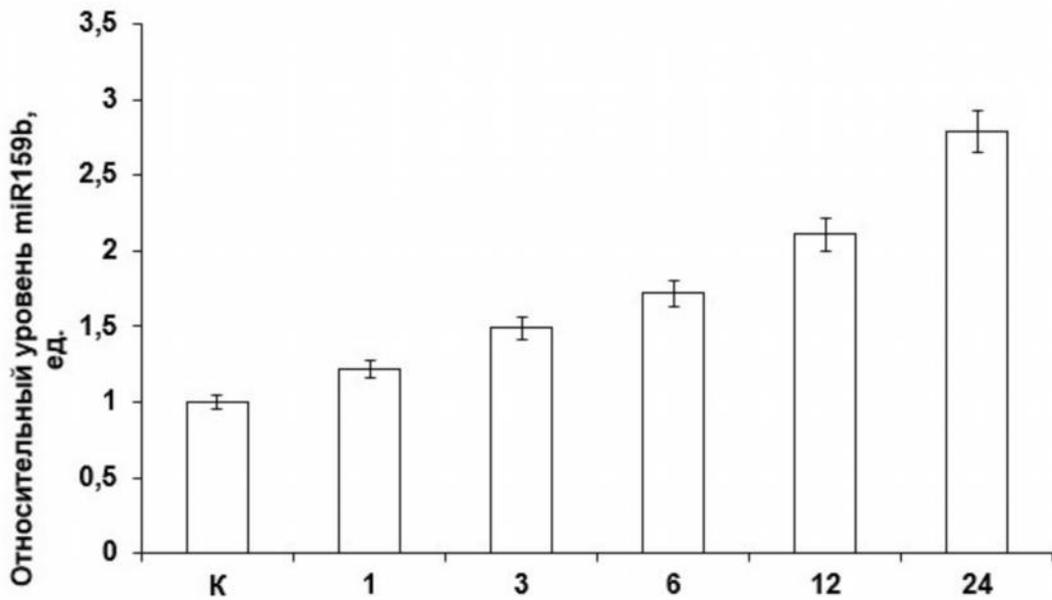
(.1).



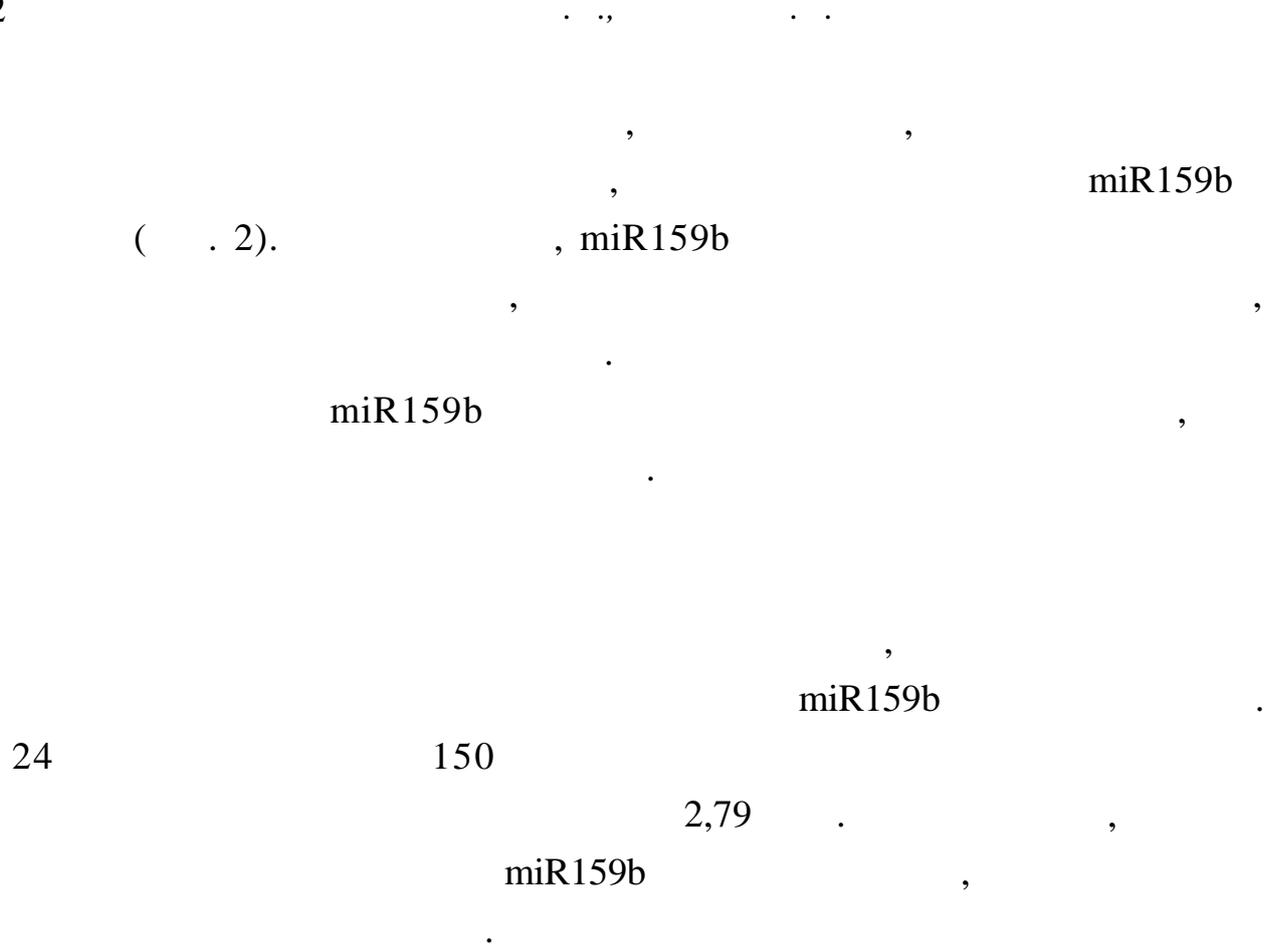
. 1. miR159b. -

~70 ..

miR159b



. 2. miR159b



1. A Uniform System for the Annotation of Vertebrate microRNA Genes and the Evolution of the Human microRNAome / B. Fromm [et al.] // *Annu. Rev. Genet.* - 2015. - V. 23. - P. 213–242.
2. Axtell M.J., Antiquity of MicroRNAs and Their Targets in Land Plants / M.J. Axtell, D.P. Bartel // *The Plant Cell.* - 2005. - V. 17. - P. 1658–1673.
3. Housekeeping gene selection for real-time RT-PCR normalization in potato during biotic and abiotic stress / N. Nicot [et al.] // *J. of Exp. Bot.* - 2005. - V.56. - P. 2907–2914.
4. Kramer M.F. STEM-LOOP RT-qPCR for miRNAs / M.F. Kramer // *Curr Protoc Mol Biol.* – 2011. - CHAPTER: Unit15.
5. Livak K.J. Analysis of relative gene expression data using real-time quantitative PCR and the 2-<sup>-</sup> Ct method / K.J. Livak, T.D. Schmittgen // *Methods.* - 2001. - V.25. - P. 402–408.
6. MicroR159 regulation of most conserved targets in Arabidopsis has negligible phenotypic effects / R.S. Allen [et al.] // *Silence.* - 2010. - V. 1:18.
7. Realtime quantification of microRNAs by stem-loop RT-PCR / C. Chen [et al.] // *Nucleic Acids Res.* - 2005. - V. 33:e179
8. Small RNAs: The Essential Regulators in Plant Thermotolerance / Z-F. Zuo [et al.] // *Front. Plant Sci.* - 2021. - V. 12:726762.



591.1

**EUBLEPHARIS MACULARIUS**

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• - , , ,

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*(Eublepharis macularius)* -

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( ) [1].

- ( , , ) .

*Lemon frost*

[4].

*Enigma.*

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Enigma snow

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[2].

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 . – 2022. – // . 4 (96).  
 – . 233-238.
2. . . . .  
 , 29 2023 , . ( IV ) – 2023.  
 – . 66.
3. : ./ . . - .:  
 - , 2005. - . 77-81.
4. ./ . - .:  
 - , 2005. - .  
 128, 219, 293.



577.12

E-mail: *saidjamel-eddine@yandex.ru*

150

<sup>2</sup>: PDH,

, DLD,

( , , , , , . .),

[2].

( (NaCl))

[7].

[6].

(PDC)

[3].

NADH.

2.3.1.12)

(E1, 1.2.4.1),

(E3, 1.8.1.4).

(E2,

[4].

PDC

[5].

NCBI

3

: PDH-

, DLAT-

, DLD-

PDH, DLAT, DLD

(*Zea mays* L.),

25 , 12-

25 / <sup>2</sup>.

150

1, 3, 6, 12, 24

24 .

12 LiCl.

MMLV RT kit ( « »)

NCBI

PrimerBlast ( 1).

1.

Название		Последовательность нуклеотидов	Размер продукта, п.н.	Температура отжига, °C
PDH	Прямой	GCTGCCAACCTTGAGAGGAT	167	59
	Обратный	TCTTTCTGCAGCACGGACAT		
DLAT	Прямой	TGGGTGCTTCTCACCCATAG	192	62
	Обратный	TACACACATCCTTACCCACCT		
DLD	Прямой	CTTGGAACCCATGTGCAGGA	134	61
	Обратный	ACCGCAACTGGCTCATTTCAT		

LightCycler96 («Roche», )

Taq-

( « »)

SYBR Green I.

..., 2025,

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EF-1A

2<sup>-</sup> Ct<sub>1</sub>

3-4-

(Excel, Microsoft Office),

[1].

PDH,

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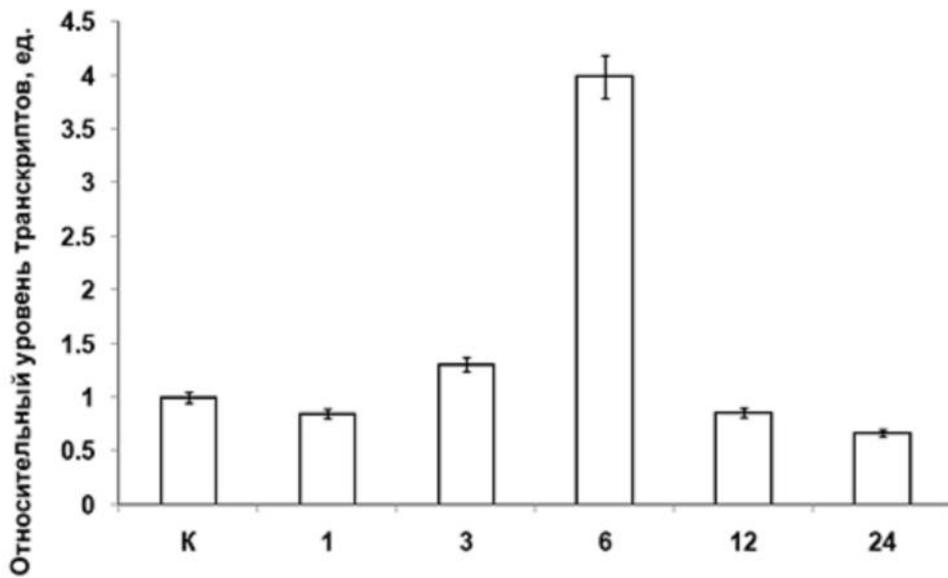
6

( 1.2.4.1),

150

NaCl

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PDH

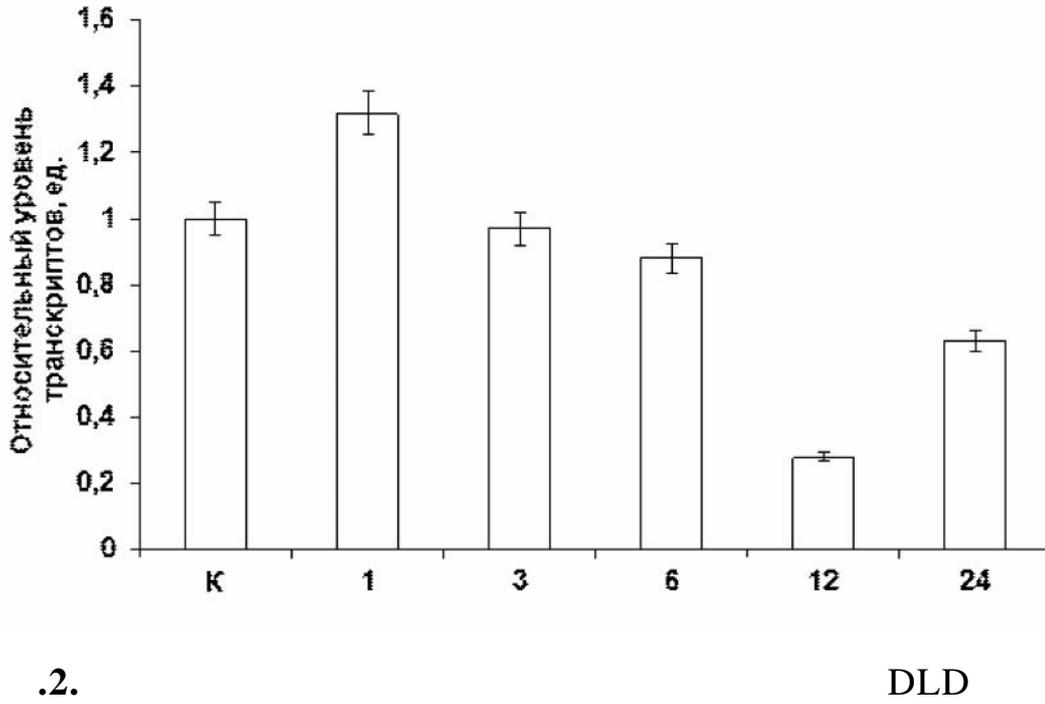
DLD,  
(E3, 1.8.1.4) – 3-

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0.4 ( . 2).



.2.

DLD

2

: PDH,

DLD,

DLAT,

24 ,

PDH

DLD

PDH

6

150

DLD

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1. . . . / . . . . - .: . . . , 1990. - 351
2. Salt Stress Tolerance in Rice: Emerging Role of Exogenous Phytoprotectants / A.Rahman [et al.] // *Advances in International Rice Research*. — 2017. — DOI: 10.5772/67098.]
3. Role of the Pyruvate Dehydrogenase Complex in Metabolic Remodeling: Differential Pyruvate Dehydrogenase Complex Functions in Metabolism / S. Park [et al.] // *Diabetes Metab.* — 2018. — Vol. 42, 4. — P. 270 — 281
4. Tovar-Mendez A. Regulation of pyruvate dehydrogenase complex activity in plant cells / A. Tovar-Mendez , J. A. Miernyk, D.D. Randall // *European Journal of Biochemistry*. — 2003. — Vol. 270, 6. — P. 1043 — 1049
5. Structure of the native pyruvate dehydrogenase complex reveals the mechanism of substrate insertion / J. Skerlova [et al.] // *Nat Commun.* — 2021. — Vol. 12.
6. Mechanisms of Salt Tolerance and Molecular Breeding of Salt-Tolerant Ornamental Plants / J. Guo [et al.] // *Front. Plant Sci.* — 2022. — Doi: 10.3389/fpls.2022.854116.
7. Stavi I. Soil Salinity and Sodicity in Drylands : A Review of Causes, Effects, Monitoring, and Restoration Measures / I. Stavi, N. Thevs, S. Priori // *Front. Environ. Sci.* — 2021. — Doi: 10.3389/fenvs.2021.712831.



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E-mail: *druzhson@mail.ru*

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[1].

1. ... // ... – 2000. – 4 (18). – P. 111-112.
2. ... // ... – 2016. – 61, 4. – P. 793-798.
3. « ... », 2021.
4. Beniczky S., Schomer D.L. Electroencephalography: basic biophysical and technological aspects important for clinical applications / S. Beniczky, D.L. Schomer // *Epileptic Disorders*. – 2020. – Vol. 22, No. 6. – P. 697-715. DOI: 10.1684/epd.2020.1217.
5. Jasper H.H., Pertuiset B., Flanigin H. EEG and cortical electrograms in patients with temporal lobe seizures / H.H. Jasper, B. Pertuiset, H. Flanigin // *Arch Neurol Psychiatry*. – 1951. – Vol. 65. – P. 272-290.
6. Penfield W., Jasper H. *Epilepsy and the Functional Anatomy of the Human Brain* / W. Penfield, H. Jasper. – Boston: Little, Brown & Co, 1954.



577.12

**CTS-ATP** -

• „ • „ • •

E-mail: *alyona55567@gmail.com*

- - , -

cts-atp

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- ( - 4.1.3.8) - ,

- - ( )

[2]. , -

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[3].

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(-CH<sub>3</sub>)  
[1].  
(5-mC), 5-  
CG, CNG CNN.

- CpG.  
5'-CpG-3'.

CpNpG - , N -  
[4].

CpNpN -  
N -

: CpG

. CpNpG

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(Zea

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cts-atp

NCBI.

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Illumina NovaSeq6000.

CG-

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CG

14, CNG -5, CG CNG  
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CG,

CG, CNG

*cts-atp*

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CG	14
CNG	5
CNN	7

(TSS).  
CpG-, CNG CNN  
TSS

TSS CG-  
CpG- ( .  
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CG, CNG  
(TSS)

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CNN  
*cts-atp*

	TSS
CG	14
CNG	5
CNN	2

TSS

CpG-

CG,

1. / . . . . . // , : , , , . - 2022. - 2. - . 44-51
2. J.A. Watson et al. Tricarballylate and hydroxycitrate: substrate and inhibitor of ATP:citrate oxaloacetate lyase Arch. Biochem. Biophys. (1969)).
3. Lucibelli F. Plant DNA Methylation: An Epigenetic Mark in Development, Environmental Interactions, and Evolution / F. Lucibelli, M. S. Valoroso, S. Aceto / International Journal of Molecular Sciences. - 2022. - Vol. 23. - P. 8299.)
4. Cao X. Locus-specific control of asymmetric and CpNpG methylation by the DRM and CMT3 methyltransferase genes / X. Cao, S.E. Jacobsen // Proc Natl Acad Sci USA. - 2002. - V. 10, N. 99. - P. 16491-16498.



: 581.13.04

24-

*Pisum sativum (L),*

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-mail: *profershova@mail.ru*

(3-6 )  
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5- -2-  
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24-

220%.

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[1].

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(*Amanita muscaria*),

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+60° ,

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18-20%.20-26 %, 2<sup>-</sup> 30-32%

[4],

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2<sup>-</sup> 3 6 ( -1 - , %)

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3	-	3,80±0,15 100%	3,00±0,15 80%	3,12±0,16 82%
		2,56±0,07 100%	2,02±0,10 79%	1,87±0,07 73%
	- -	0,26±0,01 100%	0,17±0,01 65%	0,22±0,01 85%
6	-	3,65±0,10 100%	2,85±0,07 78%	2,90±0,08 80%
		2,41±0,02 100%	1,78±0,02 74%	1,65±0,08 68%
	- -	0,24±0,01 100%	0,23±0,01 96%	0,41±0,01 170%

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1 Hatcher C., Sommer U., Heaney L., Millett J. Metabolomic analysis reveals reliance on secondary plant metabolites to facilitate carnivory in the Cape sundew, *Drosera capensis* // *Ann. Bot.* 2021. V. 128. P. 301-308.

2 Zemlianukhin A.A., Ershova A.N. // *Metabolism of Isosuccinimide-p-Glucoside in Pea Seedlings* // *Biochemie und Physiologie der Pflanzen.* 1984. V. 179, N. 8, p. 679–684.

3 . . . . . *Pisum sativum* L. // : , . 2021. . 2. . 27-33.

4 Liu T.-Y., Castelfranco P. The biosynthesis of ethyl- -glucoside in extracts of pea seedlings // *Plant Physiol.* 1970. V. 45. P. 424.-426.

5 . . . . . , // : , , . 2024. 4. C.33-40.



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E-mail: *dowi2009@mail.ru*

( , 4.1.1.15) –

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( , L- -1- , 4.1.1.15,

glutamatic acid decarboxylase) – - ,

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( – TPC1, ) [5].

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(*Zea mays* L., 76).  
25 , 12-

25 / <sup>2</sup>.  
10<sup>-4</sup> [1].

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( 4,8), 1,5 , 1,5 , 0.1  
5 3000 g [7].

, 20 (pH 4,8), 70  
,10 -5- ,2 .  
620 3 [7].

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12.0.

±

(SEM).  
( $p < 0.05$ ).

[2].

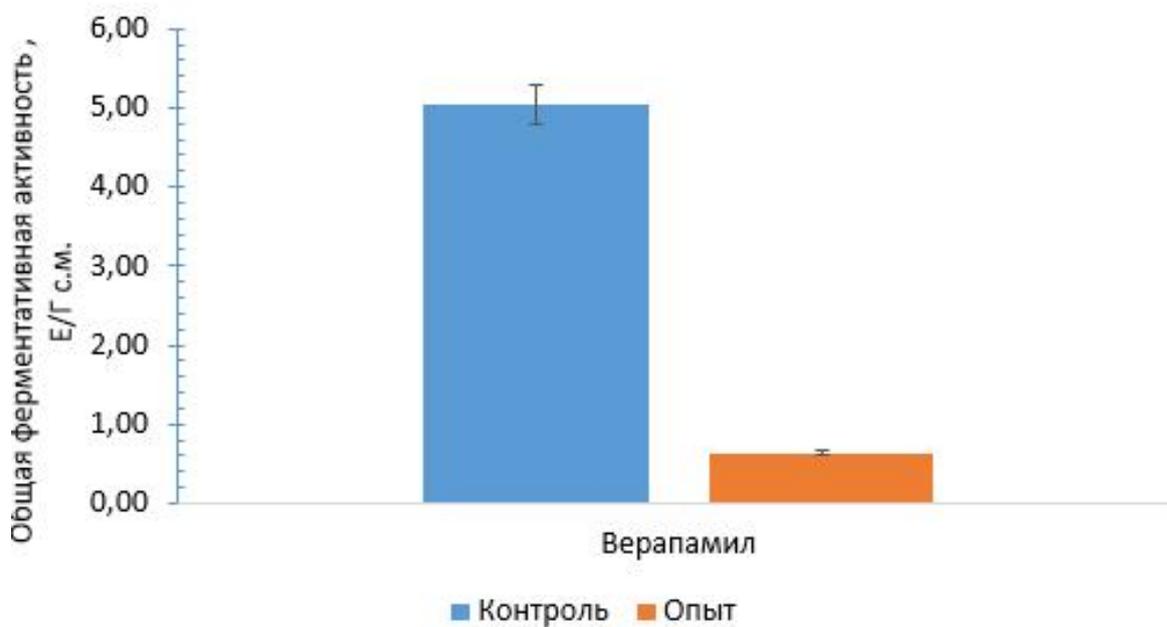
1959 ,

[6].

( .1).

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1. // . - 2020. - 3. - . 484.
2. . , . , . *GADI* . - 2024.
3. . 2. // : . . - 2001. - . 41. - . 131-162.
4. Edel KH, Marchadier E, Brownlee C, Kudla J, Hetherington AM. The evolution of calcium-based signalling in plants. *Current Biology*. 2017;27(13):667–679. DOI: 10.1016/j.cub.2017.05.020.
5. Genes for calcium-permeable channels in the plasma membrane of plant root cells / P.J. White [et al.] // *Biochim biophys acta*. – 2002. – Vol. 1564. – P. 299–309.
6. Konagalla, S. V. et al. (2025) “The psychological impact on American healthcare workers during the COVID-19 pandemic: an initial scoping review,” *Academia Mental Health and Well-Being. Academia.edu Journals*, 2(1). doi: 10.20935/MHealthWellB7570.
7. Yu K. et al. A high-throughput colorimetric assay to measure the activity of glutamate decarboxylase // *Enzyme and microbial technology*. – 2-11. – T.49.- . 3. – C. 272-276.



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-mail: *marina.gataullina@gmail.com*

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16 .

(Rattus norvegicus)

380

6 .

25%

[7]. 1-

5-

[8],

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[7].

« »( 3, 5, 12, 15- ),

65 ,

32 .

12 ,

12 .

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3 ,

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10

[6].

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Excel 2010.

: ( . 1).



380 . 3-  
: 330 . 12- 320 . 15-  
- 300 .

1. Brown R.E., Wong A.A. (2007). The neuroanatomy of the mouse: a fine-scale model for behavioral neuroscience. *Brain Research Bulletin*, 75(5), 712–715.
2. Seibenhener M. L., Wooten M.C. (2015). Use of the Open Field Maze to measure locomotor and anxiety-like behavior in mice. *J Vis Exp*, (96), e52434.
3. Spear L.P., Swartzwelder H.S. (2014). Relationship between ethanol-induced activity and anxiolysis in the open field test. *Neuropharmacology*, 85, 1–9.
4. Doremus-Fitzwater T. L. et al. (2022). Chronic alcohol alters anxiety-like responses in mice. *Alcohol Clin Exp Res*, 46(7), 1234–1245.
5. Kliethermes C. L. (2005). Anxiety-like behaviors after ethanol withdrawal in mice. *Alcohol*, 36(2), 83–90.
6. / . , . , . . . . – : . . . . , 1991. – 399.
7. / . . . . // . – 2020. – 2. – . 43-50.
8. [ . ]// / . . . . . – 2006. – 3. – . 36-41.



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• ” • ” • •  
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-mail: *anastasiya.knyazeva.03@mail.ru*

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[1].

[4].

( , 2.7.1.1)[6].

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, 4- -  
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-6- [3].

Zn<sup>2+</sup>

(*Rattus norvegicus* L. Wistar) 12  
 150-200 (n=12).  
 ( 25<sup>0</sup> ,  
 )  
 4  
 3 :« » - ,  
 0,9% NaCl, « » - ,  
 5% ( 100 / [11]); « + »  
 « + » - ,  
 « » («Woerwag Pharma GmbH &  
 Co. KG», )  
 , 36 / .  
 ,  
 .  
 (« » ).

31  
 [2].

0,9% NaCl.

Walker & Rao,1964 [11],

, : KCl (0,15 ); (5 ); MgCl<sub>2</sub> (5 ); -  
(10 ); - Cl 7,3.

-6- NADP<sup>+</sup>

-6-

, : - Cl (50 ) 7,5; MgCl<sub>2</sub> (7,5 ); (5  
); + (5 ); 2 -6- ; (100  
0,5 ).

0,5 - , - 100 -  
0,5 -

[11]

, 100 ,

, EC 1.1.1.47],

[fl-D- -NAD(P)  
Km

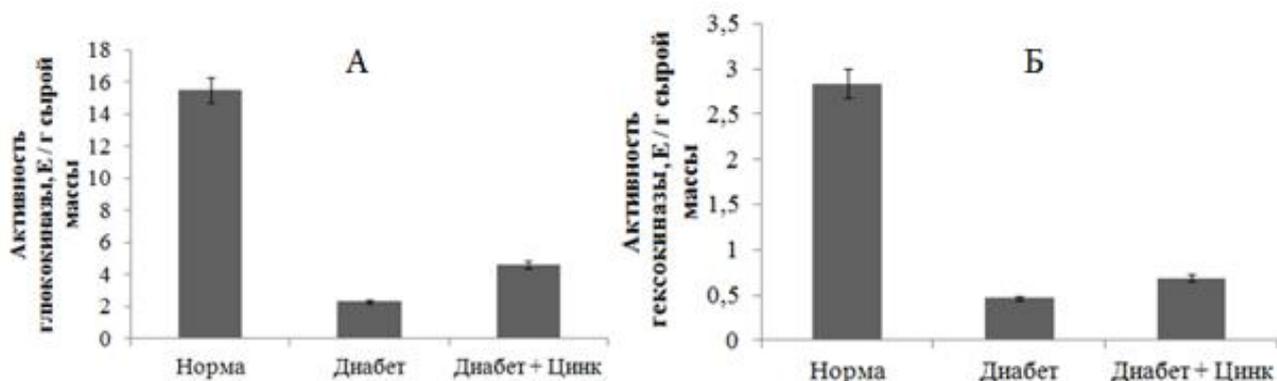
NADP<sup>+</sup> [7],

-6- [12]

15,72 \ , « »  
« + » - 2,08 \ ,  
« + » - 4,04 \ ( .1 ).  
« + » , ,

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– 2. – .71-92.

3. ... [ ] //
4. - 2016. - 19(3). - c. 190-198.
5. C rdenas, M. L. Evolution and regulatory role of the hexokinases / M. L. C rdenas, A. Cornish-Bowden; T. Ureta // Biochim. Biophys. Acta. -1998. - 1401 (3). – p. 242–264.
6. Leighton B. Small molecule glucokinase activators as novel anti-diabetic agents / Leighton B., Atkinson A., Coghlan M. P. – 2005.
7. Metzger, R. P. J. *biol. Chem.* / R. P. Metzger, S. S. Wilcox, A. N. Wick -1963. - 1769. – p. 239
8. Prediction and validation of the three-dimensional structure of glucokinase-1 from *Phytophthora infestans*. - 2021
9. Salas M. J. *biol. Chem.* / M. Salas, E. Vifiuela, A. Sols -1964. – 3535. – p. 238
10. Tang X. Zinc has an insulin-like effect on glucose transport mediated by phosphoinositol-3-kinase and Akt in 3T3-L1 fibroblasts and adipocytes. / X. Tang , N. F. Shay // *J. Nutr.* – 2001. – 131. -p. 1414-1420.
11. Walker D. G. *Abstr. Proc. 6th Int. Congr. Biochem.*, New York, -1964. p. 475.
12. Walker D. G. Department of Biochemistry, University of Birmingham / G. Walker, Holland Geraldine. - 1965.



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. .<sup>1,2</sup>, . .<sup>1</sup>, . .<sup>1,2</sup>, . .<sup>1</sup>

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E-mail: [koroleva\\_victoria@bk.ru](mailto:koroleva_victoria@bk.ru)

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( 3.4.22.3) – ,  
*Ficus*. 222 .

*Staphylococcus aureus*    *Staphylococcus epidermidis*

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[2], , .

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( ). , ,

[3].

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(Sigma Aldrich).

*N*-*-DL-**-p*- (BAPNA) (Sigma Aldrich).

60, 70 80 °C 10–60 , 20

25 000 g Jouan MR23i (Thermo Fisher Scientific, ).

(1–A/A<sub>0</sub>), A A<sub>0</sub> ( in)

60, 70 80°C 10–60

60, 70 80°C 10–60 , 20 25 000 g

..., 2025, 27.

Jouan MR23i (Thermo Fisher Scientific, ).

280

Shimadzu UV-2401PC (Shimadzu Scientific Instruments, ,

).

( $\alpha_{agg}$ )  $(1-D/D_0), D D_0$

Nano

Zetasizer ZS (Malvern Panalytical Ltd., ).

He/Ne-

632.8

4

173°

25–80°C.

Stadia 8.0 Professional.

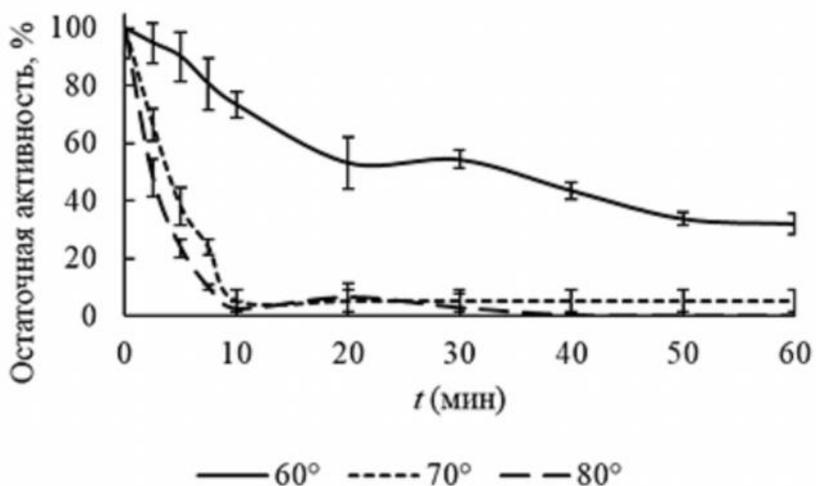
t- (  $p < 0,05$ ), ,

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60

60, 70 80°C

1



. 1.

60, 70 80°C

( 10 )

( 10 60 ).

( in)

25 000 g.

in 60°C,

70 80°C

( . 1).

1.

( in) %

Температура, °C	$\gamma_{in}$ без центрифугирования	$\gamma_{in}$ с центрифугированием
60	68±4	78±3
70	95±9	100±9
80	99±8	100±9

1 60°C

( . 2).

70°C

80°C

agg'

( ),

agg'

2.

(agg) %

Температура, °С	$\gamma_{agg}$ , метод ДРС	$\gamma_{agg}$ , метод спектрофотометрии
60	42±8	31±8
70	75±7	72±2
80	99±3	100±5

1. Baidamshina D.R., Trizna E.Y., Holyavka M.G., Bogachev M.I., Artyukhov V.G., Akhatova F.S., Rozhina E.V., Fakhrullin R.F., Kayumov A.R. Targeting microbial biofilms using Ficin, a nonspecific plant protease. *Scientific Reports*. 2017. 7(1).
2. Ol'shannikova S.S., Red'ko Y.A., Lavlinskaya M.S., Sorokin A.V., Holyavka M.G., Artyukhov V.G. Preparation of papain complexes with chitosan microparticles and evaluation of their stability using the enzyme activity level. *Pharmaceutical Chemistry Journal*. 2022. V. 55. No. 11. P. 1240-1244.
3. Li C., Wang K. Recent research and advances of material-based saturable absorber in mode-locked fiber laser. *Optics & Laser Technology*. 2021. V. 138. 106862.



( 4.2.1.3)

[1].

( 1.3.99.1)

[2].

II

BVRB\_4g088410

*Beta*

BVRB\_6g128670,  
*vulgaris* L.,

*B.vulgaris* L.,

NCBI (<https://www.ncbi.nlm.nih.gov/gene/>).

Primer-BLAST

(<https://www.ncbi.nlm.nih.gov/tools/primer-blast/>).

BLAST online

(<https://blast.ncbi.nlm.nih.gov/Blast.cgi>).

Oligo calculators (<https://>

[www.thermofisher.com/ru/ru/home/life-science/oligonucleotides-primers-probes-genes/oligos-tools-utilities.html](http://www.thermofisher.com/ru/ru/home/life-science/oligonucleotides-primers-probes-genes/oligos-tools-utilities.html)).

[3].

*BVRB\_4g088410 BVRB\_6g128670*

Gen Bank.

Primer-BLAST.

BLAST online

100%

*Beta vulgaris L.*

Oligo calculators.

1.

***BVRB\_4g088410 BVRB\_6g128670 Beta vulgaris L.***

Ген	Последовательность	T плавления	GC, %	Размер продукта
<i>BVRB_6g128670</i>	f 5'- acattgcacagattccgctc -3'	58,91	50,00	256
	r 5'- ccataaaacgctcaccctcg-3'	58,99	55,00	
<i>BVRB_4g088410</i>	f 5'- atcccatttaagcctgctcg -3'	58.03	50.00	206
	r 5'- tccatattcgctgaactgc -3'	58.05	50.00	

*BVRB\_4g088410*

(Beta

*BVRB\_6g128670*,  
*vulgaris* L.).

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1. ... / ... // ... - 2020. -  
. 1. - . 35-41.
2. ... / ... [ ... ] //  
. - 2020. - 3. - C. 146.
3. ... // ... - 2019. - . 11. - . 1. - . 23-70.



579.62

**BACILLUS SUBTILIS**

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. .1, . .1,2

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E-mail: *mihan.vrn@mail.ru*

, 14- *Firmicutes* 30-

, *Bacillus subtilis.* 30-

14- *Firmicutes* 1,4 30-  
*Bacillus subtilis,*  
Bacteroidetes 1,7

14-

[1].

..., 2025, 27.

[2]. 2006 ,

*Bacillus subtilis*,

[3].

[4].

*Bacillus*

*subtilis*

[5].

(14 30 )

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«Bio-Rad CFX96TM Real-Time System»

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; - 0,5 ; 5X qPCRmix-HS SYBR ( , )

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95°C - 2 ; 39 :

95°

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51°

30 ;

72°

45 ;

72°

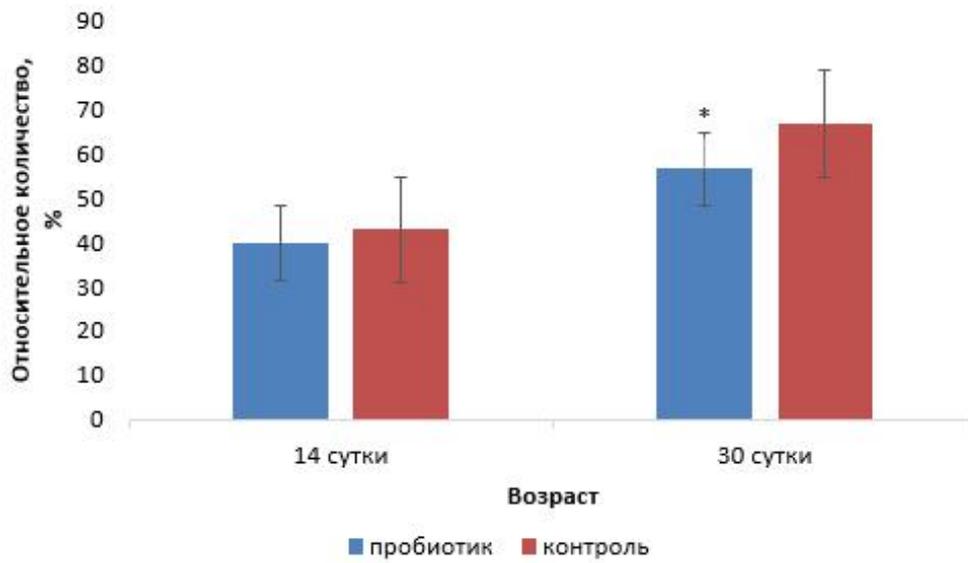
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Yun-Wen

Yang, 2015.

1

*Firmicutes*.



. 1.

14

*Firmicutes*. \* - (p<0,05).

*Firmicutes*

*Bacillus subtilis*. *Firmicutes* –

*Firmicutes*

: Bacilli, Clostridia, Erysipelotrichi Negativicutes.

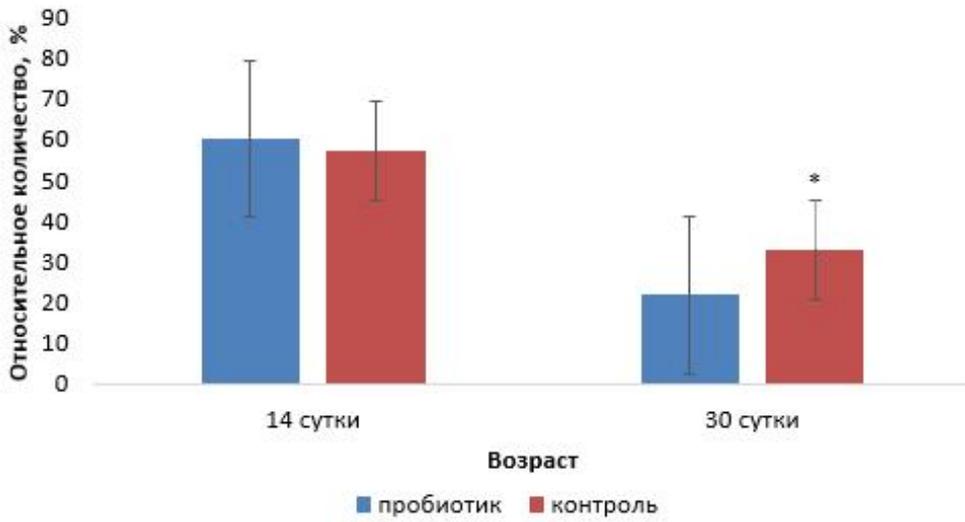
2

Bacteroidetes. 30-

*Bacillus subtilis*,

Bacteroidetes 1,7

14-



.2.

14

Bacteroidetes. \* - (p<0,05).

Bacteroidetes

Saccharibacteria, Verrucomicrobia Deferribacteres

Bacteroidetes Firmicutes

30-

Firmicutes 1,4

14-

30-

*Bacillus subtilis*,

Bacteroidetes 1,7

14-

*Bacillus subtilis*

1. . . , . . . // . - 2020. - 8. - . 21-26 ,
2. . . // . - 2019. - 3. - . 17-20.
3. . . , . . . // *Bacillus subtilis* . - 2021. - . 7,
2. - . 45-50.
4. . . //
5. . - 2018. - 6. - . 33-36.
5. . . , . . . // . - 2022. - . 38, 4.
- . 65-72.



582.475,4:631.671,3

1,2 . . , 1 . . , 1 . .

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E-mail: *mashkinaos@mail.ru*

2014) . (2008, 2011, 2015) ( 2010 2012 .) (2007, 2010, 2012, 2012 .)

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1,7 ,

[1, 2].

(*Pinus sylvestris* L.)

[3, 4].

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[6],

[7, 8].

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[11, 12].

9187 21.07.2017,

[11, 12].

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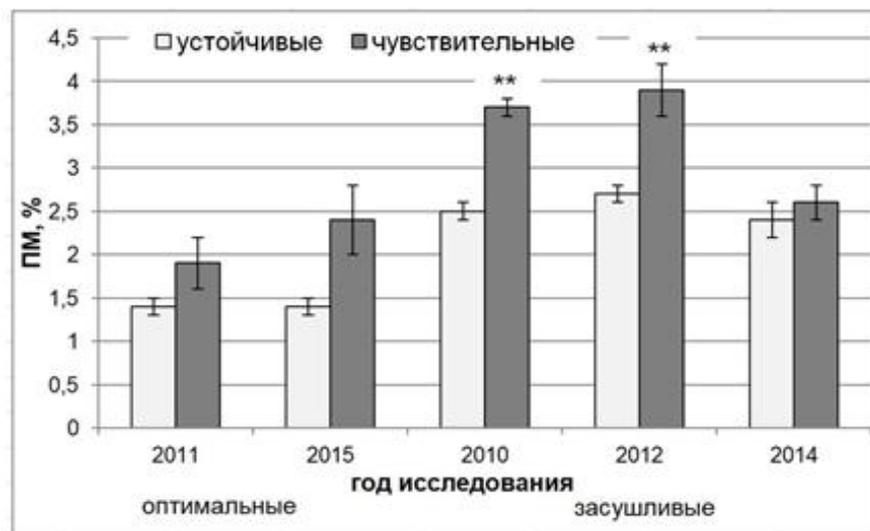
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[16, 17].

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\*\*p<0,01

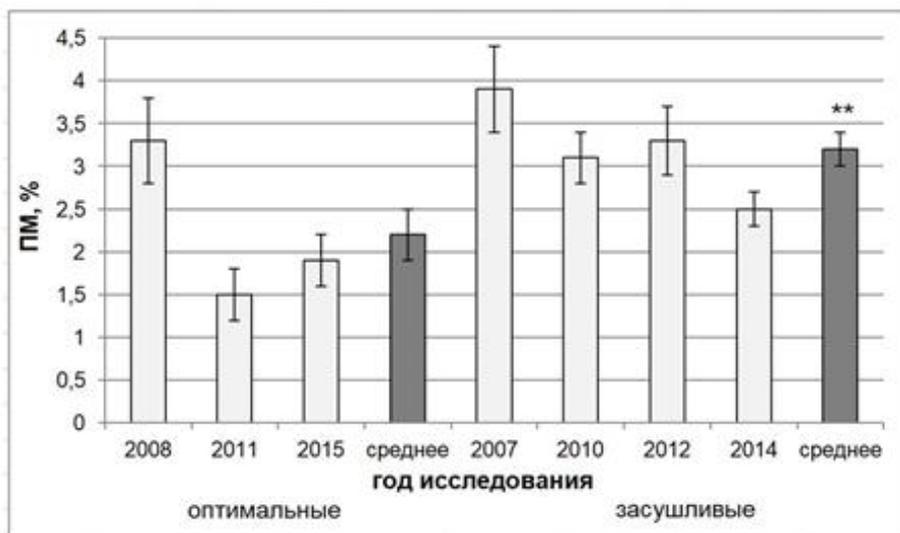
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2012 . 6 -5,1%, 3,9 ±  
0,3% . , (2010  
2012 .) 1,5

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( . 2),

2011 –  $1,5 \pm 0,3\%$ , (  $3,9 \pm 0,5\%$ ) –  
 2007 . ,



\*\*p<0,01

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[7, 8, 11, 19].

( 1, 6, 9)

(2010, 2012 .)

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0,05–0,1% 0,02–0,05% ( . 3).

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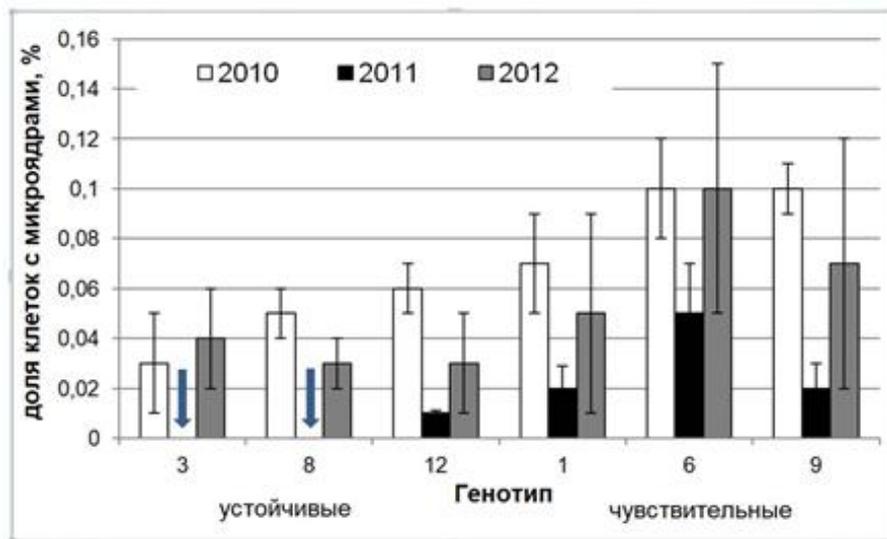
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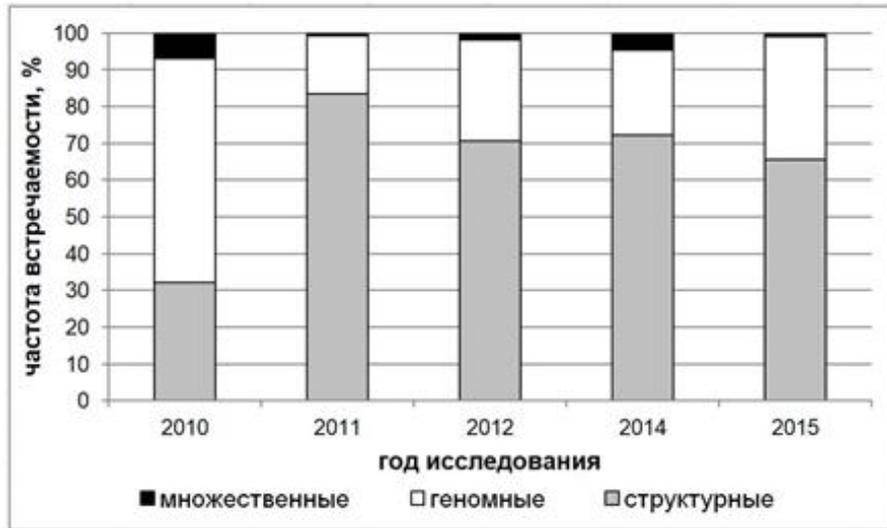
(2010, 2012 2014 .)

2011

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6-9.

2010 – 32,2 %, 2011 – 2,5 (83,4 %). : 2012 . – 70,5 %, 2014 . – 72,3 %, 2015 . – 65,6 %.



- 4. (2011, 2015) (2010, 2012, 2014)

[21–23]

(2007) (2008, 2011, 2015) (2007, 2010, 2012, 2014)



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7. // . – 2000. – 3. – . 206–210.
8. // . – 2023. – . 30,  
5. – . 591–602.
9. //  
– 2011. – . 13, 5–2. – . 179–183.
10. //  
– 2013. – 2. – . 16–21.
11. Pardayeva E.Y., Mashkina O.S., Popov V.N. State of *Pinus sylvestris* L. generative sphere according to cytogenetic analysis in changing climate conditions on the territory of Voronezh oblast // Contemporary Problems of Ecology. – 2017. – Vol. 10, 3. – . 271–276.
12. // . – 2020. – 85.  
– . 104–109.
13. // *Pinus sylvestris* L.  
– 2021. – 91. – . 181–186.
14. *Pinus sylvestris* L.  
// . – 2015. – 5. – . 332–338.
15. // . – 1975. – . 22. – . 107–114.
16. // . – 1965. – 11. – . 58–66.
17. / [ ] // :  
– 2007. – 4. – . 508–512.
18. : . . . . , 2011. 40 .
19. //  
– 1988. – . 22, 1. – . 67–72.
20. ( *Pinus sylvestris* L.)  
// . – 2004. – . 2. – . 128–140.
21. *Pinus sylvestris* (*Pinaceae*)  
// . – 2021. – T. 106,  
4. – . 353–362
22. . I.  
// . – 1967. – . 3, 5. – . 45–51.
23. // . –  
1983. – . 23, 5. – . 703–706.



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-mail: *nakvasina\_ma@mail.ru*

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[1-5].

1510 / 2)

(10<sup>-5</sup> / ) [6-8].

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1.

Исследуемый образец	Люминесцентные характеристики зонда	
	$\lambda_{\text{max}}$ флуоресценции, нм	Интенсивность флуоресценции в максимуме, отн. ед.
Интактные мембраны	478	11,7±0,8
Мембраны + ресвератрол	484	14,7±2,5
Мембраны + ЦА	484	13,6±2,6

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max

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Исследуемый образец	Люминесцентные характеристики зонда	
	$\lambda_{\text{max}}$ флуоресценции, нм	Интенсивность флуоресценции в максимуме, отн. ед.
Липосомы из ФХ	479	234±5
Липосомы из ФХ + ресвератрол	481	188±12
Липосомы из ФХ + ЦА	477-478	84±6
Липосомы из ФХ и ХЛ	484	65±8
Липосомы из ФХ и ХЛ + ресвератрол	480-481	79±6
Липосомы из ФХ и ХЛ + ЦА	476	84±3
Водный раствор ресвератрола	503	11±2
Водный раствор ЦА	500	9±1

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335 ( = 774 / . )

[7].

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(1.0·10<sup>-5</sup> / )

1.0·10<sup>-4</sup> /

1:10.

2

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0,23

-1 c

130

240-390

151 / 2,

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[8].

S-

HbO<sub>2</sub>

$$= 21,57 \pm 0,78$$

$$a = 2,36 \pm 0,35.$$

40 . . (Y<sub>40</sub>)

84,35 ± 0,98 %,

100 . . - 97,13 ± 0,87 %,

HbO<sub>2</sub> ( ) 12,32 ± 0,78 % ( 1).

1.

**GSNO**

УСЛОВИЯ ОПЫТОВ P <sub>O2</sub> , ММ рт. ст.	HbO <sub>2</sub>	HbO <sub>2</sub> +УФ	Hb-GSNO (1:10)	Hb-GSNO (1:10) + УФ
0	0	0	0	0
3,19	2,56 ± 0,97	4,94 ± 1,42 *	1,33 ± 0,84	3,24 ± 1,15
7,98	7,68 ± 2,36	12,07 ± 3,22 *	4,17 ± 0,91 *	8,81 ± 1,73 **
15,96	22,42 ± 3,93	55,71 ± 3,47 *	17,63 ± 0,25 *	27,16 ± 0,58 **
23,94	62,50 ± 3,45	78,63 ± 2,89 *	68,31 ± 0,93*	69,4 ± 0,64 **
31,92	81,90 ± 2,39	88,80 ± 1,72 *	78,67 ± 0,69	85,59 ± 1,33 **
63,84	93,98 ± 0,81	96,04 ± 0,39 *	92,12 ± 0,95	96,13 ± 0,82
95,76	97,61 ± 0,43	98,04 ± 0,36	96,53 ± 0,32	97,47 ± 0,39
159,6	100	100	100	100

: \*  
( <0,05), \*\*

( <0,05).

S -

$1 \cdot 10^{-4}$  /

$Y_{40} \quad 88,53 \pm 0,62 \%$ ,

$8,92 \pm 0,61 \%$ .

Y100

50,

S -

$(1 \cdot 10^{-4} \quad /)$

151 / <sup>2</sup>

$50 ( \quad 19,35 \pm 0,83 \quad .$   
 $, Y_{40}, Y_{100}$

%  $9,96 \pm 0,58 \%$

$2,35 \pm 0,31, 89,33 \pm 0,78 \%, 98,87 \pm 0,36$

1:10,

S-

, S-

1. ... : ... / ... . - : -  
... , 1995. - 280 .
2. ... : -  
/ ... , ... , ... ,  
... , ... . - : - , 1997. - 281 .
3. ... //  
/ ... , ... , ... //  
1993. - . 113 4. - . 456-470.
4. Anggard E. Nitric oxide: mediator, murderer and medicine / E. Anggard // Lancet.  
- 1994. - Vol.343. - P. 1199-1206.
5. Verigo B. F. Zur Frage ber die Wirkung des Sauerstoff auf die  
Kohlens ureausscheidung in den Lungen / B. F. Verigo // Archiv f r die gesammte  
Physiologie des Menschen und der Thiere - 1892. - 51. - . 321-361.
6. Bohr Chr. Concerning a Biologically Important Relationship — The Influence of  
the Carbon Dioxide Content of Blood on its Oxygen Binding / Chr. Bohr, K.  
Hasselbalch, A. Krogh. // Skand. Arch. Physiol. - 1904. - 16. - . 401-412.
7. ... S-  
S- / ... // ... - 1995. - .  
60, . 1. - . 593-602.
8. ... -  
... ,  
/ ... , ... , ... ,  
... // ... - 2016. - . 62. -N 3. - . 251-258.



577.151.6

-[N6- ]-

• ” • ” • ” • •

E-mail: *daryagosudarya@gmail.com*

-[N6- ]-

(Dam),

Dam

[3].

NaCl

[4,5,6,7,8].

[9].

[10,11].

[12].

in vivo in vitro [13].

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 , c  
 ( ) ,  
 , ( ) [14].  
 - (Dam) — ,  
 5'-GATC-3' [15]. Dam  
 [16]. *E.*  
*coli* GATC ,  
 . , Pap-  
 ( - , P- ), *E.*  
*coli* Dam GATC  
*pap* [17]. ,

*dam* *Aggregatibacter actinomycetemcomitans*

,  
 [18].  
*dam*  
 [19]. ,  
 -[N6- ]- ,

14-  
 (*Triticum aestivum* L.) -76.  
 25° C.  
 , 150 NaCl.  
 1, 3, 6, 9, 12 24 .

Dam

256

Dam

3

[1].

1

1

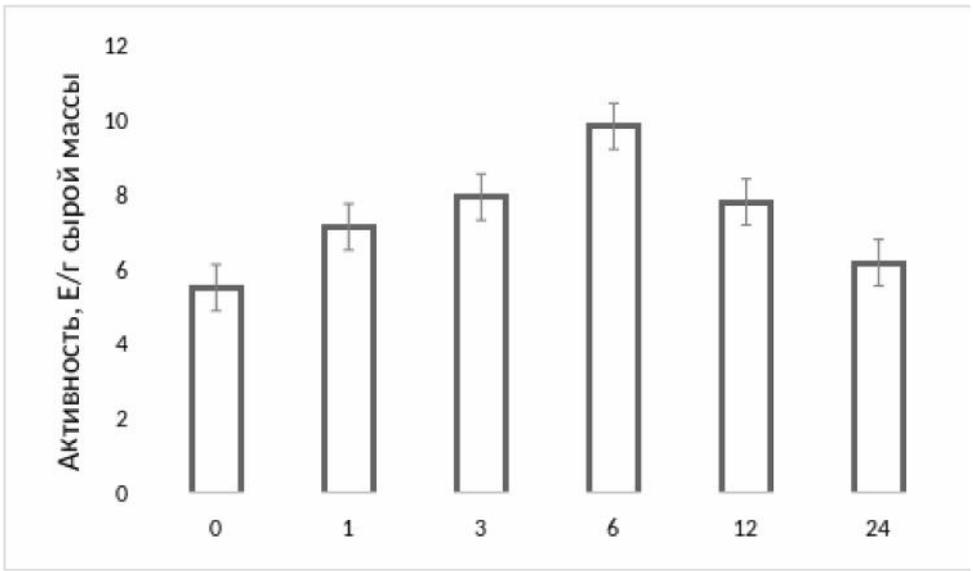
25° .

Dam

( . 1)

6-

[2].



. 1.

Dam

150

NaCl.

Dam

Dam

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[2].

6

Dam.

Dam

Dam

Dam

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NaCl

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Dam

1. ... .. // : - /  
... .. - 1996.
2. ... .. // / ... ..  
... .. :  
... .. 25. - :  
... .. , 2023. - . 43-48.
3. Abiotic stress responses in plants / Zhang H. [et al.] // Nature Reviews Genetics. – 2022. – . 23. – . 2. – P. 104-119.
4. Datir S. Effect of NaCl-induced salinity stress on growth, osmolytes and enzyme activities in wheat genotypes / Datir S., Singh N., Joshi I. // Bulletin of environmental contamination and toxicology. – 2020. – . 104. – P. 351-357.

5. Hajiyeva I. Effect of High Salt Stress on Germination and Growth of Some Varieties of Common Beet / Hajiyeva I. // ... – 2024. – . 10. – . 5. – P. 188-195.
6. Effect of salt stress on growth, physiological and biochemical parameters and activities of antioxidative enzymes of rice cultivars / Talubaghi M. J. [et al.] // Cereal Research Communications. – 2023. – . 51. – . 2. – P. 403-411.
7. Li W. Effect of environmental salt stress on plants and the molecular mechanism of salt stress tolerance / Li W., Li Q. // Int. J. Environ. Sci. Nat. Res. – 2017. – . 7. – . 3. – P. 555714.
8. Maize (*Zea mays* L.) responses to salt stress in terms of root anatomy, respiration and antioxidative enzyme activity / Hu D. [et al.] // BMC plant biology. – 2022. – . 22. – . 1. – P. 602.
9. DNA methylation in plant responses and adaption to abiotic stresses / Sun M. [et al.] // International Journal of Molecular Sciences. – 2022. – . 23. – . 13. – P. 6910.
10. DNA cytosine methylation dynamics and functional roles in horticultural crops / Liu P. [et al.] // Horticulture Research. – 2023. – . 10. – . 10. – P. uhad170.
11. Vanyushin B. F. DNA methylation in plants / Vanyushin B. F. // DNA methylation: basic mechanisms. – 2006. – P. 67-122.
12. Chomet P. S. Cytosine methylation in gene-silencing mechanisms / Chomet P. S. // Current opinion in cell biology. – 1991. – . 3. – . 3. – P. 438-443.
13. Angeloni A. Enhancer DNA methylation: implications for gene regulation / Angeloni A., Bogdanovic O. // Essays in biochemistry. – 2019. – . 63. – . 6. – P. 707-715.
14. DNA methylation and DNA methyltransferases / Edwards J. R. [et al.] // Epigenetics & chromatin. – 2017. – . 10. – P. 1-10.
15. Vanyushin B. F. Adenine methylation in eukaryotic DNA / Vanyushin B. F. // Molecular Biology. – 2005. – . 39. – P. 473-481.
16. Same modification, different location: The mythical role of N 6-adenine methylation in plant genomes / Jim nez-Ram rez I. A. [et al.] // Planta. – 2022. – . 256. – . 1. – P. 9.
17. A methylation-directed, synthetic pap switch based on self-complementary regulatory DNA reconstituted in an all *E. coli* cell-free expression system / Worst E. G. [et al.] // ACS Synthetic Biology. – 2021. – . 10. – . 10. – P. 2725-2739.
18. Inactivation of DNA adenine methyltransferase alters virulence factors in *Actinobacillus actinomycetemcomitans* / Wu H. [et al.] // Oral microbiology and immunology. – 2006. – . 21. – . 4. – P. 238-244.
19. Vanyushin B. F. DNA methylation in higher plants: past, present and future / Vanyushin B. F., Ashapkin V. V. // Biochimica et Biophysica Acta (BBA)-Gene Regulatory Mechanisms. – 2011. – . 1809. – . 8. – P. 360-368.



581.132.1:58.032:674.031.632.264

**QUERCUS L.**

• • „ • •

« — »,

E-mail: [runastep@yandex.ru](mailto:runastep@yandex.ru)

*Quercus pubescens* Wild. *Q. petraea* (Matt.) Liebl.,<sup>b</sup>

*pubescens* ( ), *Q. petraea*<sup>Q.</sup>

350 2022  
*Q. pubescens*, *Q. petraea*,

[1, 2].

*Quercus*,

[3, 4].

..., 2025, 27.

[5].

[6, 7].

b

*Quercus L.*,

*Quercus pubescens Wild.*,

( , , )

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350

( . ),

*Q. petraea (Matt.) Liebl. –*

30

350

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2022-2024 .

b

Unico 2100

[8].

2022-2024 .

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MS Excel 2007.

2022-2024 .

b

*Quercus pubescens Q. petraea.*

..., 2025,

27.

2022 .

*pubescens*

( .1).

*petraea*

30

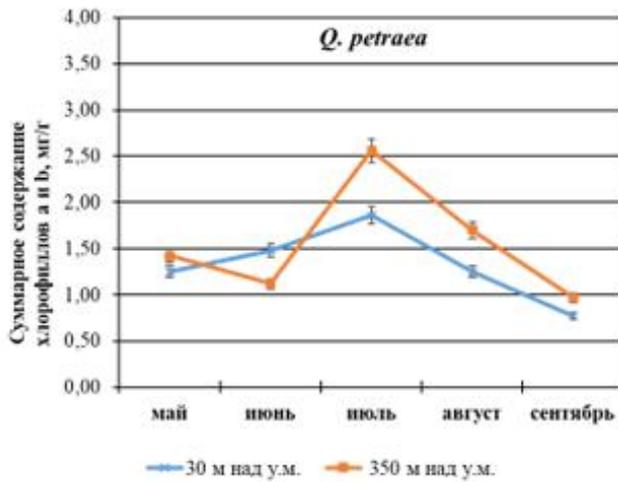
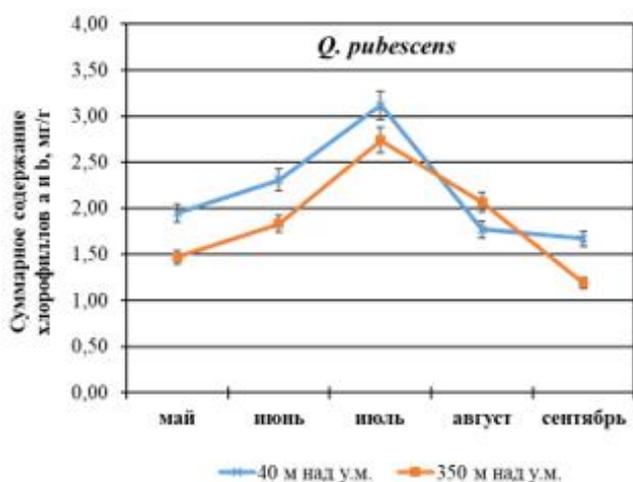
350

(2,5-2,6 /

( - ),

1,6 /

).



.1.  
*Q. petraea*

b *Q. pubescens*,  
2022 .

2023 .

*Quercus*

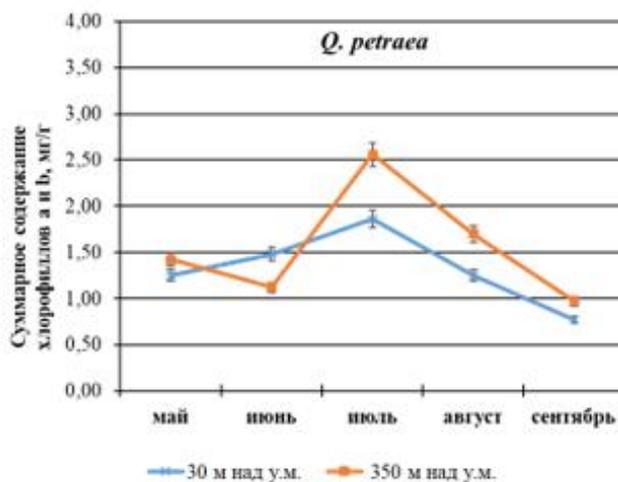
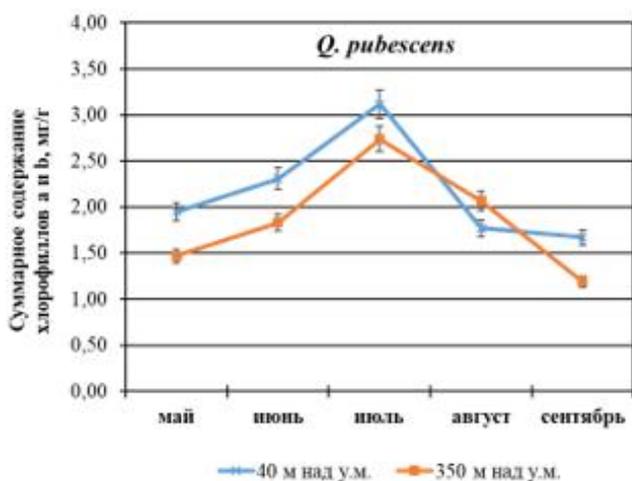
( .2).

2023 .

*Q. pubescens*

(1,19-3,12 /

), *Q. petraea* (0,77-2,56 / ).



.2.  
*Q. petraea*

b *Q. pubescens*,  
2023 .

2024 .

(  
2,7° ),

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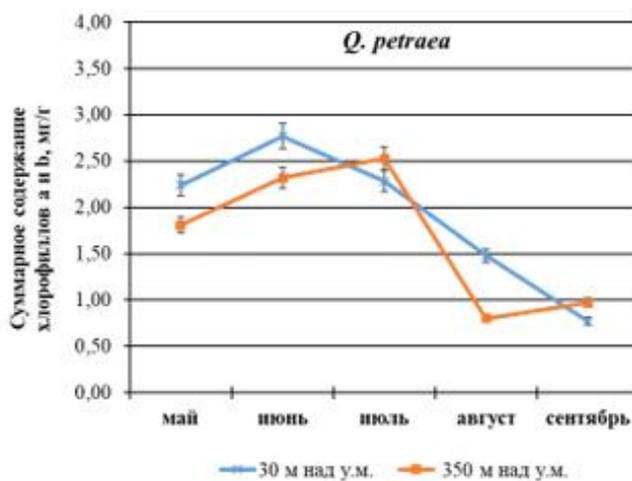
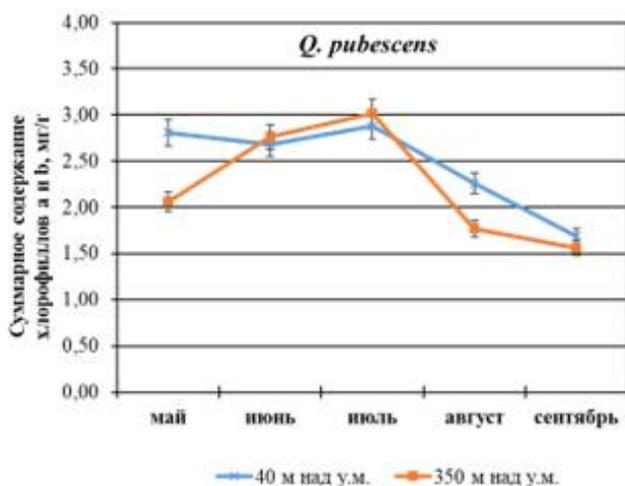
2024 . *Q. pubescens*,

40 . .

b

30%

350 ( .3).



.3.  
*Q. petraea*

b *Q. pubescens*,  
2024 .

*Q. petraea* 30

142

350

2024 .

( 2023 .) –

2022-2024 .

b

*Quercus pubescens* *Q. petraea*

*Q. pubescens*

( ),

*Q. petraea*

( - ).

2022 .

*Q. pubescens* *Q. petraea*,

350

b

*Quercus pubescens* *Q. petraea*.

1.

/[ .

. . . ]. – . : , 2017. – 106 .

2.

/ . . . , . . . , . . . , . . . , . . . //  
. . . . . – 2022. –

604. – . 55–201.

3. Cuza P., Dascaluic Al., Aproxima ia sistemic n utilizarea ra ional a speciilor i genotipurilor de stejar la mp durirea i gospod rirea durabil a p durilor din Republica Moldova, // Mediul Ambient. – 2015. - No. 3 (81).- . 7–15.

4.

//

. : . – 2011. - 2.  
– . 166–175.

5. Esteban R, Barrutia O, Artetxe U, Fern ndez-Mar n B, Hern ndez A, Garc a-Plazaola JI. Internal and external factors affecting photosynthetic pigment composition

in plants: a meta-analytical approach // *New Phytol.* – 2015. – V. 206(1). – P. 268-280.

6. . . . .

//

– 2020. – 67. – . 278-288.

7. Garcia-Plazaola J., Becerril J.M. Seasonal changes in photosynthetic pigments and antioxidants in beech (*Fagus sylvatica*) in a Mediterranean climate: implications for tree decline diagnosis // *Aust. J. Plant Physiol.* – 2001. – V. 28. – P. 225.

8. . . . .  
» , 1975. – 392 .



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[1].

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1)

1-3  
Smith et al. (2015) [2]

30

( , , - 65%)

3,2% 3,5%.

2)

0,5-2

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10-15%

, - 60%)

(Johnson,

2018) [3].

1.

[4].

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»,  
..., 2025, 27.



« » , 18%,  
- 12%. « - »  
4 / [6].

2.

3,5%,

3,2%

: — 0,2-0,3%,  
— 0,1-0,2%.

[8].

, 44

NovaPro (

2,5

) 3,0

NovaPro

1,7

[9].

[10].

[11].

Kemin,

(N)

(N, O)

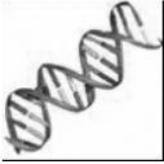
[12].

0,5-2 / 1-3 / 50-80%, (2020) [4]

1. ...
2. Smith, J. Brown, R., & Miller, T. Effects of heat-treated soybean meal on milk protein content in high-producing dairy cows / J. Smith, R. Brown, T. Miller // Journal of Dairy Science. – 2015. – Vol. 98. – P. 1234–1240.
3. Johnson, K. Impact of encapsulated sunflower meal on growth performance of dairy calves // Animal Feed Science and Technology. 2018. Vol. 245. P. 56–63.
4. ... // . — 2020. — . 45–52.
5. « » // . URL: [https://belkoff.biz/page\\_20.php](https://belkoff.biz/page_20.php) ( : 04.04.2025).
6. « » // . 2009. URL: <https://www.agroinvestor.ru/technologies/article/10974-korov-kormyat-zashchishchennym-belkom/> ( : 04.04.2025).
7. « » // . 2019. URL: <https://vitasol.ru/notes/belok-v-moloke-korov> ( : 04.04.2025).

..., 2025, 27.

8. Satter, L. D. Protected Proteins and Amino Acids in Dairy Cattle Nutrition / L. D. Satter, A. J. Roffler // Journal of Dairy Science. — 1986. — Vol. 69, No. 10. — . 2734–2744.
9. « » // Agrovesti.net. 2019. URL: <https://agrovesti.net/lib/tech/feeding-tech/menshe-soevogo-shrota-v-ratsionekorov.html> ( : 04.04.2025).
10. Santos, J. E. P. Nutritional Strategies to Improve Nitrogen Efficiency and Milk Production in Dairy Cows / J. E. P. Santos, J. T. Huber // Veterinary Clinics: Food Animal Practice. — 2005. — Vol. 21, No. 2. — . 347–368.
11. « » [ . .] // . URL: <https://kampkazan.ru/catalog/kormlenie/zashhishhennyj-belok/poliamin/> ( : 04.04.2025).
12. Kemin. « » // Kemin. 16.07.2021. URL: <https://info.kemin.com/ru/ruminants/blog/sustainable-economic-implications-amino-acids> ( : 04.04.2025).



: 577.218

**ZEA MAYS L.**

• ” • ” • ” • •

E-mail: *kate\_plotnikova36@mail.ru*

**GLYR2**

1.1.1.79) [1].

in vivo

[2].

[3].

[4].

[5].

*GLYR2*

GenBank (<https://www.ncbi.nlm.nih.gov/genbank/>).

GenBank (<https://www.ncbi.nlm.nih.gov/genbank/>).

Primer-BLAST,

(NCBI) (<https://www.ncbi.nlm.nih.gov/tools/primer-blast/>).

Multiple Primer Analyzer.

10-

*Zea mays* L.

76.

«LabTech» ( )

: 10-

25 / 2

+25 C.

R-Plants (Biolabmix,

)

1%

(Sigma Aldrich, )

0,5 /

2%

[6].

..., 2025, 27.

LV-RT Kit ( , ).

Random(dN)10 ( ,

).

MiniAmp (Thermo Scientific, )

ScreenMix ( , ).

( ) 2%

(*GLYR2*, LOC100280775),

8

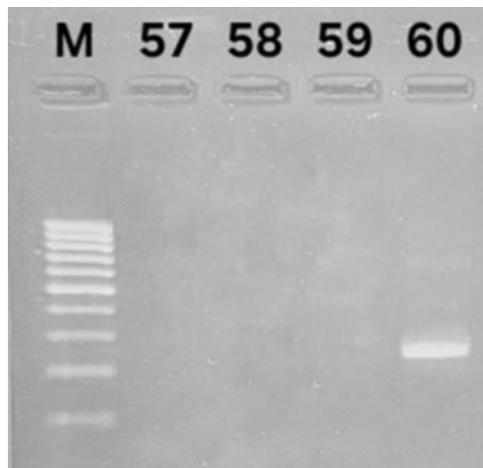
*GLYR2*

( .1)

1.

*GLYR2*,

Ген	Праймер	Последовательность	Размер продукта
<i>GLYR2</i>	Прямой	GCATTGCTGTCAGTAGTGGT	242
	Обратный	GAGTTCTGGATCAAAAGGTCGG	



. 1.

*GLYR2*

, 57-60 –

1, , 60 .  
 , - 250 ,  
*GLYR2*

(*GLYR2*).

250 ,  
 ,  
*GLYR2*

1. Allan W. L., Clark S. M., Hoover G. J., Shelp B. J. Role of plant glyoxylate reductases during stress: a hypothesis //Biochemical Journal. – 2009. – . 423. – . 1. – . 15-22.
2. Allan W. L., Breitzkreuz K. E., Waller J. C., Simpson J. P., Hoover G. J. Detoxification of succinate semialdehyde in Arabidopsis glyoxylate reductase and NAD kinase mutants subjected to submergence stress //Botany. – 2012. – . 90. – . 1. – . 51-61.
3. Yu L., Jiang J., Zhang C., Jiang L., Ye N., Lu Y. Glyoxylate rather than ascorbate is an efficient precursor for oxalate biosynthesis in rice //Journal of Experimental Botany. – 2010. – . 61. – . 6. – . 1625-1634.
4. Geigenberger P., Fernie A. R. Metabolic control of redox and redox control of metabolism in plants //Antioxidants & redox signaling. – 2014. – . 21. – . 9. – . 1389-1421.
5. Allan W. L., Simpson J. P., Clark S. M., Shelp B. J. -Hydroxybutyrate accumulation in Arabidopsis and tobacco plants is a general response to abiotic stress: putative regulation by redox balance and glyoxylate reductase isoforms //Journal of Experimental Botany. – 2008. – . 59. – . 9. – . 2555-2564.
6. / . .  
 2008. – 63 .  
 ..., 2025, 27.



636.2.084.412

• • • • •

« » ,

’ 2,7% ,

’ , [1].

’ , [2, 3].

... ..  
 . 2-4  
 8-10% ,  
 10-30% 10-25% [4, 5].

[6, 7].

t-

c

2,7%;  
 1,4%  
 1,2%;  
 : 0,85%, 0,75%,  
 0,95%.

1. ... / ... , 2018. – 260 ...
2. ... [ ... ]. – ... , 2017. – 413 ...
3. Chojnacka, K. Innovative high digestibility protein feed materials reducing environmental impact through improved nitrogen-use efficiency in sustainable agriculture / K. Chojnacka, K. Mikula, G. Izydorczyk [et al.] // Journal of Environmental Management. – 2021. – Vol. 291. – Iss. 2. – Article number: 112693.
4. // ... - 2011. - .191-206.
5. // ... - 2007. - 11. - . 22-31
6. ... « ... ».- 2011. - 372 ...
7. ... / ... - 2012. - 328 c.



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0,2%,

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1. URL: <https://vetandlife.ru/livestock/kak-kormlenie-vliyaet-na-vozproizvodstvo-korov/> (10.05.2025)
2. Rodec. Overcoming Negative Energy Balance in Dairy Cows. URL: <https://rodec.in/blog/overcoming-negative-energy-balance-in-dairy-cows> (10.05.2025)
3. [ ]. – , 2017. – 413 . / . .
4. // . – 2006. – 1-16.
5. Bewley, J.M. An Interdisciplinary Review of Body Condition Scoring for Dairy Cattle / J.M. Bewley, M.M. Schutz // The Professional Animal Scientist. – 2008. – 24:50.



631.81

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», .

E-mail:*elen\_yushkova@mail.ru*

(*Petroselinum crispum*),

0,01%.

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0,001

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[1, 2].

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0,01 0,001 %.

[3, 4].

4-

[5, 6].

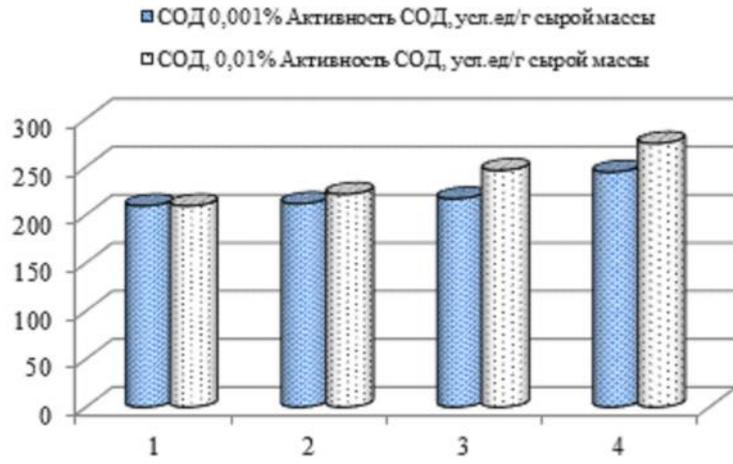
1

(*Petroselinum crispum*).

(1-3 ) 209,15 . / ( ) .

0,001% - 244,28 .

/ , 0,01% - 274,23 . / .

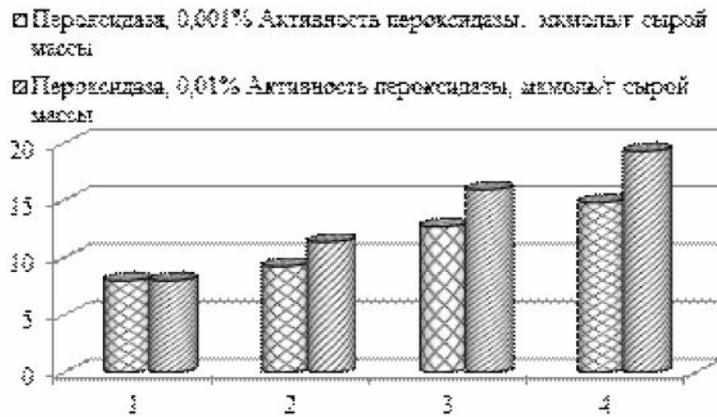


1. (*Petroselinum crispum*), / . ( )

2

(*Petroselinum crispum*),

0,001% - 17,28 0,01% -19,25 / .

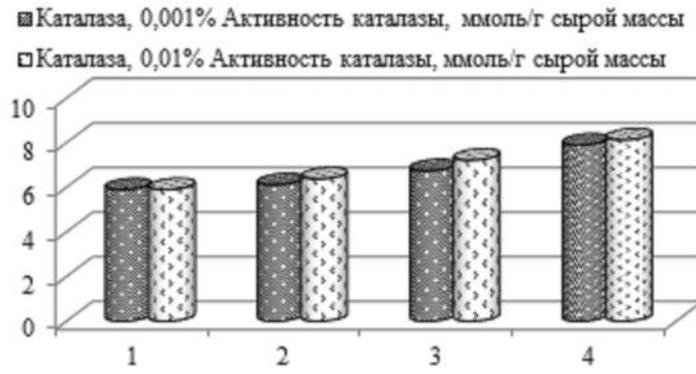


2. (*Petroselinum crispum*), / . (Petroselinum

[7, 8, 9].

..., 2025, 27.

(*Petroselinum crispum*).



.3.  
/

(*Petroselinum crispum*),

0,001%

8,12

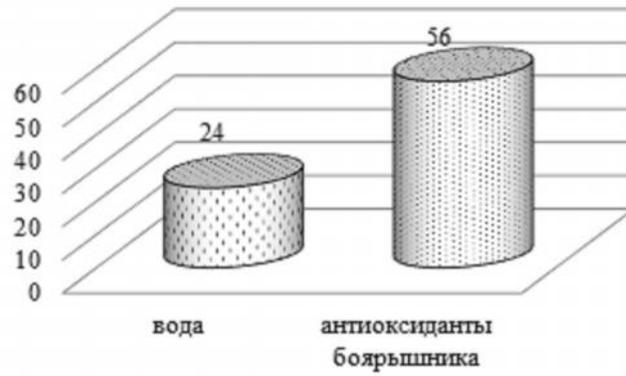
7,89

0,001%

[10].

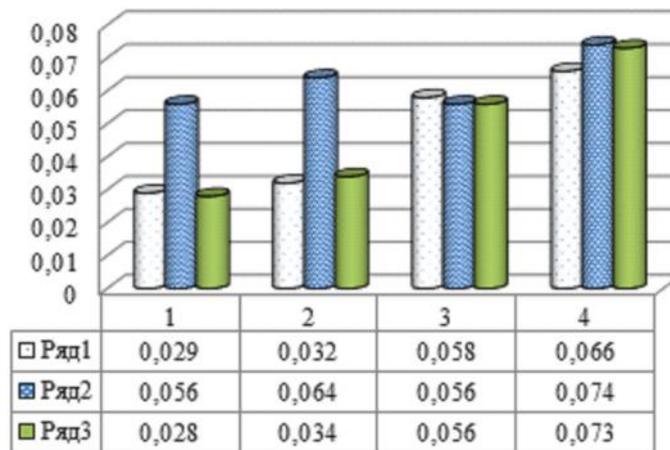
[11, 12].

2,0-2,31 ( .4).



.4. , %.

4-  
0,066 0,074 ( .5).



.5. m , .

(*Petroselinum crispum*),

1. ... // ... , 2009.
2. 8. . 42-43. ... // ... , 2010.
3. 12. . 150-155. ... : 03.01.05 : ... , 2010. 318 .
4. ... (Fagopyrum esculentum M.) : 03.01.05: ... , 2013. 24 .
5. ... // « ... » (15-17 2017 .) / ... , ... « ... » , 2017. . 633-637.
6. ... (Crataegus rhipidofilla) // ... , 2017. 28-29 . 41-43.
7. ... / ... , ... , ... [ ... ] // 2024. . 67, 5-6. . 308-313.
8. ... Populus pyramidalis // : III , 10–13 2012 . – I , 2012. . 156-158.
9. ... // : / ... . . . . : , 2009. . 11. . 56-61.
10. ... // , 2023. . 6. 3.
11. ... [ ... ]. – . : « ... » , 2006. . 556. ... , 2025, 27.

12.

/ . . , . .

...  
, . . .  
, 2019. . 62.

[ .] //  
3-4. . 242-246.

13. . . , . .

// , 2017. 12. . 34-43.

14. . . , . . , . .

// , 2010. 3. . 25-27.

15. . . , . .

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// . 2021. 5(122). . 61-64.



CBL10

...  
CBL3, CBL7, CBL9,  
*POPULUS NIGRA* L.

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- ,

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E-mail: *slavaosin@yandex.ru*

(CBL)

“Primer-Blast”. CBL ,

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CBL

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(*P. trichocarpa* Torr. t Gray),

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(*P. simonii* Carr),

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(*P.*

*maximowiczii* Henry)

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[4].

CBL1, CBL2, CBL5, CBL9

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*in vitro*.

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(*P. alba* L. *P. tremula* L.) [5].

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CBL

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CBL3, CBL7, CBL9

(*P. nigra* L.).

(National Center for

Biotechnology Information — NCBI) [6].

Primer-Blast [7].

“BLAST” NCBI,  
“highly similar

sequences” [8].

CBL  
*P.*

*nigra*, *P. trichocarpa*, *P. alba*, *P. euphratica* Oliv.

(*Salix* sp.).

( . 1).

*Populus*

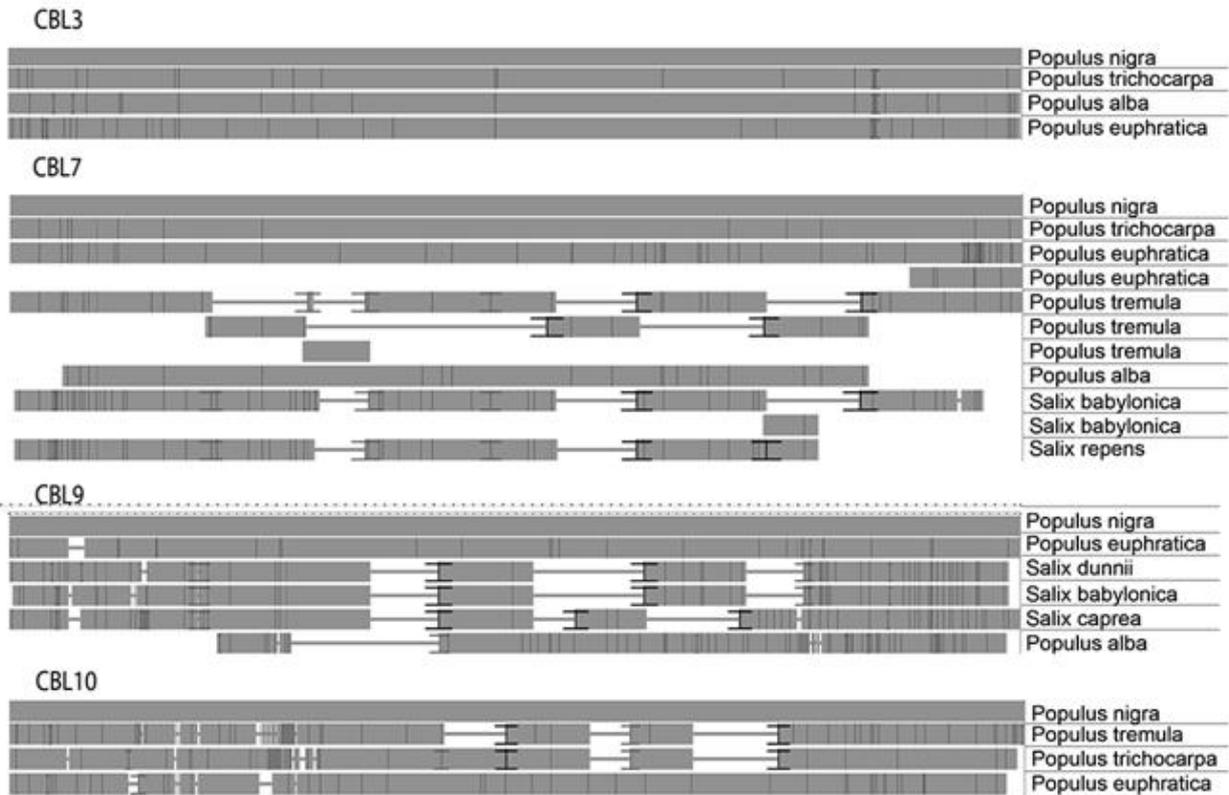
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16–25

AT GC;

[9].



1. CBL (*Populus* *Salix*).

, Tm – (F/R — / , ° ; GC % – 1.

Праймер	Последовательность (5'→3')	Длина продукта	Tm	GC%	Самокомплиментарность	Самокомплиментарность 3'
CBL3						
F	GCGTCGCAATTGTACACAACC	21	60.98	52.38	6.00	2.00
R	GCTTCTGCTAGCCAATCCCA	20	60.11	55.00	6.00	0.00
CBL7						
F	AAGCCTGCTGCTGTATTGGT	20	59.96	50.00	4.00	0.00
R	CGCGTTCGATGAAACCTGTC	20	59.9	55.00	4.00	2.00
CBL9						
F	TAGTAGCCGTCCGATTACGGT	21	60.48	52.38	4.00	3.00
R	GTTTGTGAGGCAAGGGCAAC	20	60.25	55.00	3.00	0.00
CBL10						
F	AGAATTTGCCCGAGCACTCA	20	59.96	50.00	6.00	2.00
R	ACATCTGCCGGACTTCTTCC	20	59.75	55.00	4.00	1.00

## CBL3, CBL7, CBL9, CBL10

## CBL,

1. ... //
2. Tang R.J., Yang Y., Yang L., Liu H., Wang C.T., Yu M.M., Gao X.S., Zhang H.X. Poplar calcineurin B like proteins PtCBL10A and PtCBL10B regulate shoot salt tolerance through interaction with PtSOS2 in the vacuolar membrane // *Plant, cell & environment*. – 2014. – Vol. 37. – No. 3. – P. 573–588.
3. Batisti O., Kudla J. Plant calcineurin B-like proteins and their interacting protein kinases // *Biochimica et Biophysica Acta (BBA) – Molecular Cell Research*. – 2009. – Vol. 1793. – No. 6. – P. 985–992.
4. CBL- //
2020. – 4. – 4–13.
5. //
- 3. – 15–23.
6. “The National Center for Biotechnology Information”; <https://www.ncbi.nlm.nih.gov/> (available: 03/25/2025).
7. “Primer-Blast”; URL: <https://www.ncbi.nlm.nih.gov/tools/primer-blast/index.cgi> (available: 01/25/2025).
8. “BLAST”; URL: <https://blast.ncbi.nlm.nih.gov/Blast.cgi> (available: 01/25/2025).
9. Primer-BLAST //
- 2021. – 8 (3). – 37–52.



577.15

**(RATTUS NORVEGICUS L.)**

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-mail: *bc366@bio.vsu.ru*

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*Pc,*

1q43 (Gene ID: 25104) (<https://www.ncbi.nlm.nih.gov/gene/25104>).

*Rattus norvegicus* L.,

*(Rattus norvegicus* L.) *Pc*

GenBank

(<https://www.ncbi.nlm.nih.gov/genbank/>).

BLAST (<https://blast.ncbi.nlm.nih.gov/>).

Primer-

BLAST.

BLAST-online [1,2],

FastPCR,

TermoFisher.

LiCl [4].

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( / ).

0,1%

M-MuLV

(dT) (“ ”, )

AmpliSence (“ ”, )

LightCycler 96 (“Roche”, )

*Pc*

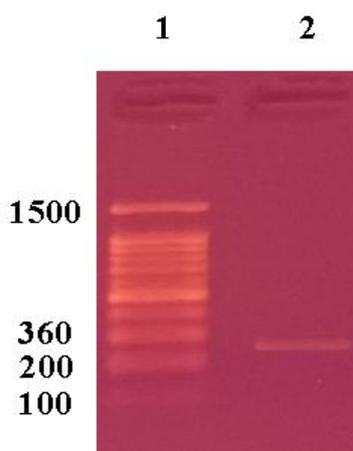
( . 1).

1.

*Pc*

Ген	Праймер	Нуклеотидная последовательность	Температура плавления, °C	Размер продукта, п.н.
<i>Pc</i>	Прямой	5'-ttcctgtcagtgaggc-3'	57.6	264
	Обратный	5'-ggatggcaatctcacctctg-3'	59.6	

59° ( . 1).



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59° .

(1)  
*Pc*

1. Alvarez-Fernandez R. Explanatory chapter: PCR primer design // *Methods Enzymol.* - 2013. - V.529. - P. 1-21.
2. Noguera D.R. Mathematical tools to optimize the design of oligonucleotide probes and primers / Noguera D.R., [et al.] // *Appl. Microbiol. Biotechnol.* - 2014. - V. 98. - 23. - P.9595- 9608.
3. Wallace J. Distribution and biological functions of pyruvate carboxylase in nature. In: Keech D, Wallace J, editors. *Pyruvate Carboxylase.* // Boca Raton: CRC Press. - 1985. - P. 5–64.
4. ... - 2008. - 62 .



(ZEA MAYS L.)

• ” • ” • ” • •

E-mal: bc366@bio.vsu.ru

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( - ; 2.6.1.19) — [1]. -

( ) [2]. -

[3].

(Zea mays L.) [4].

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*A. thaliana*.

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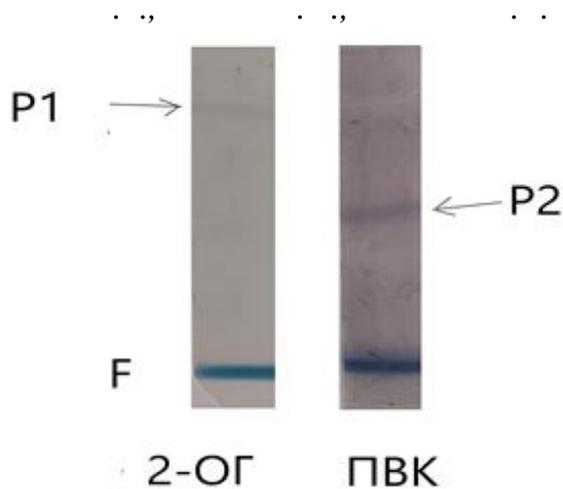
*Arabidopsis (AtGABA-T)*. *E.*  
*coli* , - - ,  
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*AtGABA-T*, , N- ,  
 [6].

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12-  
 (*Zea mays* L.) « 76»,  
 25 / <sup>2</sup>.

20 10- .  
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 2-4 [5]..





1. Khan, Muhammad Sarwar et al. Enzymes: Plant-based Production and their Applications.// Protein and peptide letters. - 2018.- V. 25, - 2. – p.136-147.  
 2. Nonaka, Satoko et al. An Agrobacterium tumefaciens Strain with Gamma-Aminobutyric Acid Transaminase Activity Shows an Enhanced Genetic Transformation Ability in Plants.// Scientific reports. - 2017. - V. 7. – . 42649  
 3. Shimajiri Y. et al. Differential subcellular localization, enzymatic properties and expression patterns of  $\gamma$ -aminobutyric acid transaminases (GABA-Ts) in rice (*Oryza sativa*) //Journal of Plant Physiology. – 2013. – V. 170. – . 2. – . 196-201.  
 4. Guo X. et al. An aminobutyric acid transaminase in *Zea mays* interacts with *Rhizoctonia solani* cellulase to participate in disease resistance //Frontiers in Plant Science. – 2022. – V. 13. – P. 860170.  
 5. ... : ... , 1971.  
 6. Clark SM, Di Leo R, Dhanoa PK, Van Cauwenberghe OR, Mullen RT, Shelp BJ. Biochemical characterization, mitochondrial localization, expression, and potential functions for an *Arabidopsis* gamma-aminobutyrate transaminase that utilizes both pyruvate and glyoxylate// J Exp Bot. – 2009. – V.60. - 6. – .1743-1757.  
 ..., 2025, 27.

1. Khan, Muhammad Sarwar et al. Enzymes: Plant-based Production and their Applications.// Protein and peptide letters. - 2018.- V. 25, - 2. – p.136-147.
2. Nonaka, Satoko et al. An Agrobacterium tumefaciens Strain with Gamma-Aminobutyric Acid Transaminase Activity Shows an Enhanced Genetic Transformation Ability in Plants.// Scientific reports. - 2017. - V. 7. – . 42649
3. Shimajiri Y. et al. Differential subcellular localization, enzymatic properties and expression patterns of  $\gamma$ -aminobutyric acid transaminases (GABA-Ts) in rice (*Oryza sativa*) //Journal of Plant Physiology. – 2013. – V. 170. – . 2. – . 196-201.
4. Guo X. et al. An aminobutyric acid transaminase in *Zea mays* interacts with *Rhizoctonia solani* cellulase to participate in disease resistance //Frontiers in Plant Science. – 2022. – V. 13. – P. 860170.
5. ... : ... , 1971.
6. Clark SM, Di Leo R, Dhanoa PK, Van Cauwenberghe OR, Mullen RT, Shelp BJ. Biochemical characterization, mitochondrial localization, expression, and potential functions for an *Arabidopsis* gamma-aminobutyrate transaminase that utilizes both pyruvate and glyoxylate// J Exp Bot. – 2009. – V.60. - 6. – .1743-1757.

..., 2025, 27.



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E-mail: *rybolov@mail.ru*

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-6- ( 6 , 1.1.1.49) -

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(GSH) ( ) .

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6 [3]. 6

[2].

*Rattus norvegicus* L. 180-200 .

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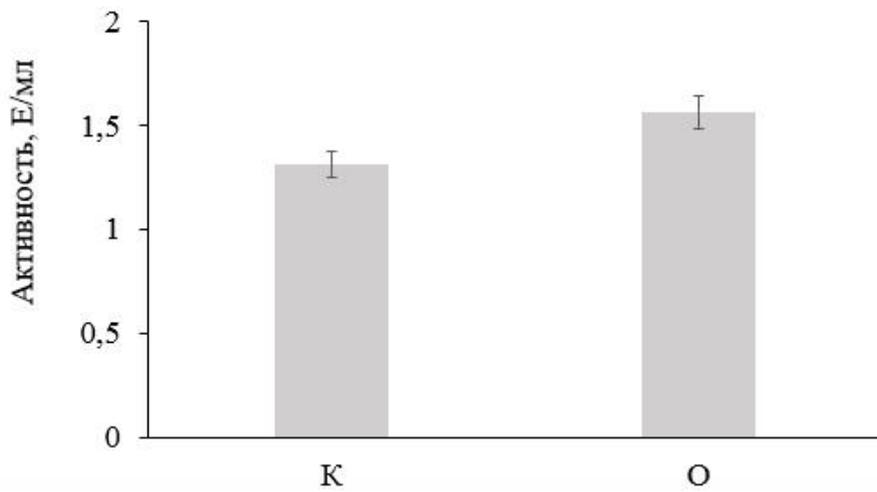
STATISTICA.

< 0.05.

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1,2 (1,56 ± 0,08 / ),

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2.

184

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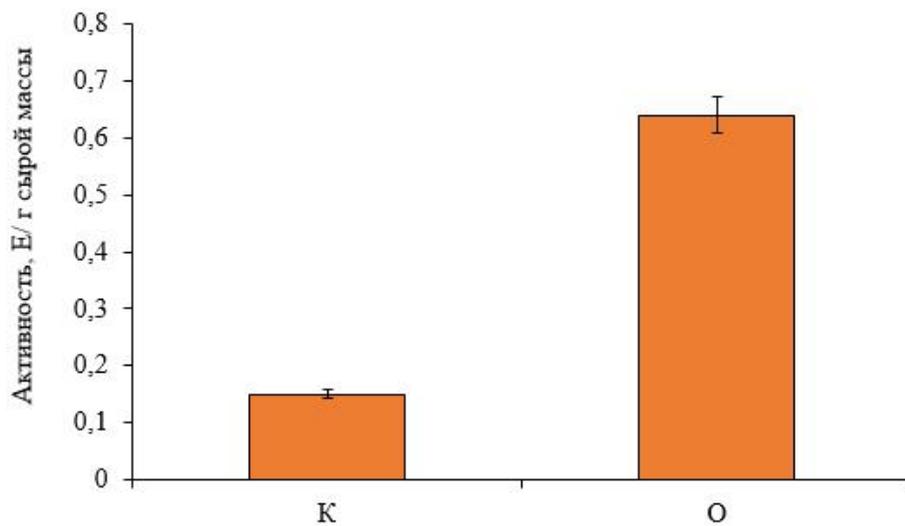
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0,64 /

4,64

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1. Luzzatto L. Glucose-6-Phosphate Dehydrogenase Deficiency. Hematology / L. Luzzatto, C. Nannelli, R. Notaro // Oncology Clinics of North America. – 2016. – V. 30. – P. 373–393.
2. Kletzien R.F. Glucose-6-phosphate dehydrogenase: a “housekeeping” enzyme subject to tissue-specific regulation by hormones, nutrients, and oxidant stress / R.F. Kletzien, P.K. Harris, L.A. Foellmi // FASEB J. – 1994. – V. 8. – P. 174–181.
3. Glucose-6-phosphate dehydrogenase expression associated with NADPH-dependent reactions in cerebellar neurons / E. Biagiotti [et al.] // Cerebellum. – 2003. – V. 2. – P. 178–183.
4. / . . . . – . : , 1987. – 368 .



577.12

**FOXA**

**-6-**

**G6PD**

E-mail: *rybolov@mail.ru*

*G6pd*

-6-

Foxa

- Foxa,

(Pepck),

-6-

(G6pc)

[1]. Foxa

Glut2,

Foxa

[2, 3].

[AC]A[AT]T[AG]TT[GT][AG][CT]T[CT]-3' [4].

24377), *norvegicus*, -6- *G6pd* (Gene ID: *Rattus*: ~900

NCBI (<https://www.ncbi.nlm.nih.gov>).

GeneCards (<https://www.genecards.org/>).

Foxa

Primer-Blast.

*G6pd* -6-

CREB.

Foxa ( . 1).

*G6pd*

Foxa

(TSS).

TCAGTCAAAGCACACGCCCTCTTGCGTTAAATGGGCCAACGAAGCTTAGCCCCGAAACGGGCTGTGCA  
CTCCAGATCTGTGAACGTGTTTGGCAGCGGCAACTAAATTCAGGTAAAGGGGTGGTCTGATGTCACATCG  
GTGGAGGGACATGCTGCCCGCTTGTATGCCGCGTGTTCCTTTGATGCGCGCTAATCTACAAGGTCTTAAC  
CCATCTCTTTGTATCCCGCACTTTCTCTGCACTGGGCCCAAGAGAAGTAGACCTTTACAAGAGGAGGTAC  
GTTCCGGAATCTCCTAACTGGGCCAGAAGAGAGCCGGAGCTGAACTCCATCATCTACACAATCAAACCTG  
CTCAGTTCATTGTGGAGTTTCTGGCAGACACCAGAGTATGATGGTCCCATATAGAGTGTGATGCCCTGAC  
TGTATTATATCTAGCTGCCCAGAGTGGATGCCTTAGGTGAATTCTATTAATGCTTTAGTACTTTCGTAAT  
GTACAGCCCCTAGTGAAGGCAATATAGGAGTTATGATTGGATCCCAGAGGCAGCAGTTTCTGAGTCTCC  
GAATGCACAGGCGCGCGCGCGTGTACACACACACACAGCATTCCCAGCCAGTGGTTTTAGTAATCGTC  
ACCGGTCTGGTGAGAAGCTTAAAGGACCCTTGAACGAGGCTAAGTTGTTAATGCTCTTCCATTCTCCTGCC  
TATCAATGAGCCTTTCTTCCACCAGAAAACATCATG

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*G6pd*

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Foxa,

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 TSS -60  
 +40 . . 200 300 . . TSS

[5].

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 [ (INR)  
 (DPE)] ( CAAT-  
 Y- ).  
 (TFBS),

[6, 7]. TFBS

[8].

Foxa

-6-

Primer-BLAST

Foxa .

Foxa *Rattus*

*norvegicus* L.

1.

**Foxa (NM\_012742)**

мРНК	Прямой праймер	Обратный праймер
Foxa	GCACAATTTTCCCCGGTTCA	CCTGCCACGGGACTAGAATG

-6-

Foxa.

(TSS),

*G6pd*

1. Friedman JR, Kaestner KH. The Foxa family of transcription factors in development and metabolism. *Cell Mol Life Sci.* 2006 Oct;63(19-20):2317-28.
2. Shen, W., Searce, L. M., Brestelli, J. E., Sund, N. J. and Kaestner, K. H. (2001) Foxa3 (hepatocyte nuclear factor 3gamma) is required for the regulation of hepatic GLUT2 expression and the maintenance of glucose homeostasis during a prolonged fast. *J. Biol. Chem.* 276, 42812–42817.
3. Kaestner, K. H., Hiemisch, H. and Schutz, G. (1998) Targeted disruption of the gene encoding hepatocyte nuclear factor 3gamma results in reduced transcription of hepatocyte-specific genes. *Mol. Cell Biol.* 18, 4245–4251
4. Lin L, Miller CT, Contreras JI, Prescott MS, Dagenais SL, Wu R, Yee J, Orringer MB, Misek DE, Hanash SM, Glover TW, Beer DG. The hepatocyte nuclear factor 3 alpha gene, HNF3alpha (FOXA1), on chromosome band 14q13 is amplified and overexpressed in esophageal and lung adenocarcinomas. *Cancer Res.* 2002 Sep 15;62(18):5273-9.
5. Roy A.L., Singer D.S. Core promoters in transcription: old problem, new insights // *Trends Biochem. Sci.* – 2015. – V. 40. – P. 165-171.
6. Yanfang Y., Kaikai Z., Liying Y., Xing L., Ying W., Hongwei L., Qiang L., Duanfen C., Deyou Q. Identification and characterization of MYC transcription factors in *Taxus* sp. // *Gene.* – 2018. – V. 675. – P. 1-8.
7. Zhou et al., Zhou T., Luo X., Yu C., Zhang C., Zhang L., Song Y., Dong M., Shen C. Transcriptome analyses provide insights into the expression pattern and sequence similarity of several taxol biosynthesis-related genes in three *Taxus* species // *BMC Plant Biol.* – 2019. – V. 19: 33.
8. Law J.A., Jacobsen S.E. Establishing, maintaining and modifying DNA methylation patterns in plants and animals // *Nat. Rev. Genet.* – 2010. – V. 11. – P. 204-220.



577.11

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E-mail: *aleck.selivanoff2013@yandex.ru*

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TBE- pH 8.3.

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(1B3A) 5 (1B3A) 5 (4A) : 1 (4B), 2 (3B1A), 3 (2B2A), 4

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pH 8.6 [1] 8,3 [2].

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GLP.

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2. 0,5% 1× , 8.3

3. 0,5% 1× TBE , 8.3

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TBE

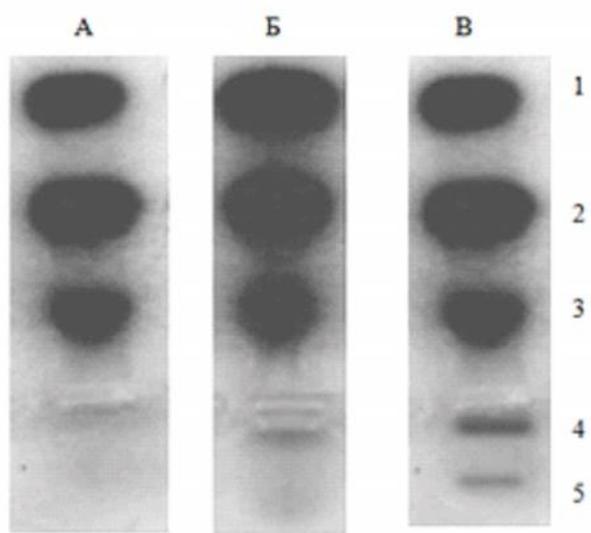
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 M . 2 37 .

1-3



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 . 1-5 - , - TBE-

, 5, , TAE-

4

0,5%

100

10

1. Barnett H. The staining of lactic dehydrogenase isoenzymes after electrophoretic separation on cellulose acetate. J Clin Pathol. 1964 Sep;17(5):567-70

2. Li P.J., Qiu N.H., Zhang L., Zhong G.S., Zeng F.C. A more universal and stable method for lactate dehydrogenase isoenzyme test. Anal. Methods. 2019;11:4173–4183.

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-mail: *polina\_02\_11@bk.ru*

4.2.1.3)

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- ( , , 4.2.1.3).

[3],

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(*Rattus norvegicus*)

150-200 ..

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 2- 1,5  
 VALUE «1 Stage Vacuum Pump»,

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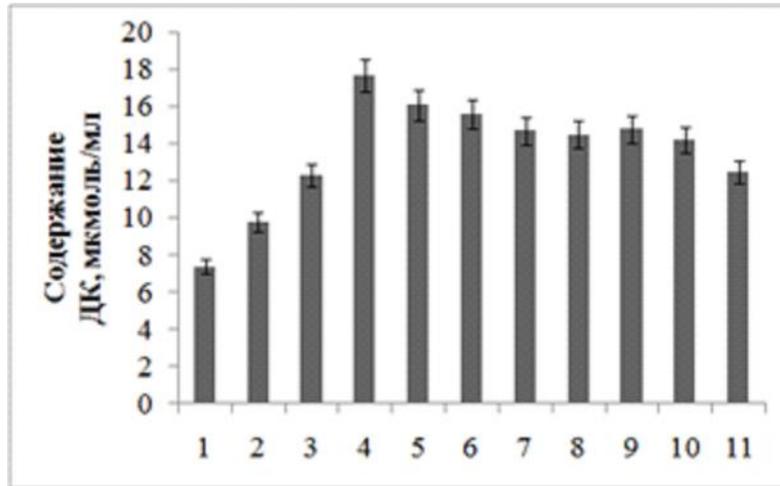
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240 [7].

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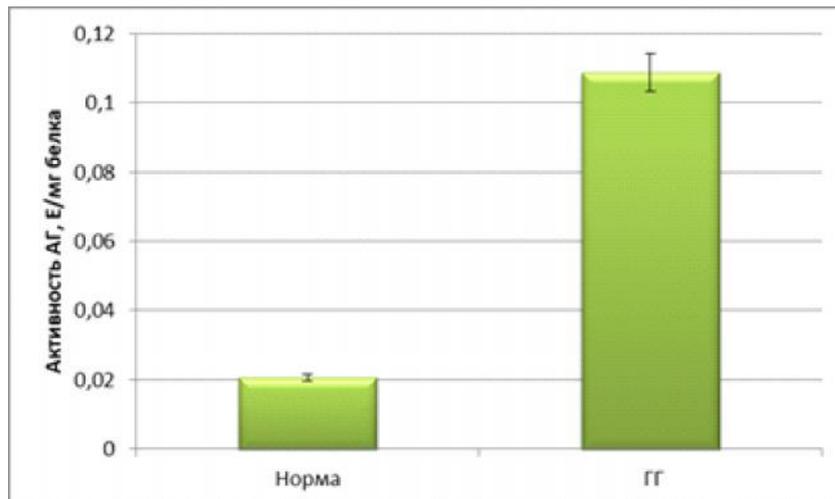
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(1), 35 (8), 40 (9), 50 (10) 60 (11)

5 (2), 10 (3), 15 (4), 20 (5), 25 (6), 30 (7),

2,

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1. ...

2. ... 2015;77(6):39-44. <https://doi.org/10.17116/patol201577639-44>

3. Chen X.J., Wang X., Butow R.A. (2007) Yeast aconitase binds and provides metabolically coupled protection to mitochondrial DNA. PNAS USA., 104, 13738-13743.

4. Bulteau A.L., O'Neil H.A., Kennedy M.C., Ikeda-Saito M., Isaya G., Szveda L.I. (2004) Frataxin acts as an iron chaperone protein to modulate mitochondrial aconitase activity. Science., 305, 242-245.

5. ... 2011;3:39-41.

6. ... 2014;1:25-31

7. Cherkasov A.A., Overton R.A.Jr., Sokolov E.P., Sokolova I.M. (2010) Temperature-dependent effects of cadmium and purine nucleotides on mitochondrial aconitase from a marine ectotherm, *Crassostrea virginica*: a role of temperature in stress and allosteric enzyme regulation. The Journal of Experimental Biology., 210, 46-55.



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-mail: *polina\_02\_11@bk.ru*

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1,7

[1].

[6].

[4].

[5].

*(Rattus norvegicus)*

150-200 .,

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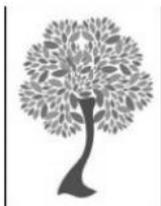
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630\* 161. 443. 66

**EX VITRO**

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Email: [tatyana.tabacky@gmail.com](mailto:tatyana.tabacky@gmail.com)

(*Populus alba L.*), . (*P. canescens Sm.*),

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(NaCl) *in vitro*,

*ex vitro.*

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*ex*

*vitro.*

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[1].

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[2-4].

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*in vitro*,

*ex vitro* [2].

*in vitro*,

*purpurea* (L.) Moench.), (*Echinacea*

[5, 9].

*in vitro*

*ex vitro*

(*Populus alba* L.), (*P. canescens* Sm.),

(NaCl)

[6, 7].

5-7 [8].

100 . 1<sup>2</sup>

(0,1 / ; 0,2 / ; 0,3 / )  
0,2 /

*ex vitro.*

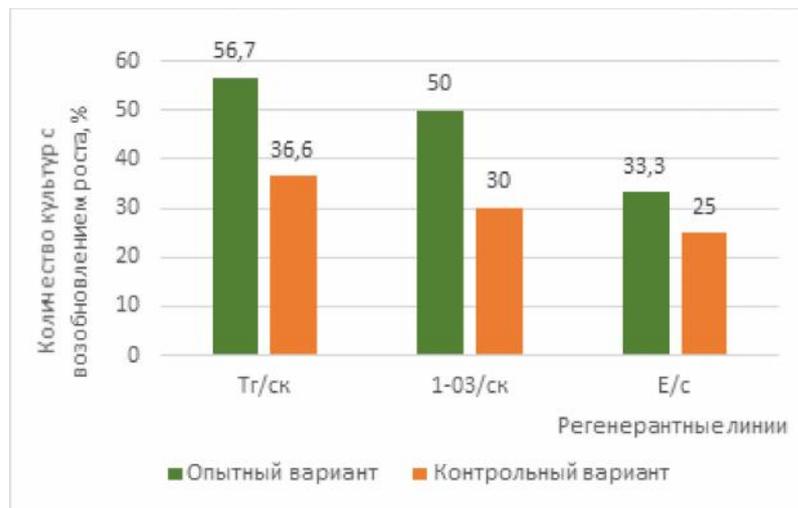
( 200

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20

15

1,6 (46,6% 29,5% ) ( 1).



.1.

*ex vitro.*

( 1).

1.

Регенерантные линии	Опыт		Контроль	
	Высота побега, см	Приживаемость, %	Высота побега, см	Приживаемость, %
Тг/ск	14,5±1,40	96,7±1,30	10,6±2,10	90,0±1,02
1-03/ск	14,8±1,20	98,3±1,40	12,3±1,40	88,3±1,20
Е/с	12,9±1,20	91,6±1,04	9,0±2,20	81,7±1,60
Среднее	14,1±0,3	95,5±1,0	10,6±0,5	86,7±1,3

[5,9].

( 91,1%)

(95,5% 86,7%, ).

( 2).

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(14,1 ) 1,3

(10,6 ).



a

b

c

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12,9-14,8

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9,0-12,3

( 2).

*ex vitro.*





574.24

**AZOSPIRILLUM**

• •

E-mail: *maryorl@mail.ru*

( ) (GSH)  
*Azospirillum brasilense* DSM 1690<sup>T</sup>, *A. picis* DSM 19922<sup>T</sup>

GSH

—

, . .)

[8, 12].

( ) [7].

[6].

*Azospirillum* spp.,

, *A. brasilense* EMCC1454

Cr 260 , *A. lipoferum* 137

..., 2025, 27.

Pb Cd, *A. brasilense* Cd

5 As(V) 250 As, As(III) [3, 5, 11].

*A. brasilense*

DSM 1690<sup>T</sup> *A. picis* DSM 19922<sup>T</sup>

Pb.

( )

(GSH).

GSH

*Azospirillum brasilense* DSM

1690<sup>T</sup>, *A. picis* DSM 19922<sup>T</sup>.

: (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> – 1,0 / , MgSO<sub>4</sub> \* 7H<sub>2</sub>O –

1,0 / , CaCl<sub>2</sub> \* 2H<sub>2</sub>O – 0,03 / ,

– 1,0 / , – 2,0 / ,

– 1,0 ,

[4, 10], pH

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3 29°C

Pb(NO<sub>3</sub>)<sub>2</sub>

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g 10

0,1M Tris-HCl

pH 7,5

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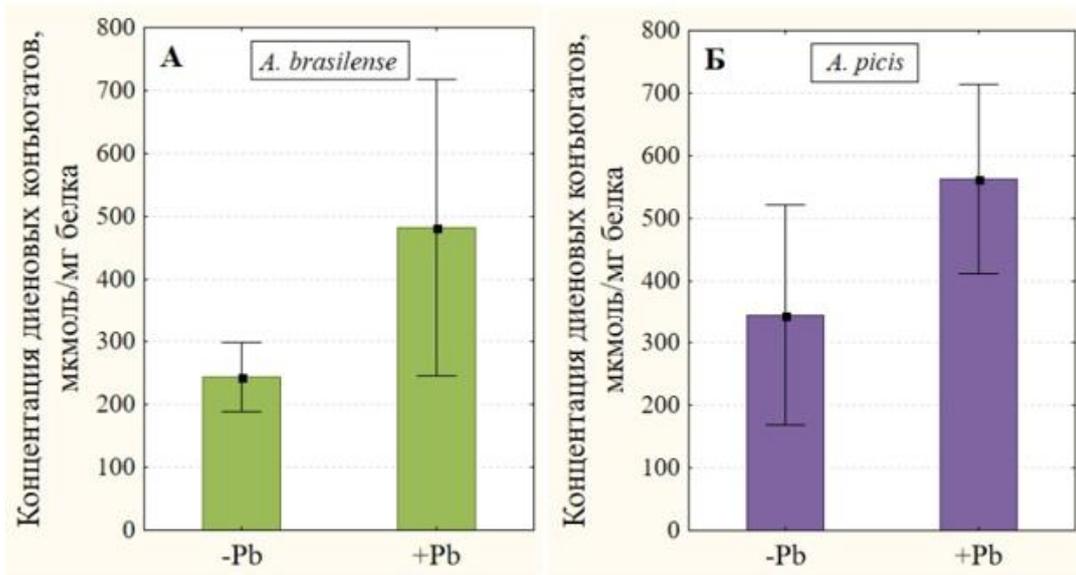
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233 [2].

GSH

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, *A. brasilense* DSM 1690<sup>T</sup> *A. picis* DSM 19922<sup>T</sup>,  
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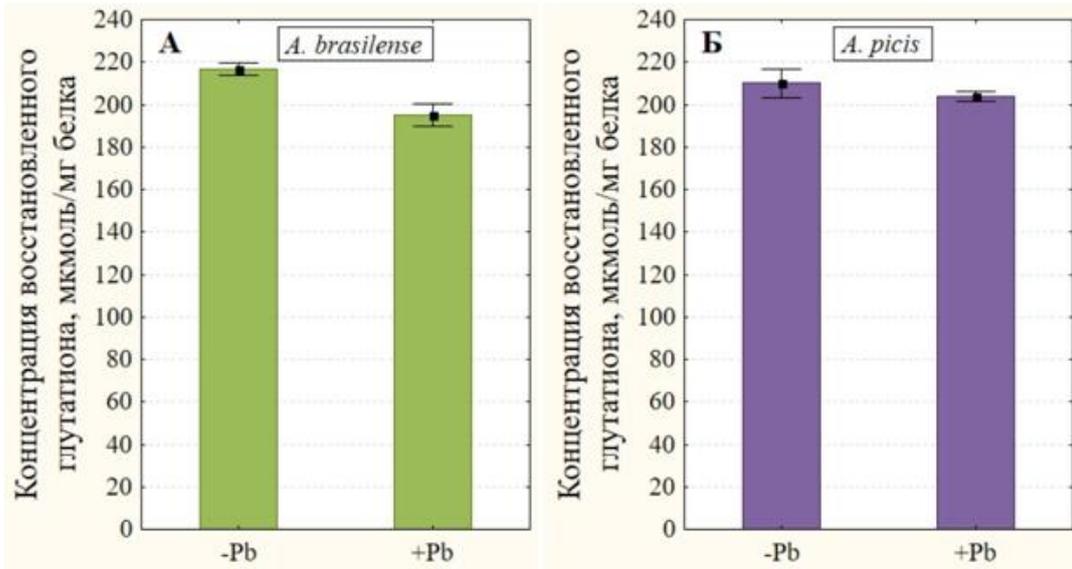


. 1. *A. brasilense* ( ) *A. picis*  
( ) / .

GSH, , (GSH)  
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GSH - , GSH

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*A. picis* ( ) .2.

*A. brasilense* ( )

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1. ... // ... - 2001. - . 381, 2. - . 262–264.
2. ... // ... / ... , 1972. - . 63–64.
3. Belimov A. A. Employment of rhizobacteria for the inoculation of barley plants cultivated in soil contaminated with lead and cadmium / A. A. Belimov [et al.] // Microbiology. - 2004. - Vol. 73. - P. 99–106.
4. Caraway B. H. Aerotaxis in *Spirillum volutans* / B. H. Caraway, N. R. Krieg // Canadian Journal of Microbiology. – 1974. – Vol. 20. – P. 1367–1377.
5. El-Ballat E. M. Metal-Resistant PGPR Strain *Azospirillum brasilense* EMCC1454 Enhances Growth and Chromium Stress Tolerance of Chickpea (*Cicer arietinum* L.) by Modulating Redox Potential, Osmolytes, Antioxidants, and Stress-Related Gene Expression / E. M. El-Ballat [et al.] // Plants. - 2023. - Vol. 12. - P. 2110.

6. Huertas M. J. Metals in Cyanobacteria: Analysis of the Copper, Nickel, Cobalt and Arsenic Homeostasis Mechanisms / M. J. Huertas [et al.] // *Life*. - 2014. - Vol. 4. - P. 865–886.
7. Hultberg B. Interaction of metals and thiols in cell damage and glutathione distribution: potentiation of mercury toxicity by dithiothreitol / B. Hultberg [et al.] // *Tox*. - 2001. - Vol. 156. - P. 93–100.
8. Li Y. Enrofloxacin at environmentally relevant concentrations enhances uptake and toxicity of cadmium in the earthworm *Eisenia fetida* in farm soils / Y. Li [et al.] // *J. Hazard. Mater.* - 2016. - Vol. 308. - P. 312–320.
9. Lowry O.H., Rosenbrough N., Farr A., Randall R.J. Protein measurement with the folin phenol reagent // *The Journal of Biological Chemistry*. – 1951. – V. 193. – P. 265–75.
10. Pfennig N. über das Vitamin B12-Bedürfnis phototropher Schwefelbakterien / N. Pfennig, K. D. Lippert // *Archiv. Mikrobiol.* – 1966. – Vol. 55. – P. 245–256.
11. Vezza M. E. Biochemical and molecular characterization of arsenic response from *Azospirillum brasilense* Cd, a bacterial strain used as plant inoculant / M. E. Vezza [et al.] // *Environ Sci Pollut Res.* - 2020. - Vol. 27. - P. 2287–2300.
12. Wahid A. Cadmium phytotoxicity: responses, mechanisms and mitigation strategies: a review / A. Wahid [et al.] // *Organic Farming, Pest Control and Remediation of Soil Pollutants*. - 2009. - P. 371–403.



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(*Gallus gallus domesticus*)

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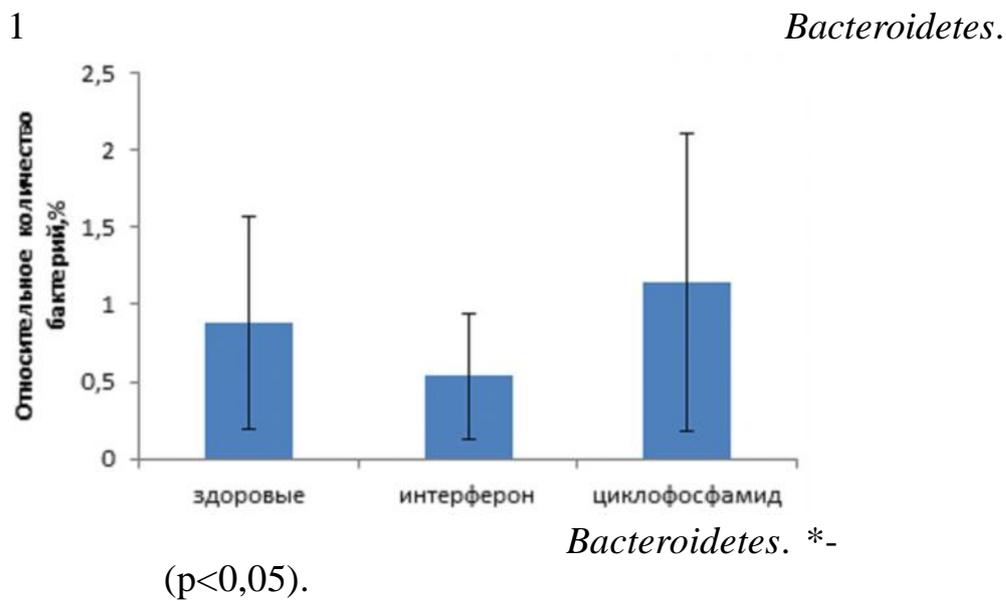
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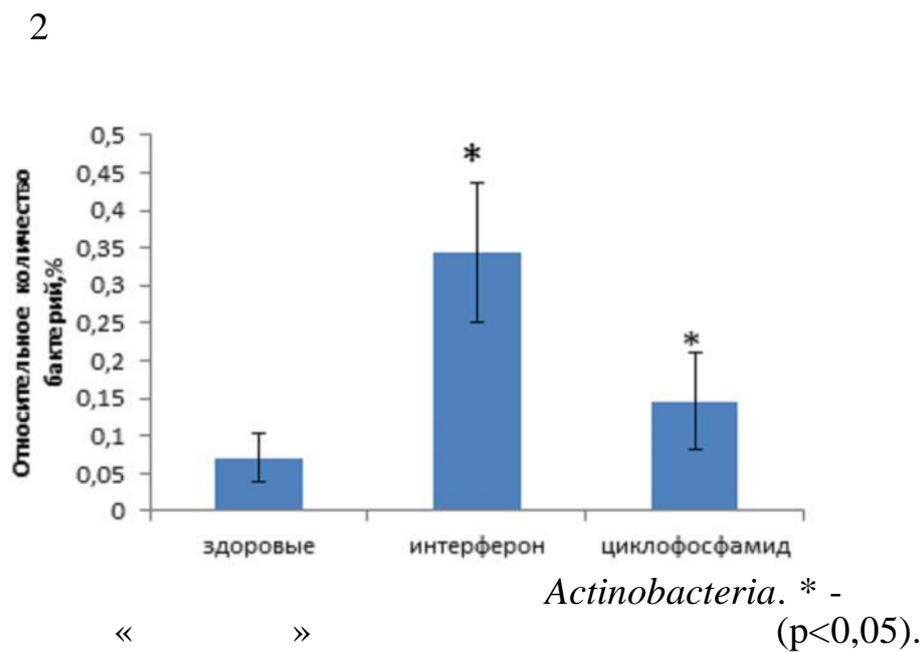
30 ; 72 ° 45 ;

72 ° 5 .

Yun-Wen Yang, 2015.



*Bacteroidetes* 40% 30%



*Actinobacteria* 5, 2, 3

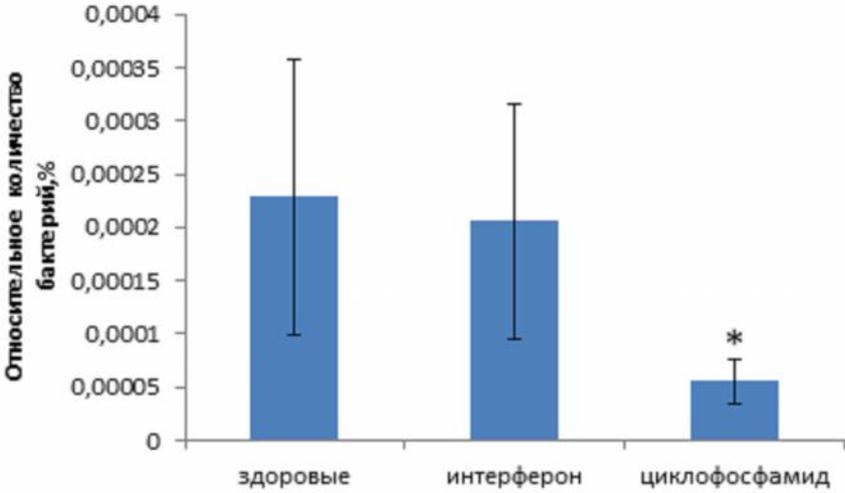
*Deferribacteres.*

80%

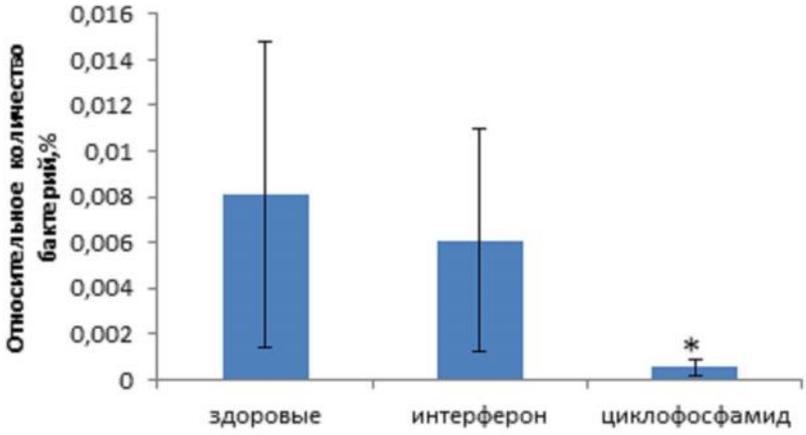
*Deferribacteres*

4

*Saccharibacteria.*



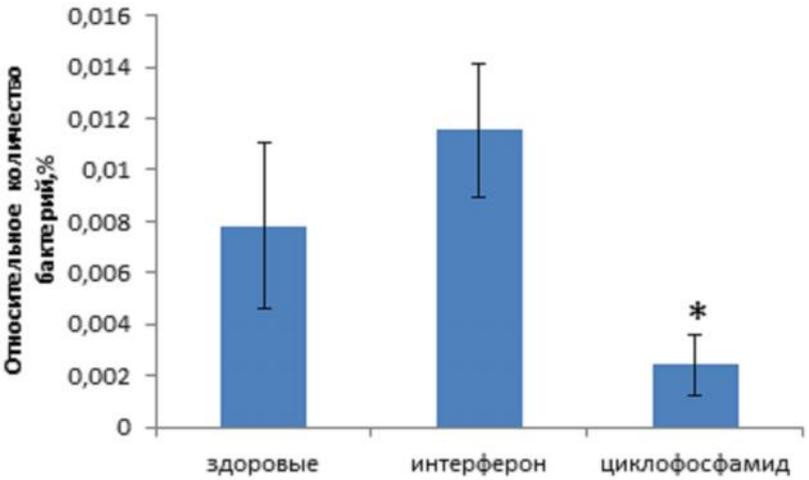
. 3. « » *Deferribacteres.* \*- (p<0,05).



. 4. « » *Saccharibacteria.* \*- (p<0,05).

93 %

*Saccharibacteria.*



. 5. « » *Verrucomicrobia.* \*- (p<0,05).

5

Verrucomicrobia.

70% *Verrucomicrobia*

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*Actinobacteria*,

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*Deferribacteres*

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1. Lebedev S. Cytokine and immunoglobulin profiles of Arbor Acres broiler chickens at different stages of physiological development / S.Lebedev, T. Kazakova, O.Marshinskaia // *Vet World*.- 2024. - 17(5) – P.988-993.
2. Li C. Editorial: Immunosuppressive disease in poultry/ C.Li, L. Wang, S.Zheng. / *Front Immunol*. – 2023. – 14 – P.1215513.
3. Nazar F.N. The immune-neuroendocrine system, a key aspect of poultry welfare and resilience / F.N. Nazar, I. // *Estevez Poult Sci*. – 2022. – 101(8) – P. 101919.
4. . . . / . . . // . – 2018. – 34(2)– . 45-52.
5. . . . // : , , . – 2017. – 1(18). – . 23-31



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**(FRAXINUS EXCELSIOR)**

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-mail: *Rima.usa@yandex.ru*

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*(Agrilus planipennis).*

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*(Fraxinus excelsior)* -

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[8,9,10].

(*Agrilus planipennis*),

[11,12].

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[13,14].

*Fraxinus*.

[15, 16, 17].

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[17, 18, 19].

*Hymenoscyphus fraxineus*.

*Hymenoscyphus*,

2006

*Chalara fraxinea.*

2009

*Chalara fraxinea*

*Hymenoscyphus pseudoalbidus,*

*Hymenoscyphus fraxineus. H. fraxineus*

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*H. fraxineus,*

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85%

[21].

(*Fraxinus excelsior*).

(*Fraxinus excelsior*)

(*F. angustifolia*) [22].

(*F. excelsior*)

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*Fraxinus.*

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*Fraxinus* ( )

*Hymenoscyphus fraxineus*,

(ADB),

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*F. Pennsylvanica* \* *F. excelsior*

*F. Ornus* \* *F. Excelsior* *F. latifolia*. \* *F. Excelsior*.

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1. (Fraxinus excelsior) / . . . // . – 2021. – . 7 (52). – . 10-12.
2. (Agrilus planipennis) (Coleoptera: Buprestidae) / . . . // . – . 33.
3. (Agrilus planipennis) (Coleoptera: Buprestidae) / . . . // . – 2022. – . 32. – . 33-41.
4. (Agrilus planipennis) / . . . 2022 / . . . // . – 2022. – . 317-319.
5. / . . . // . – . 25.
6. Valenta V. A new forest pest in Europe: a review of Emerald ash borer (*Agrilus planipennis*) invasion / V. Valenta et al. // Journal of Applied Entomology. — 2016. — Vol. 141. — P. 507—526.
7. Wang Xiao-Yi. The biology and ecology of the emerald ash borer, *Agrilus planipennis*, in China/ Xiao-Yi Wang // Journal of Insect Science. — 2010. — Vol. 10. — P. 97—116.
8. (Agrilus planipennis) / . . . // . – 2023. – . 18-21.
9. // . – Litres. - 2022.
10. Baranchikov Y. N. Changes in Climatic Range of the Emerald Ash Borer *Agrilus planipennis* Fairmaire (Coleoptera: Buprestidae) in the Northern Hemisphere/ Y. N. Baranchikov, N. Y. Dobrolyubov, S. M. Semenov // Russian Journal of Biological Invasions. – 2024. – . 15. – . 4. – . 480-490.
11. Poland T. M. Review of the emerald ash borer (Coleoptera: Buprestidae), life history, mating behaviours, host plant selection, and host resistance/ . . Poland et al. // The Canadian Entomologist. – 2015. – . 147. – . 3. – . 252-262.
12. Showalter D. N. Resistance of European ash (*Fraxinus excelsior*) saplings to larval feeding by the emerald ash borer (*Agrilus planipennis*) / D. N. Showalter, R. J. Saville et al. // Plants, People, Planet. — 2020. — Vol. 2. — P. 41—46.
13. Plumb W. J. Genetic barcodes for ash *Fraxinus* species and generation of new wide hybrids W. J. Plumb et al. // bioRxiv. – 2024. – . 958 – 1010.
14. Sollars E. S. A. Genome sequence and genetic diversity of European ash trees/ E. S. A. Sollars et al. // Nature. – 2017. – . 541. – . 7636. – . 212-216.
15. Duan J. J. Progress and challenges of protecting North American ash trees from the emerald ash borer using biological control/ J. J. Duan et al. // Forests. – 2018. – . 9. – . 3. – . 142.
16. Huff M. A high quality reference genome for *Fraxinus pennsylvanica* for ash species restoration and research / . Huff et al. // Molecular ecology resources. – 2022. – . 22. – . 4. – . 1284-1302.
17. Kelly L. J. Convergent molecular evolution among ash species resistant to the emerald ash borer/ L. J. Kelly et al. // Nature ecology & evolution. – 2020. – . 4. – . 8. – . 1116-1128

18. Kelly L. J. Genes for ash tree resistance to an insect pest identified via comparative genomics/ L. J. Kelly et al. //bioRxiv. – 2019. – . 772 – 913.
19. Kelly L. J. Analysis of the giant genomes of *Fraxinus* indicates that a lack of DNA removal characterizes extreme expansions in genome size/ L. J. Kelly et al. //New Phytologist. – 2015. – . 208. – . 2. – . 596-607.
20. Bakys R. Investigations concerning the role of *Chalara fraxinea* in declining *Fraxinus excelsior*/ R. Bakys et al. //Plant pathology. – 2019. – . 58. – . 2. – . 284-292.
21. Nielsen L. R. The susceptibility of Asian, European and North American *Fraxinus* species to the ash dieback pathogen *Hymenoscyphus fraxineus* reflects their phylogenetic history/ L. R. Nielsen et al. //European Journal of Forest Research. – 2017. – . 136. – . 59-73.
22. Bakys R. Occurrence and pathogenicity of fungi in necrotic and non-symptomatic shoots of declining common ash (*Fraxinus excelsior*) in Sweden/ R. Bakys et al. // European Journal of Forest Research. – 2019. – . 128. – . 51-60.
23. Sollars E. S. A. Genome sequence and genetic diversity of European ash trees/ E. S. A. Sollars et al. //Nature. – 2017. – . 541. – . 7636. – . 212-216.
24. Kowalski T. Virulence of *Hymenoscyphus albidus* and *H. fraxineus* on *Fraxinus excelsior* and *F. pennsylvanica* / . Kowalski, . Bila ski, . Holdenrieder //PloS one. – 2015. – . 10. – . 10. – . 141-192.
25. Nielsen L. R. The susceptibility of Asian, European and North American *Fraxinus* species to the ash dieback pathogen *Hymenoscyphus fraxineus* reflects their phylogenetic history/ L. R. Nielsen et al. //European Journal of Forest Research. – 2017. – . 136. – . 59-73.



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-mail: *f\_dmitri@mail.ru*

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Показатель	Здоровые (20)	Больные (20)	Норма
АЛАТ (Ед/л)	52,5	125,7	10-55
АСАТ (Ед/л)	43,7	95,4	10-40
ЩФ (Ед/л)	55,4	425,6	12-55
ГГТ (Ед/л)	2,8	22,8	1-10
Мочевина (ммоль/л)	4,6	24,3	3-11,3
Креатинин (мкмоль/л)	89,5	248,5	35-165
Холестерол (ммоль/л)	3,7	10,6	1,3-6
Альбумин (г/л)	35,2	22,4	25-35
Общий белок (г/л)	77,4	57,8	43-75

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20-45%,

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Показатель	Инфицированные животные	Неинфицированные животные	Физиологическая норма
Гемоглобин, г/л	71	105	80-150
Гематокрит, %	24	30	29-45
Количество эритроцитов, $\times 10^{12}$ клеток	4,2	5,2	4,8-10
MCV (объем эритроцитов), fl	54,8	48,6	38-53
MCH (среднее содержание гемоглобина в эритроците), pg	14,6	15,3	13-20
MCHC (средняя концентрация гемоглобина в эритроците), г/л	254	347	300-360

1. Jarrett, O.; Neil, J.C. Feline Leukaemia Virus. In eLS; John Wiley & Sons, Ltd.: Hoboken, NJ, USA, 2012.

2. Verbrugghe A., Bakovic M. Peculiarities of One-Carbon Metabolism in the Strict Carnivorous Cat and the Role in Feline Hepatic Lipidosis. *Nutrients*. 2013 Jul 19;5(7). P. 2811–2835.



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Показатель	После применения липотона	Без применения липотона
АЛАТ (Ед/л)	76,4	93,4
АСАТ (Ед/л)	65,3	75,8
ЩФ (Ед/л)	95,2	146,3
ГГТ (Ед/л)	5,4	15,2
Мочевина (ммоль/л)	13,6	16,3
Креатинин (мкмоль/л)	184,3	212,5
Холестерол (ммоль/л)	7,3	8,6
Альбумин (г/л)	26,7	24,3
Общий белок (г/л)	66,4	61,5

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-mail: *michael.yurievich@yandex.ru*

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(*Pinus banksiana* Lamb.) – 20-25

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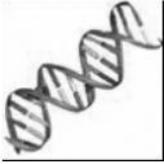
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**PRE-MIR165A**

E-mail: *v.chuykova2020@mail.ru*

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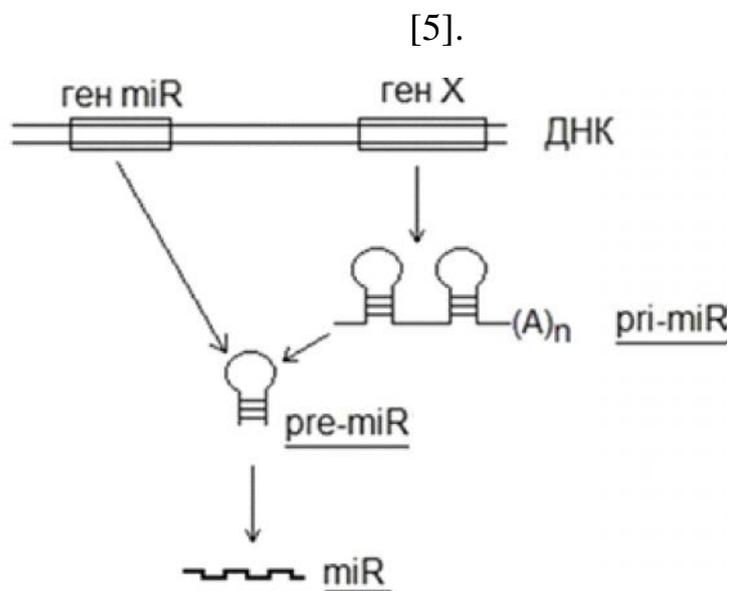
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GAGGGGAATGTTGTCTGGCT 3'; - 5'

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 CAGCTACTCTTCGCTTCCTACTCCTATCTCGATCCCTCTCTTCTTCTTCTT  
 CTTCTTCTTCCCTCCCCCTCCCTTGGATCGAGACCGAGCGGACGCAGAC  
 GAGTGGTGTAGATCTCGGACCAGGCA

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1. . . . / . . . . — . . . ., 1990. — 351 .
2. Iptekin B. A Comprehensive Prescription for Plant miRNA Identification / B. Iptekin, . . Akpınar, . Budak // *Front. Plant Sci.* — 2017. — Vol. 7.
3. Yanfei D. Emerging roles of microRNAs in the mediation of drought stress response in plants / D. Yanfei, . Yueliang, Z. Cheng // *Experimental Botany.* — 2013. — V.64. — P. 3077 — 3086.
4. Borges F. The expanding world of small RNAs in plants / F. Borges, R.A. Martienssen // *Nat. Rev. Mol. Cell Biol.* — 2015. — V. 16. — 727—741.
5. Aequorin-based luminescence imaging reveals stimulus- and tissue-specific Ca<sup>2+</sup> dynamics in Arabidopsis plants / X. Zhu [et al.] // *Mol. Plant.* — 2013. — V. 6. — P. 444—455.
6. Characterization and Function of MicroRNA's in Plants / W.W Liu [et al.] // *Front. Plant Sci.* — 2017. Vol. 8.
7. A microRNA array reveals extensive regulation of microRNAs during brain development / A.M. Krichevsky [et al.] // *RNA.* — 2003. — 9. — P. 1274—1281.
8. Deciphering the role of miRNA in reprogramming plant responses to drought stress / A. Singh [et al.] // *Crit. Rev. Biotechnol.* — 2023. — V. 43. — P. 613 — 627.



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(MANOVA).

(Wilks lambda=0,79663, F(6, 20)=0,85098, p=0,54642).

(Wilks lambda=0,36702, F(12, 40)=2,1688, p=0,03345)

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	шизоциты	эхиноциты	аканоциты	кодоциты	сфероциты
контроль	6,5±1,7	1,5±0,5	0,0±0,0	1,5±1,0	0,0±0,0
физ. Нагрузки	4,9±1,7	0,4±0,2	0,1±0,1	0,0±0,0	0,0±0,0
физ. Нагрузки + милдронат	7,7±2,8	3,0±2,0	0,0±0,0	0,2±0,1	8,0±5,4
физ. Нагрузки + L-карнитин	8,1±2,3	5,7±2,0	0,1±0,1	5,9±3,4	0,0±0,0

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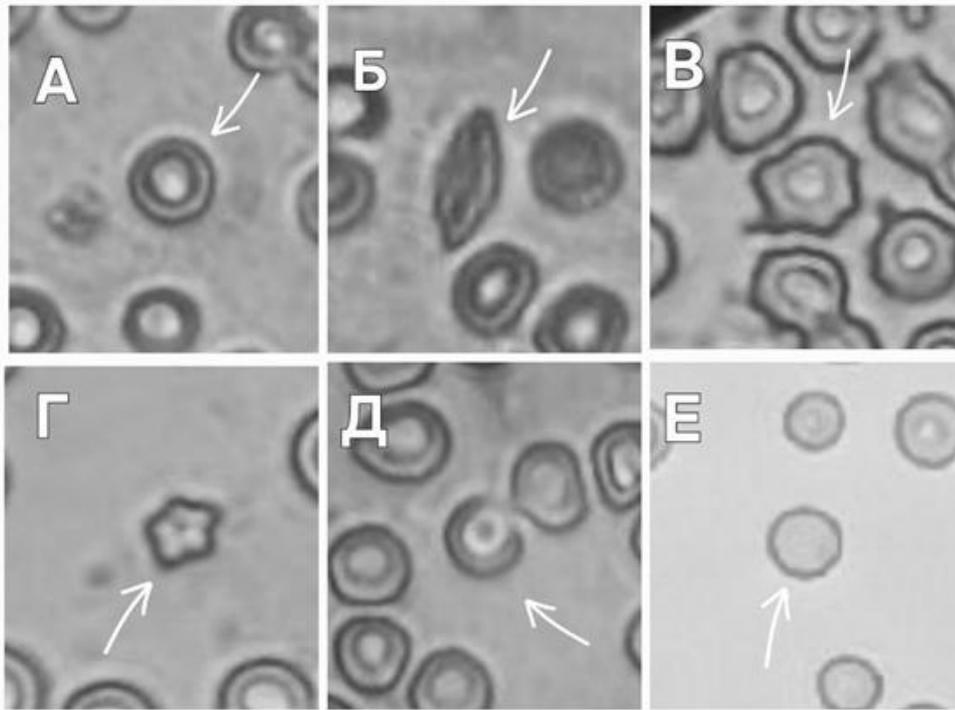
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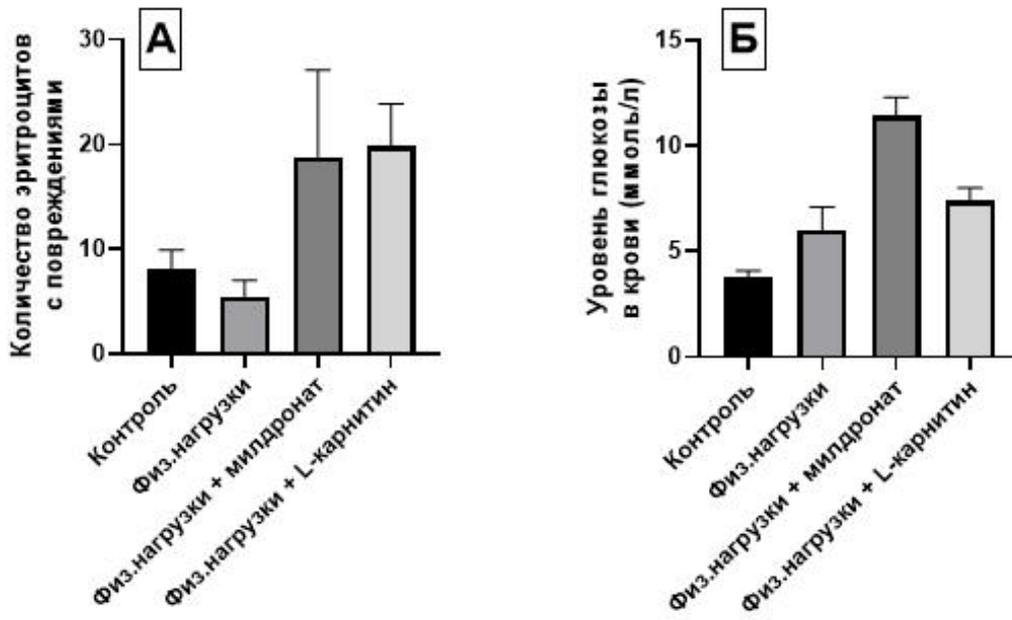
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4. : « », 2003 — 128 с. / . .
5. // . — 2012. — 1. — .3-12. / . . // . — 2022. — 5. — .128-133. / . . // . ., 2025, 27.



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E-mail: *rybolov@mail.ru*

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*Rattus norvegicus* L.,

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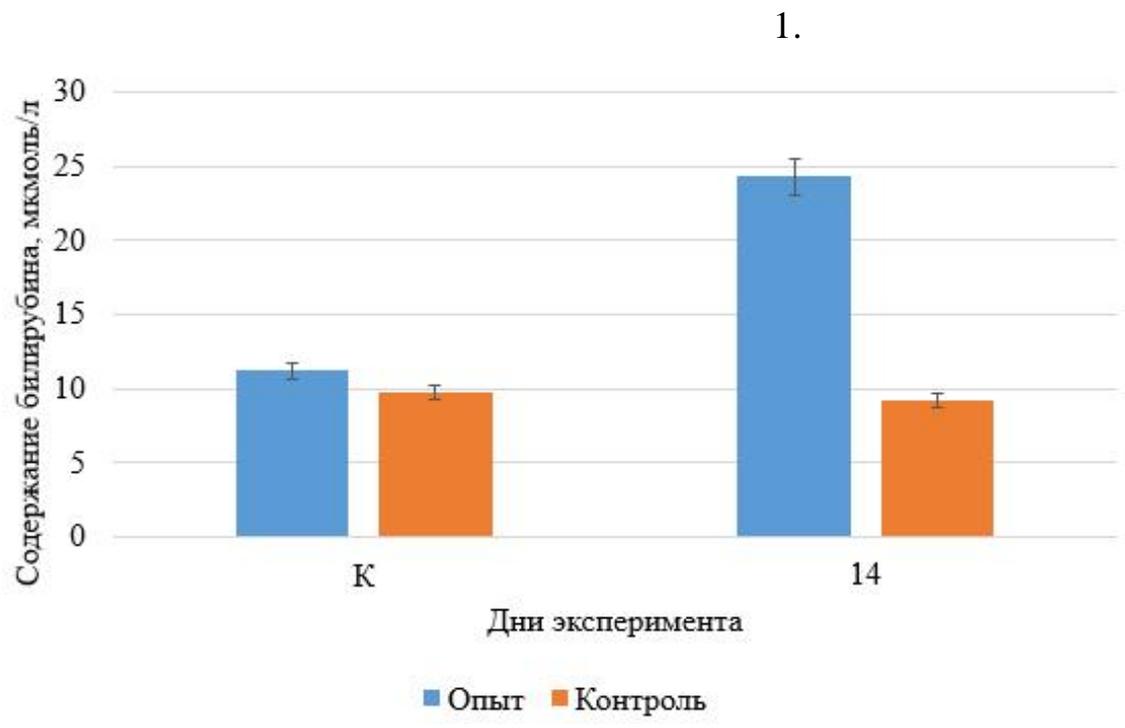
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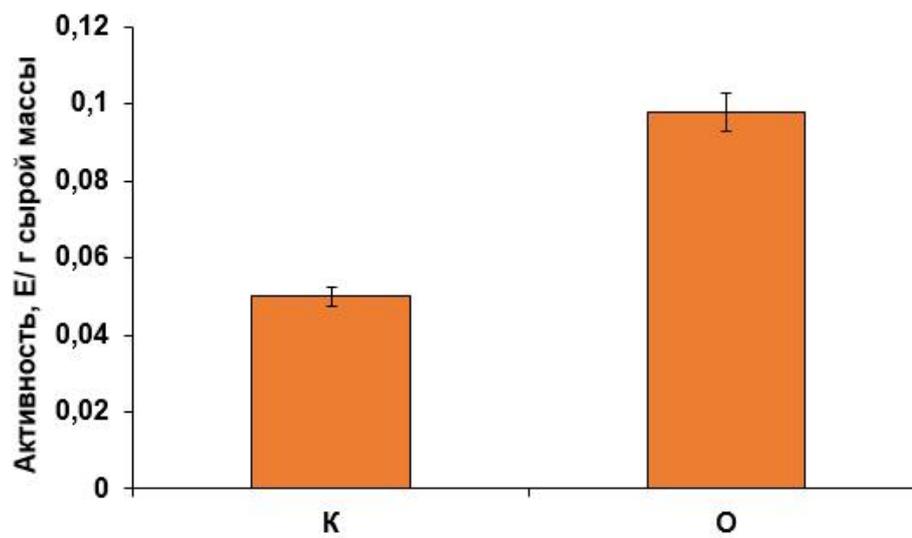
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1. Petersen M.C. Regulation of hepatic glucose metabolism in health and disease / M.C. Petersen, D.F. Vatner, G.I. Shulman // *Nat Rev Endocrinol.* – 2017. – V. 13. – P. 572–587.
2. Rui L. Energy metabolism in the liver / L. Rui // *Comprehensive Physiology.* – 2014. – V. 4, 1. – P. 177 - 197.
3. . . . / . . . , . . . // . – 2011. – . 3. – 3. – . 107-114.
4. -6- / . . . – 2007. – . 5. – 2. – . 19-20. //
5. / . . . , . . . , . . . // . – 2014. – 1. – . 52–53.
6. Metabolic oxidation of acetaminophen (paracetamol) mediated by cytochrome P-450 mixed-function oxidase and prostaglandin endoperoxide synthetase in rabbit kidney / J. Mohandas [et al.] // *Toxicol Appl Pharmacol.* – 1981. – V. 61. – P. 252–259.
7. Voloshchuk O. Activity of liver mitochondrial Krebs cycle NAD<sup>+</sup>-dependent dehydrogenases in rats with hepatitis induced by acetaminophen under conditions of alimentary protein deficiency / O. Voloshchuk., G. Kopylchuk // *Biochemistry (Moscow) Supplement Series B Biomedical.* - 2016. - V. 62. -P. 283-286.
8. Thomas D. Identification of the structural gene for glucose-6-phosphate dehydrogenase in yeast. Inactivation leads to a nutritional requirement for organic sulfur / D. Thomas, H. Cherest, Y. Surdin-Kerjan // *The EMBO Journal.* – 1991. – V. 10. – P. 547–553.



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**GTA-2** -

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-mail: *dowi2009@mail.ru*

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6–8%

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*GTA-2 (LOC103645944)*

NCBI (<https://www.ncbi.nlm.nih.gov/>).

Plant Transcriptional Regulatory Map (<http://plantregmap.gao-lab.org/>).

*GTA-2*

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***GTA2 (LOC103645944)***

ТФ	Семейство	Положение в п.н.	Направление	Таргетная последовательность	Функции
AC20765.6.3_FG002	ARF	480-500	+	TAAATATTTT TGAGAGAAAG	Фактор ответа на ауксин.
GRMZM2G164735	BBR- BPC	347-370	-	GTGAAATGAA AAGAGAAAGG GATG	Специфически связывается с GAGA-повторами, которые находятся в регуляторных последовательностях генов, участвующих в процессах развития.
GRMZM2G118690		886-906	+	CTCGCGCTCTT CCTCTCTTGC	
GRMZM2G118690		884-904	+	TTCTCGCGCTC TTCTCTCTT	
GRMZM2G118690		344-364	+	TCCCATCCCTT TCTCTTTCA	
GRMZM2G164735		345-368	-	GAAATGAAAA GAGAAAGGGA TGGG	

GRMZM2 G164735		889-912	-	GCGGAGGCAA GAGAGGAAGA GCGC	
GRMZM2 G164735		343-366	-	AATGAAAAGA GAAAGGGATG GGAC	
GRMZM2 G118690		346-366	+	CCATCCCTTTC TCTTTTCATT	Специфически связывается с GAGA-повторами, которые находятся в регуляторных последовательностях генов, участвующих в процессах развития.
GRMZM2 G118690		348-368	+	ATCCCTTTCTC TTTTCATTTT	
GRMZM2 G149040	bZIP	204-214	-	CCAAGCTGGC G	Вероятно, участвует в развитии сосудов и организации тканей побегов.
GRMZM2 G050939	C2H2	845-863	+	TCTCCCCCCTC CCCTCCCC	Незаменимый белок. Изоформа 1 является активатором транскрипции, связывает как 5S рДНК, так и 5S рРНК и стимулирует транскрипцию гена 5S рРНК, регулирует уровни 5S рРНК во время развития.
GRMZM2 G050939		894-912	+	CTTCCTCTCTT GCCTCCGC	
GRMZM2 G050939		225-243	+	ACTCATCAGCC ATCTCATC	
GRMZM2 G050939		943-961	+	CGCCGTCCGCC CCCGCCTC	
GRMZM2 G050939		897-915	+	CCTCTCTTGCC TCCGCCTG	
GRMZM2 G179677		312-328	-	TAGTAAAAGC CAAAGGT	Действует как положительный регулятор синтазы крахмала SS4. Контролирует развитие хлоропластов и образование гранул крахмала.
GRMZM2 G079653	ERF	949-963	-	CGGAGGCGGG GGCGG	Выступает как репрессор транскрипции. Связывается с GCC-боксом промотора патогенез-связанных генов. Участвует в регуляции экспрессии генов под действием стрессовых факторов и компонентов сигнальных путей передачи стрессового сигнала, а также может регулировать другие AtERF.
GRMZM2 G138396		322-336	+	TTTACTACGGC GCCA	Вероятно, действует как активатор транскрипции. Связывается с GCC-боксом промотора патогенез-связанных генов. Может участвовать в регуляции экспрессии генов под действием стрессовых факторов и компонентов стрессовых сигнальных путей.
GRMZM2 G144744	GRAS	347-366	-	AATGAAAAGA GAAAGGGATG	Выступает как репрессор сигнального пути гиббереллинов (GA). Действует, входя в состав мультибелковых комплексов, подавляющих транскрипцию GA-индуцируемых генов. При обработке GA разрушается протеасомой, что активирует сигнальный путь GA.
GRMZM2 G144744		309-328	-	TAGTAAAAGC CAAAGGTGAT	
GRMZM2 G144744		349-368	-	GAAATGAAAA GAGAAAGGGA	
GRMZM2 G144744		66-85	-	CAGTACGAAA GAAAAGGGGC	
GRMZM2 G118250	LBD	313-333	+	CCTTTGGCTTT TACTACGGCG	Способствует переходу от пролиферации к дифференциации в зародышевом мешке. Отрицательный регулятор пролиферации клеток на адиаксальной стороне листьев. Регулирует образование симметричной пластинки и установление жилкования.
GRMZM2 G118250		800-820	+	TCTTCTCCGCG AACCTCGCGT	
GRMZM2 G110153	MIKC MADS	352-365	-	ATGAAAAGAG AAAG	Участвует в развитии цветочных органов, необходим для нормального развития лодикул.
GRMZM2 G171365		885-905	+	TCTCGCGCTCT TCCTCTCTTG	Регулирует время цветения. Может контролировать удлинение междоузлий и ускорять переход к фазе цветения. Возможно, действует раньше регуляторов цветения MADS1, MADS14, MADS15 и MADS18 в пути индукции цветения.
GRMZM2 G171365		891-911	+	GCTCTTCTCT CTTGCCCTCCG	

GRMZM2 G095904	MYB	915-935	+	GCCTCCAGGCT CCAACAACCC	Выступает негативным регулятором холодоустойчивости. Подавляет транскрипцию генов бета-амилаз в ответ на холодовой стресс, взаимодействуя с TIFY11A/JAZ9. Мальтоза, продуцируемая бета-амилазами, защищает клеточные мембраны при холодовом стрессе у риса усиливая холодоустойчивость.
GRMZM2 G008374	NAC	141-158	+	TTAATTGAGA AACAAGAA	Связывается с промоторными областями генов, участвующих в процессах катаболизма хлорофилла, таких как NYC1, SGR1, SGR2 и PAO.
AC23386 5.1_FG00 3		142-158	+	TAATTGAGAA ACAAGAA	Контролирует энуклеацию сидовидных элементов и деградацию цитозоля. Не требуется для формирования литических вакуолей.
GRMZM2 G069047		141-157	+	TTAATTGAGA AACAAGA	Активатор транскрипции генов, участвующих в биосинтезе вторичных клеточных стенок. Вместе с NST2 и NST3 необходим для утолщения вторичных стенок (склеренхимных волокон), формирования вторичной ксилемы (трахеальных элементов) и развития эндотеция пыльника. Также может регулировать лигнификацию вторичных стенок в других тканях.
GRMZM2 G139700		138-156	-	CTTGTTTCTCA ATTAATTT	Участвует в регуляции формирования апикальной меристемы побега во время эмбриогенеза, разделении органов, сращении перегородок гинецея по длине завязей. Необходим для образования побегов в каллусе, развития края листа и формирования зубчатости, инициации пазушных меристем и их отделения от главного стебля. Регулирует филлотаксис на всех этапах развития растения. Вероятно, выступает как ингибитор клеточного деления.
GRMZM2 G058518		142-156	-	CTTGTTTCTCA ATTA	Регулирует развитие вторичных волокон клеточной стенки. Участвует в регуляции генов биосинтеза целлюлозы и гемицеллюлозы, а также генов, участвующих в полимеризации и сигнализации лигнина.
GRMZM2 G179885	TALE	140-156	-	CTTGTTTCTCA ATTAAT	Транскрипционный фактор семейства NAC, связанный с содержанием белка в зерне (GPC). Ускоряет старение листьев и усиливает ремобилизацию питательных веществ из листьев в развивающиеся зёрна.
GRMZM2 G017087		848-867	+	CCCCCTCCCC TCCCCGTGC	Транскрипционный фактор, связывающийся с РНК. Вероятно, регулирует гены, участвующие в развитии.
GRMZM2 G017087		858-877	+	CTCCCCGTGCC TCCCTCGGG	
GRMZM2 G017087		945-964	+	CCGTCCGCCCC CGCCTCCGG	
GRMZM2 G017087		856-875	+	CCCTCCCCGTG CCTCCCTCG	
GRMZM2 G017087		870-889	+	CCCTCGGGCCT CGTTTCTCG	
GRMZM2 G017087		846-865	+	CTCCCCCTCC CCTCCCCGT	
GRMZM2 G017087		886-905	+	CTCGCGTCTT CCTCTCTTG	
GRMZM2 G017087		844-863	+	GTCTCCCCCT CCCCTCCCC	
GRMZM2 G017087		854-873	+	TCCCCTCCCCG TGCTCCCT	
GRMZM2 G135447		303-310	+	GCTGTCAT	

GRMZM2 G047370	Trihelix	673-682	+	СТААССАСАС	Может действовать как молекулярный переключатель в ответ на световые сигналы.
GRMZM2 G047448	WOX	682-692	+	ССААТСААТС А	Репрессор транскрипции, необходимый для формирования и развития почек побега. Играет важную роль на ранних стадиях формирования пазушной меристемы. Предположительно, не участвует в поддержании апикальной меристемы побега.
GRMZM2 G069274		684-693	+	ААТСААТСАС	Фактор транскрипции, который может участвовать в процессах развития.
GRMZM2 G162481		139-149	+	ААТТААТТГА G	
GRMZM2 G162481		618-628	-	ТТТТААТТАГТ	
GRMZM2 G038252		682-689	+	ССААТСАА	
GRMZM2 G162481		49-59	-	ТГТТААТТГАА	
GRMZM2 G069274		680-689	+	САССААТСАА	
GRMZM2 G038252		50-57	+	ТСААТТАА	
GRMZM2 G038252		141-148	-	ТСААТТАА	
GRMZM2 G162481		617-627	+	ААСТААТТАА А	

1. Bouche N., Fromm H. GABA in plants: just a metabolite? // Trends in Plant Science. — 2004. — Vol. 9, No. 3. — P. 110–115.
2. Renault H., Roussel V., El Amrani A., et al. The Arabidopsis pop2-1 mutant reveals the involvement of GABA transaminase in salt stress tolerance. // BMC Plant Biology. — 2010. — Vol. 10. — Article 20.
3. Kinnersley A.M., Turano F.J. Gamma aminobutyric acid (GABA) and plant responses to stress. // Critical Reviews in Plant Sciences. — 2000. — Vol. 19, No. 6. — P. 479–509.
4. AkCay N., UCarli C., CiftCi Y. GABA Shunt and Its Role in Stress Tolerance in Plants // Plant Stress. — 2021. — Vol. 2, No. 1. — P. 100032.
5. Fait A., Fromm H., Walter D., Galili G., Fernie A.R. Highway or byway: the metabolic role of the GABA shunt in plants. // Trends in Plant Science. — 2008. — Vol. 13, No. 1. — P. 14–19.
6. Michaeli S., Fromm H. Closing the loop on the GABA shunt in plants: are GABA metabolism and signaling entwined? // Frontiers in Plant Science. — 2015. — Vol. 6. — Article 419.
7. Batushansky A., Kirma M., Grillich N., et al. Phosphorylation of GABA transaminase is critical for regulating the levels of GABA during hypoxia in plants. // Plant Cell. — 2015. — Vol. 27, No. 6. — P. 1532–1548.
8. Shelp B.J., Bown A.W., McLean M.D. Metabolism and functions of gamma-aminobutyric acid. // Trends in Plant Science. — 1999. — Vol. 4, No. 11. — P. 446–452.

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